

# The triggering role of *Clostridioides difficile* infection in relapsed IBD outpatients

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Although the exact aetiology of inflammatory bowel disease (IBD) is unknown, one hypothesis suggests that the inflammation may be the consequence of an altered or pathogenic microbiota in a genetically susceptible host. The aim of this study was to assess the frequency of enteral infections in patients with relapse of IBD, and to evaluate the clinical utility of faecal calprotectin (FC) and faecal matrix metalloproteinase-9 (MMP-9) in the differential diagnosis of relapses with different origins, and to determine the recurrence rate of *Clostridioides difficile* (*C. difficile*), the hospitalisation and colectomy rate among *C. difficile* positive IBD patients at the end of 4 years follow-up period.

**Methods:** In this prospective, “real life” study clinical data, serum and stool samples were assessed.

**Results:** Overall, 135 outpatients with IBD were enrolled [91 IBD patients who relapsed and 44 subjects in clinical remission (control group)]. *C. difficile* A/B toxins were detected in 42.2% of all cases. *Candida* was presented in 9.9% among the enrolled subjects. We found significant difference between FC and MMP-9 values in patients in relapse and remission, but not in *C. difficile* positive and negative cases. Our results revealed an association between previous antibiotic use and the rate of toxigenic *C. difficile*. Toxigenic *C. difficile* positivity recurrence rate was 4.4%. Hospitalisation during follow-up due to IBD was 45.4% and 35% in *C. difficile* positive and negative group, respectively. Value of FC and MMP-9 did not predict the need of hospitalisation.

**Conclusions:** The occurrences of toxigenic *C. difficile* and *Candida* positivity were excessively high in our patients in an acute relapse, which suggests the importance of intestinal microbiota in IBD. FC and MMP-9 has no diagnostic value to differentiate between infection-induced and natural relapse. In our study was confirmed that hospitalisation rate was higher in *C. difficile* positive cases, but we did not find any relationship on long-term period.

**KEYWORDS:** *Clostridioides difficile*, ulcerative colitis, Crohn’s disease, faecal calprotectin, matrix metalloproteinase-9

## Introduction

The course of the inflammatory bowel diseases (IBD: Crohn’s disease – CD – and ulcerative colitis – UC) are chronic or intermittent and coloured by multiple flare-ups, despite long-term maintenance treatment in about half of the patients (1, 2). Just as with the initial manifestation of IBD, the factors which cause subsequent relapses are also ambiguous in most cases; however, an infective agent may play a role in both the manifestation and the relapse (3).

Some animal experiments suggest a role is played by bacteria in the pathogenesis of IBD: interleukin-10 knockout, germ-free mice do not develop colitis; however, interleukin-2-deficient mice develop spontaneous colitis after *Escherichia coli* exposure, but not with *Bacteroides vulgatus* exposure (4). Although the aetiology of IBD has not been unequivocally clarified, an altered intestinal immune response due to the interaction of several environmental and genetic factors is observable, which may lead to inadequate gut flora compound and function, thus

making the intestine vulnerable to superinfections like the infections caused by toxigenic *Clostridioides difficile* (*C. difficile*) (5, 6, 7). *C. difficile* infections came to the front since epidemiological data showed high prevalence of toxin-producing isolates (8, 9) among IBD patients. Current data suggest that in patients with CD or UC, infections caused by this anaerobic bacterium occur more frequently than in the general population (10). The significance of toxigenic *C. difficile* positivity manifests not only in its increasing incidence, growing antibiotic resistance and severe disease course, but also in the magnitude of the infections, which also leads to high health care costs. Expansion of *C. difficile* infections is more pronounced in IBD patients; moreover, its prevalence has been reported in 5 to 19% in patients with active disease (11, 12). According to previous studies, the rate of *C. difficile* infection is approximately 4-5 times greater than in non-IBD patients (13, 14). However, most of studies are retrospectively performed.

