



Age and diet-specific trace element accumulation patterns in different tissues of chub
(*Squalius cephalus*): juveniles are useful bioindicators of recent pollution

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Highlights

- Three different feeding groups of chub from the River Szamos were analyzed.
- Feeding groups were differed based on the trace element patterns of tissue.
- Generally, the highest concentrations of elements were found in the juveniles.
- Only the hepatic copper concentrations showed positive correlation with fish age.
- The juveniles of chub can be used as a bioindicators of the recent metal pollution.

1 **Abstract**

2

3 Chub (*Squalius cephalus* L. 1758) specimens of three age groups with different types of diet were collected
4 in November 2013 in the River Szamos/Someş, Hungary. The Ca, K, Mg, Na, Cd, Cr, Cu, Fe, Mn, Pb, Sr and Zn
5 concentrations were analyzed in the muscle, gills and liver samples of chub by microwave assisted plasma-
6 atomic emission spectrometry (MP-AES). The Kruskal–Wallis test revealed significant differences among
7 different age groups based on the trace element concentrations in the liver, muscle and gills. The trace element
8 concentration pattern in muscle and liver of different age groups differed, may be according to the different diet
9 types of the groups. Meanwhile no differentiation among the age groups based on the trace element
10 concentration in the gills was observed, probably because the pattern of trace elements in the gills is related to
11 the habitat preference, which does not differ during the life-span of chub. In contrast to expectations, trace
12 element concentrations in juveniles were the highest in most cases, certainly because of their specific diet,
13 relatively fast metabolic rate and inadequately developed detoxification system. Only the copper concentrations
14 in liver increased with fish age. Considerable concentrations of trace elements in the tissues of juveniles were
15 observed in the case of elements whose concentrations in the River Szamos were higher in 2013 than in previous
16 years. According to this phenomenon, trace element patterns in the tissues of juveniles may be good indicators of
17 recent pollution of watercourses.

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19 **Keywords:** Fish, Muscle, Liver, Gills, Heavy metals, MP-AES

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21

22 **Introduction**

23

24 Nowadays, the heavy metal pollution of aquatic ecosystems is a burning issue all over the world, mainly due
25 to these elements' toxicity, long persistence, bioaccumulation and biomagnification in the food web (Miracle and
26 Ankley, 2005; Carrasco et al., 2011, Carneiro et al., 2014). Two sources of heavy metals are in water
27 ecosystems: the natural geological background and anthropogenic activities, such as industrial and agricultural
28 emissions and atmospheric deposition (MacDonald et al., 2000; Lenhardt et al., 2009). Since trace elements can
29 be accumulated in living organisms in different concentrations and their bioavailability is not equal, systematic

30 monitoring of the concentrations of pollutants in organisms to predict environmental risks should be given
31 priority (Nakata et al., 2005; Zhou et al., 2008; Rašković et al., 2018).

32 Fish communities are widely used in monitoring impacts of aquatic contaminations (Hermenean et al., 2015;
33 Jia et al., 2016; Liu et al., 2016). Fish living in polluted waters can take up various amounts of trace elements by
34 several routes; i.e. through the skin or gills, or into the digestive system by feeding (Yilmaz et al., 2007;
35 Lenhardt et al., 2012). Metal accumulation in fish depends on both the chemical properties of the elements and
36 different factors such as ecological needs, and the physiological state, size and age of individuals, as well as their
37 life cycle, life history and feeding habits (Newman and Doubet, 1989; Canpolat and Çalta, 2003; Zhang and
38 Wong, 2007; Lenhardt et al., 2015).

39 The accumulated trace element patterns are generally different among the different tissues, primarily due to
40 the various physiological features (Subotić et al., 2013a, 2013b). Gills and liver, as metabolically active organs,
41 are target organs for trace element accumulation (Yilmaz et al., 2007; Lenhardt et al., 2012). Therefore gills and
42 liver are frequently used as target organs for metal accumulation assessments (Djikanović et al., 2016; Jia et al.,
43 2017). Generally, the trace element accumulation in muscle tissue is lower; but fish muscle is the main part of
44 the fish consumed by humans (Jia et al., 2017; Subotić et al., 2013a). For this reason, monitoring the trace
45 element pattern of fish muscle is equally important (Yancheva et al., 2015, Traina et al., 2019).

46 Chub [*Squalius cephalus* (Linnaeus, 1758) previously *Leuciscus cephalus*] is one of the most common and
47 widespread cyprinid species in Europe and is a very popular game fish (Kottelat and Freyhof, 2007). Chub is
48 most abundant in small rivers and large streams, but it also occurs in slow-flowing lowland rivers and very small
49 mountain streams (Kottelat and Freyhof, 2007). Adults are predominantly piscivorous predators of freshwater
50 ecosystems (Czeglédi and Erős, 2013). Chub is a generalist fish species; it also survives in more polluted water
51 (Machala et al., 2001; Dragun et al., 2012; 2016). Therefore, chub is increasingly used to assess the
52 contamination levels of water bodies and give relevant information regarding the endangered status
53 (vulnerability) of aquatic organisms (e.g. Dragun et al., 2007; Triebskorn et al., 2007; Yilmaz et al., 2007;
54 Krasnići et al., 2013).

55 In recent years, considerable research has focused on the concentrations of trace elements in adult fish (e.g.
56 Subotić et al., 2013a, 2013b; Hermenean et al., 2015; Djikanović et al., 2018). However, the accumulation and
57 distribution of trace elements in fish tissues are influenced by size, age and feeding behavior (Jia et al., 2017;
58 Ndimele et al., 2017). The diet of a fish species can alter considerably during its life-span; therefore the pattern
59 of accumulated trace elements can vary among the different age groups of the same species (Handy et al., 2003;

60 Dragun et al., 2009; Subotić et al., 2013b). During the ontogenesis of the chub, there is a rather pronounced diet
61 shift: there are three different types of diet (Balestrieri et al., 2006; Marković et al., 2007; Sasi and Ozay, 2017).
62 Chub juveniles (age 0+) have a plankto-phytophage diet; they feed mainly on diatoms, green algae, and
63 zooplankton (Marković et al., 2007). The older, sexually immature chub (age 1+, 2+) have a plankto-phyto-
64 zoophage diet, feeding on diatoms, green algae, and macrophytes, and also on animals, such as larvae and
65 imagines of Odonata, Trichoptera, Diptera and Ephemeroptera insects (Marković et al., 2007). The sexually
66 mature adult chub (age 3+, and thereafter) have a mainly carnivorous diet. They prey predominantly on fish, but
67 also feed on insects, molluscs, and amphibians (Ünver and Erk'akan, 2011). These significant shifts in diet can
68 lead to different bioaccumulation and biomagnification features in the different age groups of chub (Merciai et
69 al., 2014). Furthermore, several studies have highlighted that trace elements can be accumulated in the tissues of
70 juveniles in the highest concentrations as a result of various mechanisms: e.g. the specific diet, the higher
71 relative metabolic rate, and the inadequately developed detoxification system of young specimens (Merciai et al.,
72 2014; Jia et al., 2017; Ndimele et al., 2017). Therefore, the pollution indicator capability of different age groups
73 of the same species can be different (Jia et al., 2017).