The usual diagnostic tools in case of a relapse of IBD consist of faecal microbiological examination and an evaluation of inflammatory biochemical and stool biomarkers. A simply and quickly performing marker would be useful as strong point in the differentiation of an infective origin or exacerbation. Faecal calprotectin (FC) and matrix metalloproteinase-9 (MMP-9) are markers that correlate with disease activity. FC is a neutrophil leukocyte-derivate protein eligible for distinguishing IBD from functional disorders (15). In vitro resistance to degradation allows faecal samples to be assayed for reliable calprotectin determination. FC has a high sensitivity in the determination of IBD activity; however, elevated levels were also shown in intestinal infections, tumours and colon polyps, therefore its specificity is less favourable. MMP-9 is a Zn-binding neutral proteinase produced by polymorphonuclear cells. MMP-9 is one of the major contributors to the breakdown and reconstruction of the extracellular matrix (16). Faecal MMP-9 and FC in UC patients is an excellent marker of disease activity and shows significant correlation with clinical and endoscopic scores (17).

The objective of this study was to prospectively assess the prevalence of enteral infections (especially toxigenic *C. difficile* and *Candida* sp.) among our outpatients with IBD in relapse. Patients in remission were examined as a control group. In addition, our secondary endpoint was to estimate the utility of clinical and differential diagnoses of FC and MMP-9 in cases of both with infection and without infection. The tertiary endpoint was to assess the long-term outcome of *C. difficile* infection in IBD patients. We aimed to determine the recurrence rate of *C. difficile*, the hospitalisation and colectomy rate among *C. difficile* positive IBD patients at the end of 4 years follow-up period.

## Patients and methods

### Patient selection

91 IBD patients with symptoms of relapse were enrolled in our study with 44 control IBD subjects in clinical remission from September, 2012 to July, 2013. All of them were outpatients at the 1st Department of Medicine,

Szeged, Hungary. The medical data of patients were collected prospectively. Clinical data, activity indexes, faecal specimens and serum samples were analysed of 91 IBD patients who manifested worsening symptoms – indicating a potential relapse – and of 44 IBD patients in remission. Blood samples were taken from all patients for the determination of CRP level (C-reactive protein), platelet and leukocyte count, serum iron level and haemoglobin level. Faecal samples from every subject were sent to the microbiological laboratory of the Institute of Clinical Microbiology, University of Szeged. The clinical activity of CD and UC were assessed using the Crohn's Disease Activity Index (CDAI) and with a partial Mayo Score (pMayo), respectively. Relapse was defined as  $\geq 150$  points and/or an increase of  $\geq 80$  points in the CDAI in the case of CD and  $\geq 2$  points in the pMayo score in the case of UC. Immuno-

**Table 1. Clinical characteristics of patients at the time of enrolment**

	Relapsing group (n=91)	Control group (n=44)
Gender (male/female) No. of patients	43/48	26/18
Age Years, mean (min-max)	36.8 (16 to 79)	40 (19 to 75)
UC/CD No. of patients	49/42	25/19
Disease location (based on Montreal classification of IBD)		
L1	15	6
L2	7	6
L3	20	7
E1	11	8
E2	20	6
E3	18	11
Disease duration (years) Years, mean (min-max)	6.9 (0 to 26)	10.9 (0 to 46)
Appendectomy No. of patients	4	1
Smoking No. of patients	17	5
Extraintestinal manifestation No. of patients	36	9
Immunosuppression No. of patients	46	25
Gastric acid-suppressant agent No. of patients	25	8
Antibiotic use in previous 3 months No. of patients	18	4
Previous <i>C. difficile</i> positivity No. of patients	23	7

suppression was defined by Consensus of the European Crohn's and Colitis Organisation (ECCO) (3). Data of the patients regarding smoking status, previous appendectomy, perianal involvement, the presence of extraintestinal manifestations of IBD, previous antibiotic use and concomitant medications were collected from each subject. The clinical characteristics of the patients are presented in *Table 1*.

### Microbiological examination

A microbiological analysis for the presence of enteric pathogens (including toxigenic *C. difficile*, *Campylobacter* spp., *Salmonella* spp., *Shigella* spp., *Yersinia* spp., *Candida* spp. and in reasonable cases viruses and parasites) was carried out at the Institute of Clinical Microbiology, Szeged, Hungary. Laboratory diagnosis of toxigenic *C. difficile* was based on ESCMID guideline (18): faecal samples were tested for the presence of glutamate dehydrogenase (GDH) and toxin A/B using the QUIK CHEK COMPLETE (TechLab). Samples with a GDH-positive, but toxin A/B negative results were further tested with toxigenic culture method. Toxigenic culture is one of reference method for diagnosing *C. difficile* infection. The basis of this method is isolation of *C. difficile* and testing the toxin production in vitro by a sensitive A/B toxin detection method (18).