74 The River Szamos (Romanian: Someş) is one of the most polluted rivers in Europe (Kraft et al., 2003; 2006).
75 In early 2000, both its water and a huge amount of sediment were contaminated with cyanide and heavy metals
76 entering the River Szamos (Lakatos et al., 2003). These events caused an ecological disaster in the River Szamos
77 and in its recipient watercourse, the River Tisza; moreover the sediment of these rivers were strongly enriched
78 by contaminants compared to the local geochemical background concentrations (Van der Veen et al., 2002; Fleit
79 and Lakatos, 2003; Kraft et al., 2003; Lakatos et al., 2003). Thereafter, trace element concentrations decreased in
80 both the Szamos and the Tisza (Óvári et al., 2004). However, recent pollution of the River Szamos with Pb, Cd,
81 Cu and Zn has occurred due to intensive mining activities in Romania (Simon et al., 2017).

82 In this paper, we analyzed concentrations of twelve elements in the muscle, gills and liver of different age
83 groups of chub living in the River Szamos. Different trace element concentrations in varying age groups of chub
84 were hypothesised, because of their different type of diet and other physiological features. It was also
85 hypothesised that trace element concentration would be higher in older groups than in younger ones, due to the
86 bioaccumulation and biomagnification of these elements. Diverse patterns of accumulated trace elements in the
87 chub groups which differed by age, size, and diet were analyzed to test these hypotheses; at the same time, the
88 potential risks of contaminated fish consumption on human health were evaluated. Furthermore, this study aims

89 to understand which age group of chub performs better as a bioindicator of the trace element contamination of
90 freshwater ecosystems.

91

92

93 **Material and methods**

94

95 **Study area and sample collection**

96 The River Szamos is the second largest tributary of the River Tisza. Its catchment basin collects waters over
97 a 15 882 km² area of Romania and Hungary. Its total length is 415 km, but the Hungarian section is only 50 km
98 long. Fish were caught at the village of Csenger, near the Hungarian-Romanian border, where the water depth
99 varies between a few centimeters to 2-3 meters, and the main sediment types are pebbles and sand. The
100 coordinates of the sampling site are 47°50'17.89"N, 22°41'37.48"E (Fig. 1).

101 A total of 33 chub specimens were collected in November 2013 by electrofishing (Hans Grassl IG200/2b,
102 PDC, 75–100 Hz, 350–650 V, max. 10 kW, Hans Grassl GmbH, Germany). All chub specimens were captured
103 alive and killed immediately by spinal severance. Measurements of the standard length (SL) and body weight
104 (W) of each specimen were made to the nearest 0.1 mm and 0.01 g, respectively. Samples were stored at -18 °C
105 until sample processing. We confirm that all procedures were performed in compliance with the relevant laws
106 and institutional guidelines, and that the appropriate institutional committees have approved them (permission
107 number: HBH/01/00971-2/2013).

108 Scales were used to determine the age of each specimen. Scale samples were taken below the dorsal fin, from
109 the area above the lateral line (Tesch, 1968). Preparations of scales were assessed using a binocular microscope
110 (10× magnification). Age assessment was applied in accordance with Tesch (1968). Age estimations were
111 validated by a second independent assessment.

112

113 **The feeding groups**

114 Several studies showed that there are three different types of diet of chub, depending on their age (Balestrieri
115 et al., 2006; Marković et al., 2007; Caffrey et al., 2008). Therefore, three different feeding groups of chub were
116 created. Each specimen was assigned to one of the three different feeding groups, according to age and size:

117 First feeding group: the juveniles (age 0+; N = 8);

118 Second feeding group: the sexually immature individuals (age 1+ and 2+; N = 16);

119 Third feeding group: the sexually matured adults (age 3+ onwards; N = 9).

120

121 **Sample processing and element analysis**

122 Muscle, gills and liver samples were removed with plastic tools in order to avoid any metal contamination of
123 the samples. Gills were sampled by removing the whole branchial apparatus and collecting the second gill arch
124 from the left side. The dissected samples were then rinsed with double deionised water (Milli-Q) and were
125 weighed into glass beakers using an analytical balance (Precisa 240A). They were dried overnight at 105 °C until
126 reaching constant weight and reweighed to determine their dry mass. Samples were digested on an electric hot
127 plate using 4.0 mL 65% (m/m) nitric acid (reagent grade, Merck) and 1.0 mL 30% (m/m) hydrogen-peroxide
128 (reagent grade, Merck) in the same container at 80 °C for 4 h. Digested samples were diluted with 1 % (m/m)
129 nitric acid (reagent grade, Merck and Milli-Q water) up to a final volume of 10 mL (Braun et al., 2009; 2012).
130 The concentrations of Ca, K, Mg, Na, Cd, Cr, Cu, Fe, Mn, Pb, Sr and Zn were measured by a microwave
131 plasma-atomic emission spectrometer (MP-AES 4100, Agilent Technologies) system. The values of the limits of
132 detection (LOD) in $\mu\text{g l}^{-1}$ were: Ca, 0.070; K, 0.270; Mg, 0.080; Na, 0.484; Cd, 1.863; Cr, 0.489; Cu, 0.414; Fe,
133 0.856; Mn 0.057; Pb, 1.235; Sr, 0.051; and Zn, 3.125, respectively. The limit of detection of each sample was
134 verified based on the instrumental detection limit, sample mass and volume to which it had been diluted. The
135 average detection limits for each of the assessed elements were (mg kg^{-1} wet weight): Ca, <0.001; K, 0.002; Mg,
136 0.001; Na, 0.004; Cd, 0.009; Cr, 0.004; Cu, 0.003; Fe, 0.006; Mn <0.001; Pb, 0.009; Sr, <0.001; and Zn, 0.023,
137 respectively. The following wavelength lines of the MP-AES analysis were used: Ca 422.673 nm, K 766.491
138 nm, Mg 285.213 nm, Na 588.995 nm, Cd 228.802 nm, Cr 425.433 nm, Cu 324.754 nm, Fe 371.993 nm, Mn
139 403.076 nm, Pb 405.781 nm, Sr 407.771 nm, and Zn 481.053 nm.

140 An auto sampler (Agilent SPS3), a Meinhard type nebulizer and a double pass spray chamber were used. We
141 applied a five-point calibration procedure prepared from the multi-element standard solution (Merck ICP multi-
142 element standard solution IV). Certified reference material was used (ERM-BB422, fish muscle) during the
143 measurement. The recoveries were within the 10% of the certified values for the metals. The wavelengths and
144 measuring parameters were chosen on the basis of suggestions provided by the instrument's software (MP
145 Expert). Concentrations of all elements were expressed as mg kg^{-1} wet weight (ww).