### Measurement of faecal calprotectin and MMP-9

Samples for FC and MMP-9 determination were stored at  $-20^{\circ}\text{C}$  until analysed. Faecal specimens were thawed and prepared for a calprotectin assay as described by the manufacturer. FC levels were measured by

**Table 2. Inflammatory biomarkers and clinical activity indices of IBD patients at the time of enrolment**

	Relapsing group (n=91)	Control group (n=44)
CDAI mean (min-max)	219.6 (105 to 450)	60.7 (0 to 127)
pMayo Score mean (min-max)	5.2 (2 to 9)	0.6 (0 to 2)
FC ( $\mu\text{g/g}$ ) median (min-max)	640 (29 to 14000)	79 (29 to 860)
Fecal MMP-9 (ng/mL) mean (min-max)	11.4 (0 to 168.8)	5.6 (0 to 23.2)
CRP (mg/L) mean (min-max)	23.1 (1 to 202.6)	6.5 (1 to 40.8)
Leucocytes (G/L) mean (min-max)	9.8 (3.8 to 90.6)	7.4 (3.6 to 15.1)
Thrombocytes (G/L) mean (min-max)	326 (88 to 657)	292.7 (125 to 602)
Serum iron ( $\mu\text{mol/L}$ ) mean (min-max)	11.5 (1.9 to 25.3)	15.1 (4.3 to 33.1)
Hematocrit (%) mean (min-max)	39 (21 to 49)	40 (33 to 46)

a quantitative lateral flow assay (Quantum Blue, Bühlmann Laboratories, Switzerland). Samples for MMP-9 determination were thawed and prepared as described by the manufacturer. In a few words, one gram of the sample was diluted and homogenised in 4 ml of an ice-cold buffer (0.15M NaCl + 20 mM Tris-HCl, pH: 8.3). The suspension was then centrifuged at 1500g for 10 min and the supernatant was recentrifuged at 10,000 g for 10 min. The final supernatant was filtered and stored at  $-20^{\circ}\text{C}$  until analysed. MMP9 concentrations were determined by ELISA method (Quantikine MMP9 assay, R&D System, UK).

### Statistical analysis

The collected data was analysed statistically, using Pearson's chi-square test, Pearson's chi-square test with Yates' continuity correction and Wilcoxon rank sum test with continuity correction.  $P < 0.05$  was considered as statistically significant. For the statistical analysis, SPSS15.0 (SPSS Inc., Chicago, IL, USA) was used.