146

147 **Bioconcentration factor and metal pollution index**

148 The bioconcentration factor (BCF) of a chemical compound is defined as the ratio between the concentration
149 of that chemical in an organism (or in a certain tissue of the organism) and the concentration of the chemical in
150 the aqueous environment (Ivanciuc et al., 2006):

$$151 \quad \text{BCF} = C_{\text{fish}}/C_{\text{water}},$$

152 where C_{fish} means the concentration of the chemical in the whole body or tissue of the organism, expressed as
153 mg kg^{-1} wet weight, and C_{water} is the concentration of the chemical in the aqueous environment in which the
154 respective organism lives, expressed as mg l^{-1} . Data relating to concentrations of the trace elements of water
155 were obtained from the data base of the National Environmental Information System of Hungary (OKIR in
156 Hungarian). These were compared with criterion chronic concentrations (CCCs) for freshwater of the National
157 Recommended Water Quality Criteria (USEPA, 2017).

158 Metal pollution index (MPI) was assessed to compare the total content of trace elements excluding the
159 macroelements (Cd, Cr, Cu, Fe, Mn, Pb, Sr, Zn) for the different tissues of feeding groups of chub. The MPI
160 formula is the following (Usero et al., 1997; Ju et al., 2017):

$$161 \quad \text{MPI} = (C_1 \times C_2 \times C_3 \times \dots C_n)^{1/n},$$

162 where C_n is the mean concentration of trace element n in the analyzed tissue (mg kg^{-1} wet weight).

163

164 **Statistical analysis**

165 IBM SPSS Statistics for Windows (Version 20.0) (IBM, 2011) and Past 3.03 (Hammer et al., 2001) software
166 packages were used for statistical analysis. The normal distribution was tested with the Shapiro-Wilk test. The
167 homogeneity of variances was tested with the Levene's test. Since the variables lacked a normality of
168 distribution (Shapiro-Wilk test, $p < 0.05$), the non-parametric Kruskal–Wallis test was used to display the
169 differences among the concentrations and BCF values of elements in three different tissues (muscle, gills, and
170 liver) of three feeding groups of chub. LSD Multiple Comparison test as a post hoc test was used to explore the
171 significant differences between the groups. Spearman's non-parametric correlation test was used to check for
172 significant relationships between trace element concentrations and feeding groups of chub. Principal component
173 analysis (PCA) was used to assess the differentiation of feeding groups among the analyzed fish tissues, based
174 on the concentration of trace elements (Cd, Cr, Cu, Fe, Mn, Pb, Sr, Zn), except for the macroelements.

175 To evaluate the health risks of chub for human consumption, trace element concentrations of muscle (fillet)
176 were compared with the maximum acceptable concentrations (MACs) set by the European Union (EU, 2008)
177 and the Food and Agriculture Organization of the United Nations (FAO, 1983).

178

179

180 **Results**

181

182 **Age analysis**

183 Age analysis indicated that fish belonged to age classes from 0+ to 4+. The average standard length and
184 weight of the analyzed chub specimens per feeding group are shown in Table 1. The Kruskal–Wallis test
185 revealed significant differences (standard length: $H = 27.27, p < 0.001$; weight: $H = 27.27, p < 0.001$) in mean
186 standard length and mean weight among the three feeding groups.

187

188 **Characterization of the study area**

189 The descriptive statistics of the concentrations of elements in the water, obtained from the the data base of
190 the National Environmental Information System of Hungary (OKIR in Hungarian) are presented in Table 2. The
191 mean concentrations of Cu, Pb and Zn were above the criterion chronic concentrations (CCCs) for freshwater of
192 National Recommended Water Quality Criteria prescribed by the USEPA (2017) (Table 2).

193

194 **Macroelements**

195 Mean values of the concentrations of macroelements in muscle, gills and liver are presented in Table 3. The
196 highest concentrations of Ca, Mg and Na were observed in gills, while the highest concentrations of K were
197 measured in muscle.

198 The Kruskal–Wallis test revealed significant differences among the feeding groups with regard to element
199 concentrations in muscle for Ca, K and Mg ($p < 0.05$), in gills for Ca, K, Mg and Na ($p < 0.05$), and in liver for
200 K, Mg and Na ($p < 0.05$). The LSD Multiple Comparison test revealed that the concentrations of these elements
201 were significantly lower ($p < 0.05$) in the case of juveniles than in older groups (Table 3).

202 There was a significant ($p < 0.05$) increase in concentrations of certain macroelements with fish age (Table
203 4). Both K and Mg concentrations in all examined tissues, and Na concentrations in gills and liver, were
204 positively correlated with fish age (Table 4).

205

206 **Essential and non-essential trace elements**

207 Mean values of the concentrations of the trace elements in muscle, gills and liver are shown in Table 3. The
208 concentration of Cd in the muscle of the oldest group (age 3+ \geq) was below the detection limit. The highest
209 concentrations of Cd, Cr, Cu, Fe and Pb were detected in liver, while Mn, Sr and Zn reached the highest
210 concentrations in gills. The lowest concentrations of most of the studied trace elements were observed in the
211 muscle tissue.

212 The Kruskal–Wallis test revealed significant differences among the feeding groups with regard to trace
213 element concentrations in muscle for Cr, Cu, Fe, Mn, Pb, Sr and Zn ($p < 0.05$), in gills for Cr and Pb ($p < 0.05$),
214 and in liver for Cr, Cu, Pb and Sr ($p < 0.05$).

215 The LSD Multiple Comparison test indicated that concentrations of Cr, Fe, Pb, Sr and Zn in muscle were
216 significantly higher in the cases of juveniles than in older groups ($p < 0.05$) (Table 3). The highest concentrations
217 of Pb in gills and liver were also observed in fish from the youngest age class (0+) (LSD Multiple Comparison
218 test, $p < 0.05$) (Table 3). The Cu concentration measured in the liver was significantly higher in the oldest group
219 (3+ \geq) than in the younger groups (LSD Multiple Comparison test, $p < 0.05$).

220 In the case of trace elements, only the concentrations of Cu in the liver showed a significant increase with
221 fish age ($p < 0.05$) (Table 4). On the contrary, opposite trend was observed for some trace elements, which had a
222 significant trend, showing decreasing concentrations with age ($p < 0.05$). We experienced a significant ($p < 0.05$)
223 decrease in concentrations of Pb with fish age, in all studied tissues (Table 4). In addition, Cr, Fe, Mn and Zn
224 concentrations in muscle were negatively correlated with fish age (Table 4).

225 The PCA showed separations among the different feeding groups of chub based on trace element
226 concentrations in the muscle (Fig. 2) and the liver (Fig. 3). In the case of muscle, the first component (PCA 1)
227 contributed 88.03% of the total variance, while the second (PCA 2) contributed 7.34% of the total variance.
228 Based on the trace element pattern, juveniles were completely separated from the other groups (Fig. 2). The PCA
229 showed less separation between the 1+ year-old, 2+ year-old and older chub (age 3+ onward) (Fig. 2).

230 According to trace element concentrations in liver, the PCA showed less differentiation among the different
231 feeding groups (Fig. 3); the first component (PCA 1) contributed 36.56% of the total variance, while the second
232 (PCA 2) contributed 31.19% of the total variance. Juveniles were clearly separated from the other groups,
233 especially from the older chub (age 3+ onward) (Fig. 3). The 1+ and 2+ year-old chub showed notable overlaps
234 with other groups (Fig. 3). There were no notable separations among the feeding groups based on the trace
235 element concentration in the gills, according to the PCA.

236

237 **Bioconcentration factor and metal pollution index**

238 The bioconcentration factor values are given in Table 5. The highest BCF values for Cd, Cr, Cu, Fe and Pb
239 were observed in the cases of liver tissues. The highest BCF values for Mn and Zn were recorded in gills. The
240 BCF values also showed that for most of the trace elements accumulation was more higher in the tissues of
241 juveniles.

242 The highest MPI values were observed in juveniles in all examined tissues (Fig. 4).

243

244 **Human health implications**

245 MACs set by the European Union (EU) for Cd and Pb in fish meat are 0.05 and 0.30 mg kg⁻¹ ww,
246 respectively. MACs prescribed by the FAO (1983) for Cr, Cu, Fe, Mn and Zn in fish meat are 1.0, 30.0, 43.0, 1.0
247 and 40.0 mg kg⁻¹ ww, respectively.

248 Both the concentrations of Cd in juveniles (0+) and the 1+, 2+ age groups, and the concentrations of Pb in
249 juveniles (0+) were over the limit permitted by the EU (EU 2008) in the muscle samples. The concentrations of
250 the other analyzed elements were below the MAC levels prescribed by the EU (2008) and the FAO (FAO, 1983)
251 in all of the muscle samples.

252

253

254 **4. Discussion**

255

256 Both our results and earlier studies have demonstrated that metals deriving from industrial, mining and
257 agricultural activities have serious impacts in the River Szamos. Woelfl et al. (2006), Málnás et al. (2014) and
258 Simon et al. (2017) reported that the contamination level of Szamos is moderately with trace elements, especially
259 with Cu, Cr, Mn, Pb, Sr, and Zn than adjacent rivers. At the same time, our present study and the water
260 chemistry data of the National Environmental Information System of Hungary have also revealed continuous
261 pollution of the River Szamos. Moreover, Cu, Pb and Zn concentrations in the water were above the criteria for
262 chronic concentrations set by the USEPA (2017), indicating a considerable degree of contamination with these
263 elements.

264 The distribution of macroelements (Ca, K, Mg, Na) were found according to their functions in the organism.

265 Generally, the lowest concentrations of macroelements were in the tissues of juveniles, because of various

266 physiological features (Venugopal and Shahidi, 1996; Uysal et al., 2008). For example, in the case of Ca and Mg

267 - elements which accumulate mainly in the bony structures of gills (e.g. arches and rakers) (Mayer-Gostan et al.,
268 1983; Playle, 1998) - lower concentrations of these elements were observed in the gills of juveniles. This is
269 related to ontogenetic development features, namely that the bones of juveniles were not fully developed (Witten
270 et al., 2001). Similar results were found in a study based on laboratory experiments (Harangi et al., 2016).

271 Seventeen trace elements are required by some terrestrial and aquatic organisms (Yılmaz et al., 2017), such
272 as Cu, Cr, Fe, Mn, and Zn. For organisms trace amounts of these elements are essential for normal growth and
273 development. However, at higher levels of exposure and adsorption all trace elements can potentially be harmful
274 for most aquatic organisms (Yılmaz et al., 2017). Contrary to our hypothesis, only the Cu concentrations in the
275 liver showed an increase with fish age. Higher Cu concentrations in the liver of older chub (age 3+ onward) may
276 be the result of their predominantly piscivorous feeding (Djedjibegovic et al., 2012). Milošković et al. (2016)
277 also revealed a higher concentration of Cu in the liver of older chub, which is probably caused by the
278 biomagnification features of Cu. In the muscle of chub Cr, Fe, Mn and Zn were negatively correlated with fish
279 age, and differed significantly among feeding groups. Earlier studies demonstrated a considerable degree of
280 continuous pollution of these elements along the River Szamos (e.g. Óvári et al., 2004; Woelfl et al., 2006,
281 Málnás et al., 2014, Simon et al., 2017). According to the water chemistry data of the National Environmental
282 Information System of Hungary, there are recent contaminations involving Cr, Fe and Zn, because
283 concentrations of these elements were about 2–10 times higher than they had been in previous years. Similarly to
284 our results, negative correlations between Mn and Zn concentrations in muscle and fish size/age were also found
285 in the study by Djikanović et al. (2016), in the case of nase (*Chondrostoma nasus*) from the Medjuvršje
286 Reservoir (Republic of Serbia). The Medjuvršje Reservoir had been contaminated by Mn and Zn in the year of
287 the study (Djikanović et al., 2016), which implies a potential bioindicator capacity of juvenile fish in monitoring
288 recent environmental trace element pollution events.

289 Strontium (Sr) is a rarely investigated element in fish. The beneficial effects of Sr are questionable, but it can
290 also be bioaccumulated in the body, and can replace Ca in the organism. Woelfl et al. (2006), Málnás et al.
291 (2014) and Simon et al. (2017) also reported that the River Szamos is contaminated with a high amount of Sr
292 which may be originated mainly from agricultural activities. The highest concentrations of Sr were measured in
293 the gills of chub. Farrel and Campana (1996) found that water was a more important source of bioaccumulation
294 for Sr than consumed food. Dragun et al. (2016) observed that the Sr concentrations in the soluble gill fractions
295 of chub have not reflected the exposure level in water, because Sr mainly accumulated in the bony structures of

296 gills (e.g. arches and rakers) by Ca replacement, rather than in soft tissues (e.g. soluble gill fractions)
297 (Hermenean et al., 2017).

298 Neither vital nor beneficial effects caused by non-essential trace elements on organisms have been
299 established yet; these elements (e.g. Cd and Pb) are toxic to all organisms at low concentrations (Eisler, 1985). In
300 all examined tissues of chub, Pb was negatively correlated with fish age, and its concentration in juveniles was
301 significantly higher than that of older groups. Similar results were found in different fish species by several
302 authors (e.g. Demirak et al., 2006; Merciai et al., 2014; Ndimele et al., 2017). The occurrence of higher
303 concentrations of Pb in juveniles could be assigned to both high metabolic rate and inadequately developed toxic
304 metal neutralization mechanisms in younger fish (Kljaković Gašpić et al., 2002; Jia et al., 2017). The higher Pb
305 concentration in juveniles might be also caused by a recent pollution event. The Pb concentration of the water of
306 the Szamos was approximately 14 times higher in 2013 than that in previous years, based on data from the
307 National Environmental Information System of Hungary. This recent pollution incident affected all fish, but that
308 effect on juveniles was the most pronounced due to the phenomena mentioned above. According to this
309 phenomenon, the microchemistry pattern of the tissues of juveniles can be used as a bioindicator of recent
310 pollution events, not only for essential, but also for non-essential trace elements.

311 Earlier studies and our results also showed that muscle is the tissue with the lowest concentration of the
312 majority of trace elements (Djikanović et al., 2016; Jia et al., 2017), respectively. According this phenomenon,
313 fish muscle is not sufficient to indicate trace elemental contamination in the whole fish body, due to its lower
314 accumulation potential (Jia et al., 2017). In spite of this, in recent years, a considerable number of studies on
315 trace element contamination in fish muscle have been published, since fish muscle is the main part of the fish
316 consumed by humans (Jia et al., 2017; Subotić et al., 2013a, 2013b; Yancheva et al., 2015). Furthermore, this
317 present study demonstrated that the trace element pattern of muscle is the most variable among the different age
318 groups of fish, as a results of several mechanisms (e.g. specific diet). Therefore, monitoring the trace element
319 pattern of fish muscle is equally important.