### Ethical Considerations

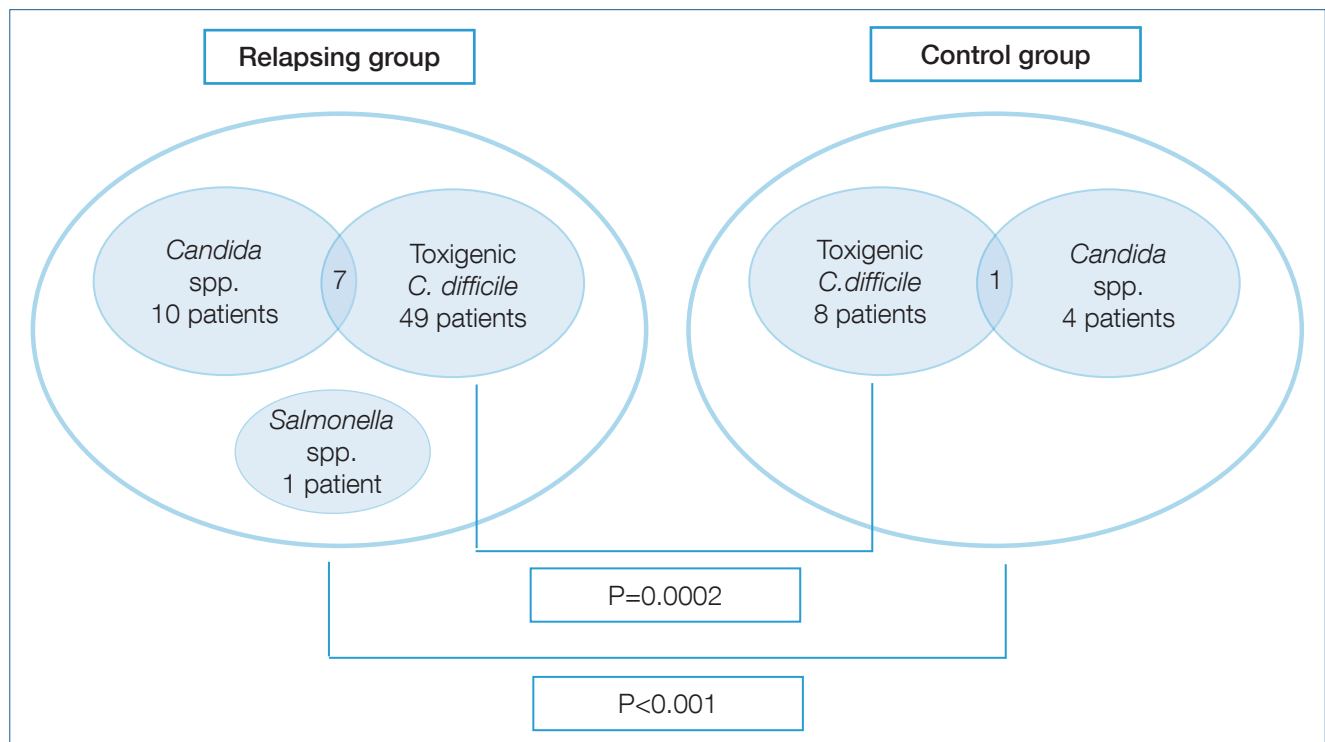
The study was approved by the Regional and Institutional Human Medical Biological Research Ethics Committee of the University of Szeged (ethical approval number: SZTE-170/2011).

## Results

### Infection Rates

Out of the 135 enrolled patients (91 with relapse and 44 control), 74 subjects were diagnosed with UC and 61 with CD. Forty-nine UC and 42 CD patients participated in the relapse group. The number of UC and CD patients was 25 vs. 19 subjects in the control group (*Table 1*). In the relapsing group the mean value of the Crohn's Disease Activity Index (CDAI) was 219.6 points (in the range of: 105 to 450) and the mean value of the pMayo score in the case of UC was 5.2 (in the range of: 3 to 9) points at the time of enrolment (*Table 2*). The most frequent disease locations in the relapse cohort were left sided and pancolitis (20 and 17 subjects) in the case of UC, and pure ileal and the ileocolonic in the case of CD (15 and 20 subjects). Approximately half of the patients were immunosuppressed in the relapse group and also in the control group (*Table 1. and 2*).

Toxigenic *C. difficile* or *Candida* spp. was identified in 53 of 91 faecal samples taken from relapsing patients; on the other hand, 11 of 44 patients had positive microbiological stool findings in the control group in remission ( $p < 0.001$ ). 49 participants with positivity of *C. difficile* A/B toxins had been detected among the patients in relapse (53.8%), while toxigenic *C. difficile* positivity was observed in 8 patients (18.2%) in the remission group. Other microbiological findings in the relapse group were the presence of *Candida* species (10 patients) in all except one subject, who had *Salmonella* infection. *Candida* species occurred in 4 patients in the control group. In 7 cases we found simultaneous presence of toxigenic

**Figure 1. Infection rates among patients with relapse and remission of IBD at the time of enrolment**

*C. difficile* and *Candida* sp. in relapse group, and in one case in the control group (Figure 1.).

A significant difference was revealed between the relapse and the control group regarding the positive cases of *C. difficile* ( $P=0.0002$ ). We found that relapse of IBD is a predicting factor for *C. difficile* positivity ( $P=0.01$ ). 23 subjects in the relapse group had previous toxigenic *C. difficile* positivity in their medical history, although only 14 of the currently infected patients were positive in the past. Six patients had recurrent toxigenic *C. difficile* positivity in relapsing group during follow-up period. Seven of the control subjects had positive toxigenic *C. difficile* results previously, but only 2 of currently *C. difficile*-positive subjects had a medical history of *C. difficile* infections. We did not find a difference between CD and UC regarding positive microbiological examination findings or toxigenic *C. difficile* positivity (29 subjects with UC and 26 with CD). There was no significant connection between disease extension, extraintestinal manifestations, perianal involvement, smoking, age, gender, disease duration, immunosuppression and gastric acid-suppressant agent use and toxigenic *C. difficile* positivity. Previous antibiotic use was 19.8% in the relapse group and 9% in the control group, moreover it was more frequent among *C. difficile*-positive subjects ( $p=0.05$ ). Therefore, previous antibiotic use was shown to have a connection to toxigenic *C. difficile* positivity, but other assessed factors did not predict the presence of *C. difficile* toxins.

Patients in whom *C. difficile* toxins had been detected in the relapse group were treated with metronidazole. A combination of antibiotics was used, or a substitute was used for metronidazole (rifaximin or vancomycin) in case of metronidazole inefficiency, or if repeated stool exami-

nations continued to be positive. Ciprofloxacin and metronidazole combination was used in case of suspicions natural relapse of CD (5 patients). Antibiotic switch was needed in 18.8% of *C. difficile* positive cases in relapsing group (50% to vancomycin). Three patients needed hospitalisation in a Department of Infectious Diseases due to a therapy-resistant case of *C. difficile* positivity. However, the reasonable administration of a supplementary therapy or a modification of the maintained treatment was sufficient in 45.1% of the cases (oral or topical corticosteroids, 5-ASA and anti-TNF-therapy). Twenty two subjects required hospitalisation because of a relapse. Hospitalisation rate was not associated with initial level of FC or MMP-9 in natural relapse or in *C. difficile* positive cases.

#### Long-term outcome

The mean follow-up period was 51 (SD 8.2) months in our study. Toxigenic *C. difficile* positivity recurrence rate was 4.4%. The hospitalisation rate was 54.8% during follow-up period, within that hospitalisation due to IBD was 39.2%, due to *C. difficile* was 2.2% and due to other reasons was 31.8%. More patients with relapse needed hospitalisation during follow-up period ( $P=0.002$ ) than patients in remission. Despite that, value of FC and MMP-9 did not predict the need of hospitalisation. The rate of hospitalisation due to any reason during follow-up period was 54.4% in *C. difficile* positive group and 55% in *C. difficile* negative group therefore *C. difficile* positivity itself did not predict need of hospitalisation on long-term period. On the other hand, hospitalisation during follow-up due to IBD was 45.4% and 35% in *C. difficile* positive and negative group, respectively. Colectomy rate during follow-up was 11.8%.

### Laboratory Findings

The mean values of CRP in the relapse and the control group were 23.08 mg/L (in the range of: 1 to 202.6 mg/L) and 6.46 mg/L (in the range of: 1 to 40.8 mg/L), respectively. CRP was slightly higher in those with CD than in those with UC (the mean values of which are: CD: 25.17 mg/L vs. UC: 21.31 mg/L) in relapse group, but on the other hand, this difference was similar in the control group (the mean values of which are: CD: 9.68 mg/L vs. UC: 4.12 mg/L). The mean values of the leukocyte and the thrombocytes count, the serum iron and the hematocrit levels were 9.79 G/L, 326.05 G/L, 11.48  $\mu$ mol/L and 39% in the relapse group, and 7.4 G/L, 292.7 G/L, 15.1  $\mu$ mol/L and 40% in the control group (Table 2.).

### Faecal Calprotectin Levels

The median value of FC in those in relapse (640  $\mu$ g/g, in range 29-14000  $\mu$ g/g) was significantly higher compared to the control group (79  $\mu$ g/g, in range 29-860  $\mu$ g/g) ( $P < 0.001$ ). FC did not showed difference in relapsing patients without (with the median value of: 510  $\mu$ g/g) vs. with *C. difficile* positivity (with the median value of: 690  $\mu$ g/g).

### Faecal MMP-9 Levels

Similarly to FC, faecal MMP-9 levels were higher in the relapse group (11.36 ng/mL; in the range of: 0 to 168.79 ng/ml) compared to the control group (5.61 ng/mL; in the range of: 0 to 23.28 ng/mL;  $P = 0.001$ ). We found obvious numerical difference between the mean values in cases of microbiological (*C. difficile* or *Candida* spp.) positive (mean: 8.0 ng/ml) and negative cases (mean: 14.97 ng/ml) in the relapse cohort; however, statistical analysis did not confirm the difference.