320 Although the accumulation of trace elements in muscle tissue is also relevant, the main target tissue of the
321 bioaccumulation of trace elements is the main detoxifying organ, the liver (Dragun et al., 2012; Lenhardt et al.,
322 2012). Once the trace elements cross biological barriers and enter into the bloodstream, they will reach the liver
323 and will accumulate there (Yancheva et al., 2015); therefore, fish liver is the most important bioindicator organ
324 of water contamination (Jovanović et al., 2011). Concentrations and BCF values of the majority of examined
325 trace elements (Cd, Cr, Cu, Fe and Pb) were the highest in the liver, which implied that an examination of the

326 liver is also important. In spite of this, trace element concentrations in the liver did not differ among the feeding
327 groups of chub, as much as they did in the case of muscle.

328 A few differences in the trace element concentrations were observed in gills among the feeding groups, as the
329 pattern of trace elements in gills is accounted for by the habitat preference (Subotić et al., 2013a). Although the
330 diet of chub alters during its life-span, the habitat preference is permanent, because chub is a pelagic fish species
331 of riverine habitats (Kottelat and Freyhof, 2007). The highest concentrations and BCF values of Mn, Sr and Zn
332 were observed in gill samples because these elements are mainly taken up from water via gills, rather than by
333 feeding (Jia et al., 2017; Subotić et al., 2013a, 2013b). This phenomenon indicated that an examination of gills is
334 also important in monitoring accumulated trace elements in fish.

335 We hypothesized that trace element concentrations in different feeding groups of chub may be different. This
336 hypothesis was confirmed: the trace element concentration pattern of different feeding groups of chub differed,
337 especially in the cases of muscle and liver. We also hypothesized that the trace element concentrations would be
338 higher in older groups than in younger ones. However, this hypothesis was not confirmed by the results:
339 concentrations of most trace elements were higher in juveniles, and the concentrations of certain elements also
340 showed negative correlations with fish age, especially in the case of muscle. Higher trace element concentrations
341 in juveniles, and therefore a significant negative correlation between fish size and accumulated element
342 concentrations, have also been reported by several authors (Merciai et al., 2014; Jia et al., 2017; Ndimele et al.,
343 2017). There are several mechanisms which could cause this phenomenon. Chub is a relatively large fish species,
344 and its growth is relatively fast in the first years (Vlach et al., 2005), therefore tissue growth could be more rapid
345 than trace element intake (Ndimele et al., 2017). On the one hand, several authors hypothesized that the lipid
346 content of tissues has a relative dilution effect (Braune et al., 1999; Farkas et al., 2003), therefore the lower fat
347 content of juveniles tissue could cause higher trace element concentrations in their body than is the case with
348 adults (Merciai et al., 2014). On the other hand, lipid as a percentage of body weight reaches its peak at the end
349 of the main feeding period – autumn - and so in November this relative dilution effect in adults can also explain
350 this phenomenon (Farkas et al., 2003). Furthermore, the metabolic rate of fish is size-specific and the relative
351 metabolic rate of juveniles is higher (Newman and Doubet, 1989), therefore both the food intake and the relative
352 quantity of respiratory water passing through the gills (as the main sources of trace elements) are higher in
353 juveniles. Higher trace element concentrations in juveniles may also occur as a result of the inadequately
354 developed detoxification mechanisms in young fish (Ndimele et al., 2017). The specific diets of different feeding
355 groups of chub are also important. Juveniles feed mainly – and continuously - on phyto- and zooplankton, and

356 according to published data, high amounts of trace elements can accumulate in green algae and diatoms (Bácsi et
357 al., 2015; González-Dávila et al., 2000; Novák et al., 2014). According to data from the National Environmental
358 Information System of Hungary, the trace element pollution in the River Szamos was higher in 2013 than in the
359 previous years, especially in the case of Cr, Fe, Pb and Zn. Furthermore, the relative trace element exposure of
360 juveniles was higher than that of adults due mainly to higher metabolic rate. In addition, other factors could also
361 have influenced this phenomenon, e.g. changes in body surface-volume ratio during growth, or even faster short-
362 term uptake by juveniles (Merciai et al., 2014; Jia et al., 2017). Finally, many of these factors are likely to
363 interact (Merciai et al., 2014). According to the phenomena mentioned above, the bioaccumulation patterns of
364 trace elements in juveniles can be utilized as an effective bioindicator of recent environmental trace element
365 contamination (Jia et al., 2017). A recent pollution event can cause a relatively higher exposure over the entire
366 life-span of juveniles, than is the case in older specimens (Merciai et al., 2014). Moreover, juvenile chub are
367 primary/secondary consumers in the aquatic food web, because they feed mainly on phytoplankton and
368 zooplankton, therefore trace elements can be accumulated earlier in their organs than is the case in piscivorous,
369 tertiary consuming adults (Croteau et al., 2005).

370 In the case of muscle, the mean concentrations of Cd and Pb in juveniles were over the prescribed MACs.
371 The trace element concentrations in the muscle of larger specimens did not exceed MACs, and the minimum size
372 limit of chub in Hungary is 25 cm standard length, so the consumption of fillets of older specimens does not
373 seem to be harmful. Nevertheless, higher concentrations of Cd and Pb in the muscle of younger (age 0+, 1+, and
374 2+) specimens require further examinations. It can be dangerous to both the ecosystem (e.g. due to
375 biomagnifications) and to human health (e.g. due to poaching), thus the regular consumption of chub from the
376 River Szamos can be potentially hazardous for human health. Trace element concentrations in the gills and liver
377 exceeded the prescribed MACs several fold in the cases of all feeding groups; consequently, consumption of the
378 inner organs (e.g. gill and liver) of chub from the River Szamos is not recommended, according to prescribed
379 MACs.

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382

383 **Conclusions**

384

385 In conclusion, the results of the present study indicate a significant differentiation among the feeding groups
386 of chub based on metal accumulation patterns. The differences observed could be explained by the specific
387 physiological characteristics and diets of different age classes of chub.

388 Contrary to our hypothesis, the highest trace element concentrations were measured in juveniles. The reasons
389 for this phenomenon could be found in various mechanisms: e.g. the specific diet, higher relative metabolic rate,
390 and inadequately developed detoxification system of young specimens. Consequently, the prescribed MACs
391 were exceeded mainly in juveniles. Our results also show that the trace element pattern of juveniles was a good
392 indicator of the recent pollution events of the river in question, because all environmental factors (such as trace
393 element contaminations) which had an effect on the young of the year chubs over a season, were integrated in
394 their different tissues. Based on the investigation of trace element pattern of juvenile chubs in 2013, we detected
395 recent contaminations in the cases of several elements, which are also confirmed by water chemistry data from
396 the National Environmental Information System of Hungary.

397 Trace element concentrations in fillets of adult chub, which can be considered for consumption by humans,
398 did not exceed the prescribed MACs, but inner organs (e.g. gills and liver) contained higher concentrations of
399 trace elements than the prescribed MACs in several cases. Therefore, consumption of the inner organs of chub
400 living in the River Szamos is not recommended.