## Discussion

Enteral infection rate during an IBD relapse, the clinical utility of faecal inflammatory markers (FC and MMP-9) and long-term outcome after presence of toxigenic *C. difficile* were assessed in this study. The occurrence of toxigenic *C. difficile* positivity was remarkably high in our IBD patients (42.2%) compared to data of previous studies. Incidence was even higher in cases of relapse, suggesting some connection between flare-ups of IBD and *C. difficile* positivity. A differential diagnosis of a relapse of IBD and *C. difficile* infection is difficult in some cases; furthermore, both can occur at the same time, thus a faecal microbiological examination and tests for both toxins A/B is recommended in the event of a relapse, as the evidence-based consensus of ECCO also suggests (3).

Some studies suggest that the aetiology of IBD with altered intestinal microbiome is a result of innate genetic defects in the intestinal epithelial and mucosal barriers. In addition, the aetiological background may involve a microbial imbalance in the intestine or dysfunction in the intestinal inflammatory cascade, leading to pathologic proliferation of cytokines (4). Although an exact aetiology of IBD has not been confirmed yet, the importance of microorganisms in the intestine is obvious. Toxigenic *C. difficile*

positivity (53.8%) and *Candida* (9.9%) colonization were the most frequent results of faecal microbiological examinations in our relapse cohort, suggesting an imbalance of intestinal microflora.

It is well-known that the infection rate in patients with an IBD is greater than in the normal population; infectious agents are involved in the etiopathogenesis, but it can also initiate a relapse of the IBD (15). IBD is an independent risk factor of *C. difficile* positivity, mainly UC patients are susceptible to infection, and thus microorganisms have considerable significance in patient management (3, 18–20). In our study, the UC/CD ratio in the *C. difficile* cohort was approximately equal (25 UC vs. 23 CD subjects).

The disease course of a *C. difficile* infection can be extremely serious, even life-threatening complications may occur, which is confirmed beyond a doubt by the mortality rate of hospitalised IBD patients with a *C. difficile* infection (21). *C. difficile* infection was not severe in most of the cases in our study, serious complications did not occur during the tenure of the study. However, hospitalisation was needed in 13 of 49 cases with positive result of toxigenic *C. difficile* in relapsing group, most of them because of a severe flare-up requiring intravenous treatment. It seems that toxigenic *C. difficile* positivity is also a trigger and an additive factor in relapse of IBD. Hospitalisation, colectomy and mortality rates are higher in a population with toxigenic *C. difficile* positivity (12). In our study was confirmed that hospitalisation rate was higher in *C. difficile* infections, but we did not find any relationship on long-term period. In some studies, antimicrobial and immunomodulator agent-exposure has been shown to increase the possibility of a *C. difficile* infection and carriage (11). Basically, any type of antibiotic may be linked with the manifestation of a *C. difficile* infection; however, the antibiotics most commonly connected to an occurrence *C. difficile* infection include penicillins, clindamycin, cephalosporins and fluoroquinolones (22). Regardless, antibiotic use for IBD patients in general seems to be a critical risk factor for the development of a *C. difficile* positivity infection, as our results showed. The use of immunomodulators specifically has not been confirmed in our study to be predictive of a *C. difficile* positivity risk, however. The data available on immunomodulators is conflicting; some studies showed an increasing risk for *C. difficile* positivity (11), although others did not confirm these results (23). The number of toxigenic *C. difficile*-positive subjects taking immunosuppressants did not differ substantially compared to those patients not treated with immunosuppressants in our study. A correlation between *C. difficile* positivity and the use of gastric acid-suppressive agents, especially histamine blockers and proton pump inhibitors, has been suggested (24). However, similarly to our own findings, other observational studies did not yield results to support the connection (25).

A strong correlation between intestinal inflammation and FC has been confirmed in IBD (26); to note, its value can increase in enteral infections also (27). Enhanced levels of MMP-9 have been observed in the intestinal tissue in the case of IBD and seem to be actively involved in the inflammatory and remodelling process (28). In this study, levels

of FC and MMP-9 were measured both in cases of infection and non-infection. Although we discover higher levels of FC and faecal MMP-9 in natural relapses than in infection-induced relapses, the statistical analysis did not confirm this observation.