401 Chub is one of the most abundant fish species in relatively fast flowing sub-mountain, highland and lowland
402 streams and rivers; it has an important role in the food web and angling of these habitats, and furthermore, it is
403 widespread in the rivers of Europe. In view of these facts, and following our results, the juveniles of chub can be
404 used as a widespread, effective bioindicators in monitoring the recent trace element pollution of aquatic
405 ecosystems. However, further studies are needed to identify such patterns in other habitats.

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681

Table 1

Number of sampled individuals (n) of chub (*Squalius cephalus*) caught in the River Szamos/Someş, and their minimum, maximum, mean and standard deviation of standard length (SL, mm) and weight (W, g) per feeding group.

Feeding group	n	SL ± SD (mm)	Min.	Max.	W ± SD (g)	Min.	Max.
Age 0+	8	65.8 ± 8.2	54.2	75.5	5.11 ± 1.73	2.49	6.97
Age 1+, 2+	16	119.2 ± 18.0	93.6	140.5	34.09 ± 15.23	15.64	53.19
Age 3+≥	9	171.9 ± 17.3	152.0	212.3	99.93 ± 30.50	65.54	169.55

Table 2

Descriptive statistical data of total trace element concentrations in water from the River Szamos/Someş obtained from the the data base of the National Environmental Information System of Hungary (OKIR in Hungarian).

Element	Trace element concentration ($\mu\text{g l}^{-1}$)			Threshold values ($\mu\text{g l}^{-1}$) ^a
	Min	Max	Mean ± SD	
Cd	0.2	1.4	0.58 ± 0.44	0.72
Cr	1	12	3.18 ± 3.68	74
Cu	5	20	9.47 ± 5.72*	3.1
Fe	45	658	365.78 ± 226.59	1000
Mn	1.8	388	145.98 ± 110.72	–
Pb	3	20	6.92 ± 6.25*	2.5
Zn	1	270	136.22 ± 93.55*	120

^a Criterion chronic concentrations (CCCs) for freshwater of National Recommended Water Quality Criteria (USEPA 2017)

* Mean concentration of trace element in the water was higher than the threshold value.

Table 3

Concentrations of trace elements in the muscle, gills, and livers (in mg kg⁻¹ wet weight) of different feeding groups of chub in the River Szamos/Someş (mean value ± standard deviation).

Tissue	Element	Age 0+	Age 1+, 2+	Age 3+ _≥	
Muscle	Ca	150.70 ± 156.86 ^a	307.07 ± 89.64 ^b	263.30 ± 114.23 ^b	
	K	1063.41 ± 1130.07 ^a	2745.29 ± 154.38 ^b	3070.33 ± 227.90 ^c	
	Mg	105.29 ± 129.10 ^a	236.26 ± 15.36 ^b	264.11 ± 33.99 ^c	
	Na	185.55 ± 207.07 ^a	363.83 ± 71.76 ^a	320.63 ± 41.11 ^a	
	Cd	0.12 ± 0.19 ^a	0.05 ± 0.09 ^a	BDL	
	Cr	0.27 ± 0.34 ^a	0.07 ± 0.01 ^b	0.06 ± 0.01 ^b	
	Cu	0.33 ± 0.13 ^a	0.16 ± 0.06 ^b	0.25 ± 0.10 ^{a,b}	
	Fe	8.27 ± 3.19 ^a	1.62 ± 0.54 ^b	1.58 ± 0.77 ^b	
	Mn	0.33 ± 0.16 ^a	0.25 ± 0.07 ^a	0.17 ± 0.11 ^b	
	Pb	0.30 ± 0.25 ^a	0.02 ± 0.01 ^b	0.01 ± 0.01 ^c	
	Sr	1.25 ± 0.64 ^a	0.33 ± 0.14 ^b	0.49 ± 0.26 ^b	
	Zn	10.48 ± 4.92 ^a	3.40 ± 0.68 ^b	3.21 ± 0.77 ^b	
	Gills	Ca	4253.80 ± 5131.17 ^a	9283.52 ± 1273.91 ^b	9014.05 ± 1337.04 ^b
		K	529.26 ± 553.50 ^a	1676.47 ± 260.34 ^b	1688.21 ± 236.43 ^b
Mg		155.25 ± 174.05 ^a	418.39 ± 70.92 ^b	439.64 ± 72.62 ^b	
Na		220.30 ± 243.82 ^a	796.50 ± 168.31 ^b	905.11 ± 168.73 ^b	
Cd		0.16 ± 0.35 ^a	0.07 ± 0.13 ^a	0.01 ± 0.01 ^a	
Cr		0.29 ± 0.10 ^a	0.14 ± 0.03 ^b	0.23 ± 0.01 ^a	
Cu		0.66 ± 0.27 ^a	0.51 ± 0.18 ^a	0.62 ± 0.20 ^a	
Fe		30.29 ± 18.69 ^a	22.35 ± 8.05 ^a	25.24 ± 11.88 ^a	
Mn		8.37 ± 5.15 ^a	7.64 ± 3.11 ^a	6.20 ± 1.92 ^a	
Pb		1.36 ± 1.92 ^a	0.08 ± 0.04 ^b	0.06 ± 0.03 ^b	
Sr		40.22 ± 36.56 ^a	18.81 ± 3.22 ^a	21.33 ± 3.80 ^a	
Zn		61.78 ± 9.12 ^a	60.97 ± 10.26 ^a	68.88 ± 9.48 ^a	
Liver		Ca	97.40 ± 63.12 ^a	165.50 ± 87.64 ^a	165.87 ± 95.00 ^a
		K	496.23 ± 637.97 ^a	1760.90 ± 370.74 ^b	2110.18 ± 397.94 ^c
	Mg	66.51 ± 114.83 ^a	139.03 ± 44.13 ^b	161.54 ± 38.14 ^b	
	Na	200.89 ± 297.06 ^a	557.94 ± 117.29 ^b	744.42 ± 128.18 ^c	
	Cd	0.18 ± 0.35 ^a	0.17 ± 0.49 ^a	0.07 ± 0.17 ^a	
	Cr	0.81 ± 0.40 ^a	0.41 ± 0.15 ^b	1.21 ± 1.32 ^{a,b}	
	Cu	2.03 ± 1.41 ^a	2.54 ± 1.03 ^a	4.33 ± 1.36 ^b	
	Fe	52.32 ± 43.23 ^a	66.15 ± 35.52 ^a	70.46 ± 11.64 ^a	
	Mn	2.54 ± 2.08 ^a	2.13 ± 1.45 ^a	2.00 ± 1.47 ^a	
	Pb	1.48 ± 1.02 ^a	0.29 ± 0.24 ^b	0.04 ± 0.03 ^c	
	Sr	1.85 ± 0.90 ^a	0.69 ± 0.55 ^b	1.07 ± 0.67 ^{a,b}	
	Zn	36.52 ± 29.58 ^a	23.56 ± 9.36 ^a	18.59 ± 6.79 ^a	

BDL: Below detection limit.

^{a,b,c} The values with different letters in the same row are significantly different (LSD Multiple Comparison test, $p < 0.05$).

Table 4

Correlation coefficients between trace element concentrations in tissues of chub from the River Szamos/Someş and feeding groups, significant at $p < 0.05$ (N = 33).

Elements	Tissues		
	Muscle	Gills	Liver
K	0.834	0.575	0.708
Mg	0.625	0.564	0.526
Na	n.s.	0.688	0.693
Cr	-0.749	n.s.	n.s.
Cu	n.s.	n.s.	0.549
Fe	-0.643	n.s.	n.s.
Mn	-0.395	n.s.	n.s.
Pb	-0.791	-0.624	-0.888
Zn	-0.643	n.s.	n.s.

n.s.: Non-significant.

Table 5

Values of the bioconcentration factor (BCF) based on the concentration of trace elements (mg kg^{-1} wet weight) in the specific tissue and the concentration of trace elements (mg l^{-1}) in the water of the River Szamos/Someş (mean value \pm standard deviation).

Elements	Tissue	Age 0+	Age 1+, 2+	Age 3+ \geq
Cd	Muscle	203.40 \pm 336.81 ^a	88.05 \pm 159.41 ^a	–
	Gills	275.12 \pm 600.70 ^a	123.12 \pm 232.64 ^a	12.40 \pm 28.21 ^a
	Liver	319.63 \pm 609.11 ^a	303.77 \pm 851.01 ^a	126.67 \pm 288.73 ^a
Cr	Muscle	85.96 \pm 108.46 ^a	20.68 \pm 4.05 ^b	19.07 \pm 2.25 ^b
	Gills	92.39 \pm 32.59 ^a	43.52 \pm 8.07 ^b	72.41 \pm 5.74 ^a
	Liver	253.62 \pm 126.2 ^a	130.40 \pm 46.42 ^b	380.88 \pm 416.36 ^{a, b}
Cu	Muscle	34.75 \pm 14.08 ^a	17.24 \pm 5.99 ^b	26.44 \pm 10.54 ^a
	Gills	69.28 \pm 28.22 ^a	53.59 \pm 19.28 ^a	65.63 \pm 21.42 ^a
	Liver	214.54 \pm 149.01 ^a	268.41 \pm 109.13 ^a	457.36 \pm 143.25 ^b
Fe	Muscle	22.62 \pm 8.71 ^a	4.42 \pm 1.48 ^b	4.31 \pm 2.11 ^b
	Gills	82.82 \pm 51.09 ^a	61.11 \pm 22.01 ^a	69.02 \pm 32.48 ^a
	Liver	143.04 \pm 118.19 ^a	180.84 \pm 97.11 ^a	192.64 \pm 31.83 ^a
Mn	Muscle	2.26 \pm 1.06 ^a	1.69 \pm 0.47 ^{a, b}	1.19 \pm 0.75 ^b
	Gills	57.33 \pm 35.27 ^a	52.32 \pm 21.27 ^a	42.46 \pm 13.16 ^a
	Liver	17.38 \pm 14.27 ^a	14.56 \pm 9.94 ^a	13.70 \pm 10.08 ^a
Pb	Muscle	42.69 \pm 36.55 ^a	2.36 \pm 1.50 ^b	1.17 \pm 0.85 ^c
	Gills	196.14 \pm 277.06 ^a	12.26 \pm 6.19 ^b	8.74 \pm 4.65 ^b
	Liver	214.18 \pm 147.40 ^a	41.25 \pm 35.39 ^b	6.42 \pm 4.06 ^c
Zn	Muscle	76.90 \pm 36.13 ^a	24.95 \pm 5.02 ^b	23.57 \pm 5.67 ^b
	Gills	453.54 \pm 66.92 ^a	447.54 \pm 75.35 ^a	505.63 \pm 69.61 ^a
	Liver	268.12 \pm 217.18 ^a	172.97 \pm 68.71 ^a	136.45 \pm 49.83 ^a

Notation: The Cd concentration was below the detection limit in the case of the muscle samples of the older chubs (age 3+ and thereafter).

^{a, b, c} The values with different letters in the same row are significantly different (LSD Multiple Comparison test, $p < 0.05$).

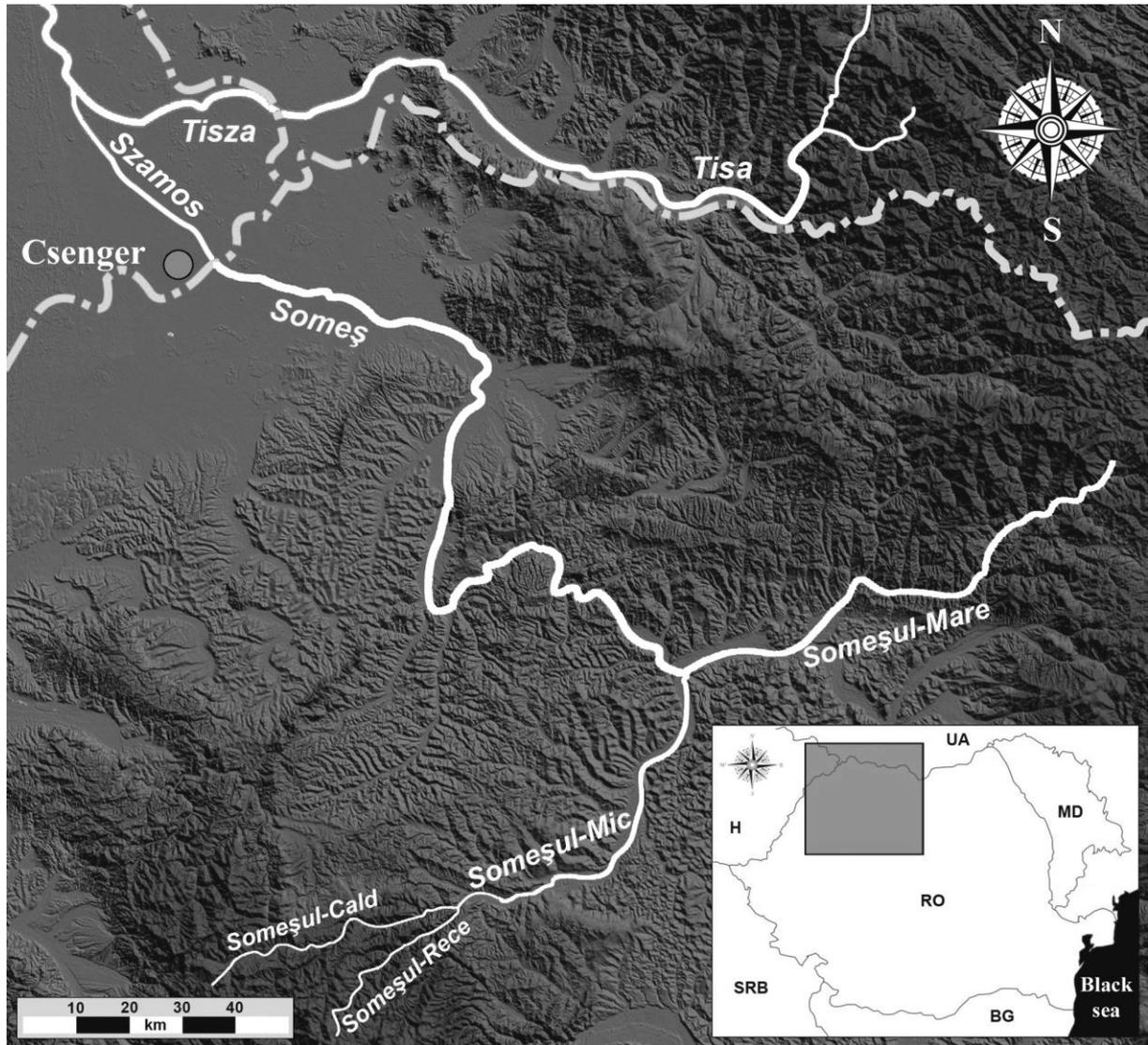


Fig. 1. Map of the Water System of River Szamos/Someș and the sampling area at the village of Csenger