Colonising microbes inhabiting the intestinal mucosa are important for normal intestinal function and homeostasis. Dysbiosis develops when the composition of intestinal microbiome is unfavourable, and this condition may lead to various diseases. Altered microbiome may be a cause or consequence of a disease, but it may develop parallel with an abnormal condition (29). The balance of intestinal homeostasis is a sensitive system of intestinal microbiota, epithelium and immune cells (30). An alteration of colonic microbiota has been suggested in IBD (31), given that intestinal flora is altered in patients who relapse, which in turn allows the unopposed proliferation of *C. difficile* and *Candida*. The frequency of the community-acquired *C. difficile* infection has been increasing worldwide with incidence rates of 3 to 40% (10–12). *Candida* was presented in 9.9% among the enrolled subjects. Presence of *Candida* in gastrointestinal tract is commonly benign, but in susceptible individuals with low level of inflammation (eg. IBD) may promote colonisation that promotes further inflammation, thus may become a trigger factor (32). The role of *C. difficile* may be the similar. Our data showed 42.2% of occurrences of *C. difficile* positivity in IBD outpatients, and 53.8% of the patients in the relapse cohort were *C. difficile* toxin positive. We revealed a significant difference regard-

ing *C. difficile* positivity between IBD patients in relapse and those in remission. The occurrences of toxigenic *C. difficile* and *Candida* positivity were excessively high in our patients in an acute relapse, which suggests the importance of intestinal microbiota in IBD; therefore, in conclusion we recommend a faecal microbiological examination in every relapse to optimise therapy.

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## Irodalom

1. Stange EF, Travis SPL, Vermeire S, Reinisch W, Geboes K, Barageuskienė A, Feakins R, Fléjou JF, Herfarth H, Hommes DW, Kupcinskas L, Lakatos PL, Mantzaris GJ, Schreiber S, Villanacci V, Warren BF; European Crohn's and Colitis Organisation (ECCO). European evidence-based Consensus on the diagnosis and management of ulcerative colitis: Definitions and diagnosis. JCC 2008; 2(1): 1–23 (PMID: 21172194 DOI: 10.1016/j.crohns.2007.11.001.)
2. Van Assche G, Dignass A, Panes J, Beaugerie L, Karagiannis J, Allez M, Ochsenkühn T, Orchard T, Rogler G, Louis E, Kupcinskas L, Mantzaris G, Travis S, Stange E; European Crohn's and Colitis Organisation (ECCO). The second European evidence-based consensus on the diagnosis and management of Crohn's disease: Definitions and diagnosis. JCC 2010; 4(1): 7–27. (PMID: 21122488 DOI: 10.1016/j.crohns.2009.12.003.)
3. Rahier JF, Ben-Horin S, Chowers Y, Conlon C, De Munter P, D'Haens G, Domènech E, Eliakim R, Eser A, Frater J, Gassull M, Giladi M, Kaser A, Lémann M, Moreels T, Moschen A, Pollok R, Reinisch W, Schunther M, Stange EF, Tilg H, Van Assche G, Vigeat N, Vucelic B, Walsh A, Weiss G, Yazdanpanah Y, Zabana Y, Travis SP, Colombel JF; European Crohn's and Colitis Organisation (ECCO). European evidence-based Consensus on the prevention, diagnosis and management of opportunistic infections in inflammatory bowel disease. JCC 2009; 3: 47–91. (PMID: 21172250 DOI: 10.1016/j.crohns.2009.02.010.)
4. Kaur N, Chen CC, Luther J, Kao JY. Intestinal dysbiosis in inflammatory bowel disease. Gut Microbes 2011; 2(4): 211–216. (PMID: 21983063 DOI: 10.4161/gmic.2.4.17863.)
5. Machiels K, Joossens M, Sabino J, De Preter V, Arijis I, Eeckhaut V, Ballet V, Claes K, Van Immerseel F, Verbeke K, Ferrante M, Verhaegen J, Rutgeerts P, Vermeire S. A decrease of the butyrate-producing species *Roseburia hominis* and *Faecalibacterium prausnitzii* defines dysbiosis in patients with ulcerative colitis. Gut 2014; 63(8): 1275–1283. (PMID: 24021287 DOI: 10.1136/gutjnl-2013-304833.)
6. Knights D, Lassen KG, Xavier RJ. Advances in inflammatory bowel disease pathogenesis: linking host genetics and the microbiome. Gut 2013; 62(10): 1505–1510. (PMID: 24037875 DOI: 10.1136/gutjnl-2012-303954.)
7. Pruitt RN, Borden LD. Toward a structural understanding of *Clostridioides difficile* toxins A and B. Front in Cell Infect Microbiol 2012; 2: 28. (PMID: 22919620 DOI: 10.3389/fcimb.2012.00028)
8. Honda H, Dubberke ER. The changing epidemiology of *Clostridioides difficile* infection. Curr Opin Gastroenterol 2014; 30(1): 54–62. (PMID: 24285002 DOI: 10.1097/MOG.0000000000000018.)