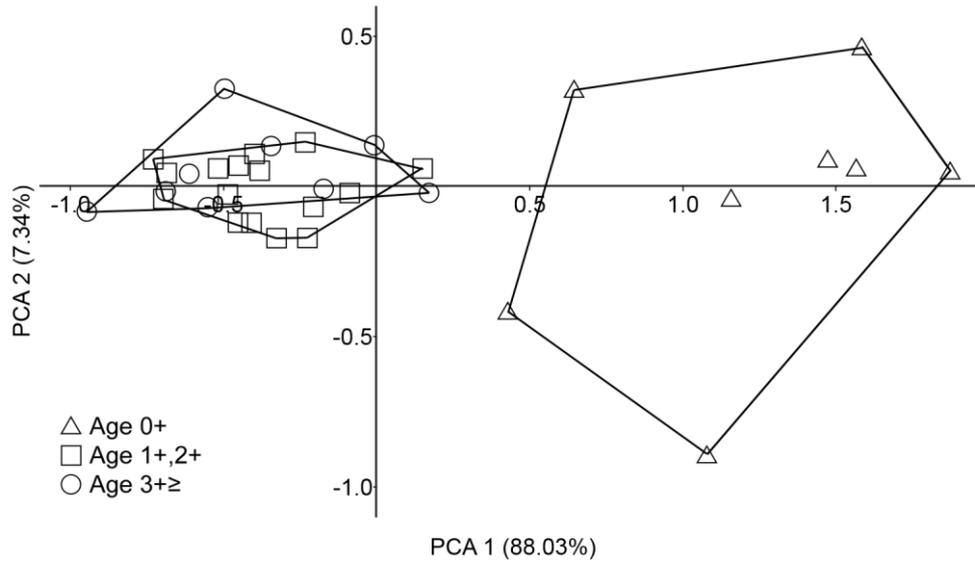


Fig. 2. Principal component analysis of trace element concentrations (mg kg^{-1} , wet weight) in the muscles of the three feeding groups of chub in the River Szamos/Someş.

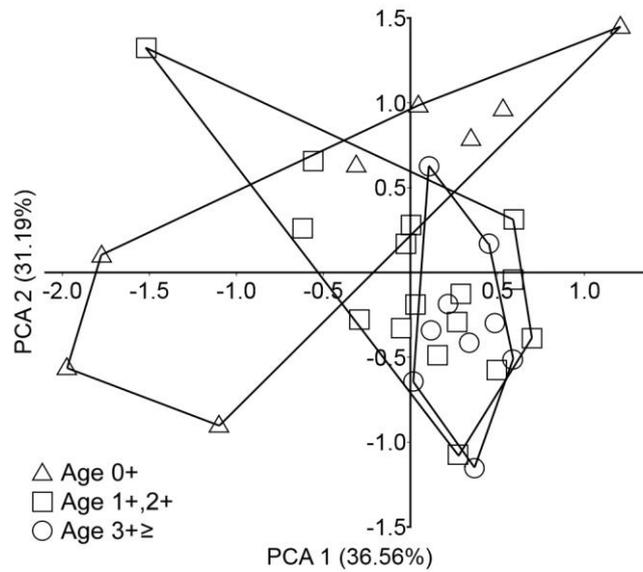


Fig. 3. The PCA plot element concentrations (mg kg^{-1} , wet weight) in the livers of the three feeding groups of chub in the River Szamos/Someş.

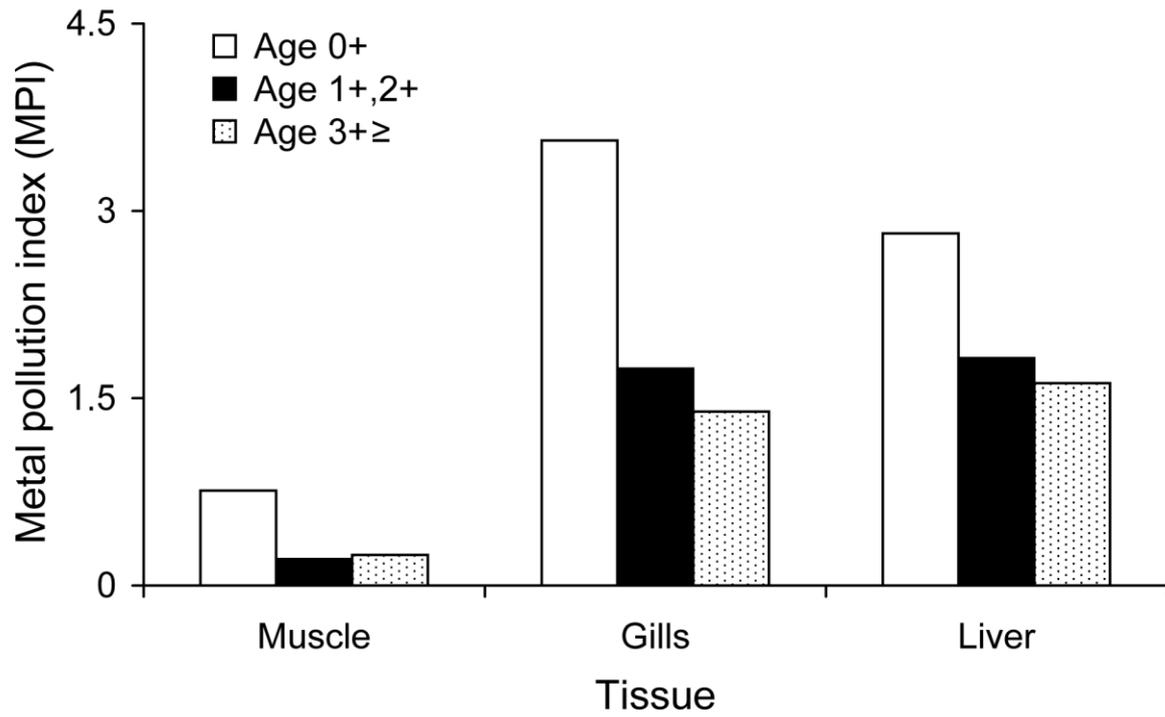


Fig. 4. Mean metal pollution index (MPI) for various tissues of different feeding groups of chub in the River Szamos/Someş.