9. Kazanowski M, Smolarek S, Kinnarney F, Grzebieniak Z. *Clostridioides difficile*: epidemiology, diagnostic and therapeutic possibilities—a systematic review. Tech Coloproctol. 2014; 18(3): 223–232. (PMID: 24178946 DOI: 10.1007/s10151-013-1081-0)
  10. Bossuyt P, Verhaegen J, Van Assche G, Rutgeerts P, Vermeire S. Increasing incidence of *Clostridioides difficile*-associated diarrhea in inflammatory bowel disease. J Crohns Colitis 2009; 3: 4–7. (PMID: 21172241 DOI: 10.1016/j.crohns.2008.09.003.)
  11. Meyer AM, Ramzan NN, Loftus EV Jr, Heigh RI, Leighton JA. The diagnostic yield of stool pathogen studies during relapses of inflammatory bowel disease. J Clin Gastroenterol 2004; 38(9): 772–775. (PMID: 15365403)
  12. Mylonaki M, Langmead L, Pantes A, Johnson F, Rampton DS. Enteric infection in relapse of inflammatory bowel disease: importance of microbiological examination of stool. Eur J Gastroenterol Hepatol 2004; 16(8): 775–778. (PMID: 15256979)
  13. Hourigan SK, Oliva-Hemker M, Hutflless S. The prevalence of *Clostridioides difficile* infection in pediatric and adult patients with inflammatory bowel disease. Dig Dis Sci 2014; 59: 2222–7. doi: 10.1007/s10620-014-3169-4
  14. Harminder Singh, Zoann Nugent, B. Nancy Yu, Lisa M. Lix, Laura E. Targownik, Charles Bernstein. Higher Incidence of *Clostridioides Difficile* Infection Among Individuals With Inflammatory Bowel Disease. Gastroenterology 2017; 153(2): 430–438.e2. DOI: 10.1053/j.gastro.2017.04.044
  15. Irving PM, Gibson PR. Infections and IBD. Nat Rev Gastroenterol 2008; 5: 18–27. (PMID: 18174904 DOI: 10.1038/npgasthep1004.)
  16. Gao Q, Meijer MJ, Kubben FJ, Sier CF, Kruidenier L, van Duijn W, van den Berg M, van Hogezaand RA, Lamers CB, Verspaget HW. Expression of matrix metalloproteinases-2 and -9 in intestinal tissue of patients with inflammatory bowel diseases. Digestive and Liver Disease. 2005; 37: 584–592. (PMID: 15869913)
  17. Annaházi A, Molnár T, Farkas K, Rosztóczy A, Izbéki F, Gecse K, Inczei O, Nagy F, Földesi I, Szűcs M, Dabek M, Ferrier L, Theodorou V, Bueno L, Wittmann T, Róka R. Faecal MMP-9: A new noninvasive differential diagnostic and activity marker in ulcerative colitis. Inflamm Bowel Dis 2013; 19(2): 316–320. (PMID: 22550024 DOI: 10.1002/ibd.22996.)
  18. Crobach MJT, Planche T, Eckert E, Barbut F, Terveer EM, Dekkers OM, Wilcox MH, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases: update of the diagnostic guidance document for *Clostridioides difficile* infection. Clinical Microbiology and Infection 2016; 22: S63–S81.
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