Eriophyoid Mites (Acariformes: Eriophyoidea) Collected from *Phyllostachys* spp. in Hungary

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Aceria bambusae ChannaBasavanna, 1966 is reported from Hungary for the first time. The species was collected from the leaf sheaths of the introduced bamboo species, *Phyllostachys rubromarginata* McClure and *Phyllostachys tianmuensis* Z.P. Wang et N.X. Ma (both Poaceae) in Hungary. Morphological differences distinguishing this species from other bambusoid inhabiting congeners are discussed. In addition, new date-locality-host records for 3 eriophyoid species collected from 7 bamboo species are given.

Keywords: Eriophyidae, Aceria, Phyllostachys, ornamental bamboos, new hosts, Hungary.

The subfamily Bambusoideae (fam. Poaceae) is represented by 1,575 species belonging to 115 genera (Ohrnberger, 2002). On bamboo species (*Bambusa* spp., *Phyllostachys* spp., *Sasa* spp., *Yushania* spp.) (Poaceae) 39 eriophyoid mite species have been found (Amrine and Stasny, 1994; Davis et al., 1982; Huang, 2001; Lin et al., 2000; Sukhareva, 1994; Xue et al., 2006). In Hungary three further species were recently described: from greenwax golden bamboo (*Phyllostachys viridiglaucescens* A. Rivière et C. Rivière) *Abacarus korosicsomai* Ripka, 2011 and *Adventacarus turulae* Ripka, 2011, from cock bamboo (*Phyllostachys iridescens* C.Y. Yao et S.Y. Chen) *Mucotergum nigrum* Ripka et al., 2015 (Ripka, 2011; Ripka et al., 2015). No eriophyoid species have previously been recorded from *Phyllostachys angusta*, *Ph. aureosulcata*, *Ph. edulis*, *Ph. fimbriligula*, *Ph. meyeri*, *Ph. rubromarginata* and *Ph. tianmuensis* (Amrine and Stasny, 1994; Davis et al., 1982).

The genus Aceria Keifer, like many eriophyoid mites, is well-known for being host specific. However some feed on multiple species within the same plant genus, and some species live on related plant genera (Oldfield, 1996). Majority of the Aceria spp. inhabits dicotyledonous hosts. Currently four bamboo-dwelling Aceria species are known from

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the different parts of the world (Amrine and Stasny, 1994; Baker et al., 1996; Davis et al., 1982; Wang and Huang, 2011; Xue et al., 2006).

Herein we report on our discovery of a species of the genus *Aceria* found on *Phyllostachys rubromarginata* McClure and *Phyllostachys tianmuensis* Z.P. Wang et N.X. Ma (both Poaceae), which are introduced species from China. We provide evidence that this species is different from *Aceria gilloglii* Keifer, 1963, *Aceria havenensis* (Keifer, 1979) (sensu Keifer) and *Aceria niitakayamensis* Wang et Huang, 2011 in consideration of the Asian origin of the hosts.

Materials and Methods

The eriophyoid mite fauna of Asian bamboo collections, among other bamboo species of Asian origin, was studied on *Phyllostachys angusta* McClure, *Phyllostachys aureosulcata* McClure, *Phyllostachys edulis* (Carrière) J. Houzeau, *Phyllostachys fimbriligula* T. H. Wen, *Phyllostachys meyeri* McClure, *Phyllostachys nigra* var. *henonis* (Mitford) Stapf ex Rendle, *Phyllostachys rubromarginata* and *Phyllostachys tianmuensis* taken from plant samples collected in the Botanical Garden of Szent István University, Gödöllő (Pest county, Central Hungary), Szada (Pest county, Central Hungary), Debrecen (Hajdú-Bihar county, East Hungary) and Egerág (Baranya county, South Hungary) in the years of 2016, 2017 and 2019. The eriophyoid mites were cleared in lactic acid then mounted in Keifer's F-medium with sorbitol and Hoyer's medium on microscope slides (Keifer, 1975). Specimens were examined with the aid of a research phase contrast compound microscope (Nikon Eclipse E600) equipped with a drawing tube (Nikon Y-IDT). A Zeiss Axio Imager A2 microscope was used for making digital microscopic images on slide mounted specimens.

The generic classification was made according to Amrine et al. (2003) together with further updating. The terminology and setal notation used in the morphological descriptions follow Lindquist (1996). The number of measured specimens (n) is given in parentheses following the body length. All measurements of mites were made according to Amrine and Manson (1996) and are given in micrometers. Measurements and means are rounded off to the nearest integer. All measurements, unless specified otherwise, are lengths. For females, the mean and the ranges are given. For males and nymphs, data of five specimens are given.

The scientific name of host plant is used according to The Plant List (2013).

TAXONOMY Superfamily ERIOPHYOIDEA Nalepa Family ERIOPHYIDAE Nalepa Subfamily ERIOPHYINAE Nalepa Tribe ACERIINI Amrine and Stasny Genus Aceria Keifer

Aceria bambusae ChannaBasavanna, 1966 (Figs 1–10)

Aceria bambusae ChannaBasavanna, 1966: 58–59. Plate XIII-1. Aceria havenensis (Keifer, 1979) — Amrine and Stasny, 1994: 51, 493. Aceria bambusae — Sukhareva, 1994: 33–34. Fig. 1. Aceria bambusae — Elhalawany, 2015: 49–52. Fig. 3.

Re-description. Female – Body whitish, vermiform, 185 (173–202, n = 10), 39 (38–40) wide, 40 (36–45) thick. Gnathosoma 14 (13–16), projecting obliquely downwards; dorsal palp genual setae d 3 (3–4), unbranched, pedipalp coxal setae ep 3 (2–3). Chelicerae 12 (11–14). Prodorsal shield 28 (26–30), 33 (29–40) wide, oval; shield pattern composed of an incomplete median line on rear $\frac{1}{3}$, two broken admedian lines, sinuate and subparallel to each other anteriorly, on each side of median line farther apart posteriorly, broken at rear $\frac{1}{3}$; and two incomplete curving and broken submedian lines at anterior $\frac{2}{3}$, forking in front of dorsal tubercle, the inner fork curving back toward tubercle, the outer arm extending laterally to lateral line; posteriorly few granules between the submedian line and lateral shield margins. Tubercles of scapular setae *sc* on rear shield margin, 18 (16–23) apart, diverging, scapular setae *sc* 33 (30–39), directed rearwards. Granules situated in lateral rows on epicoxal areas, i.e. laterally between shield margin and dorsal coxae of legs I and II (*sensu* Chetverikov and Craemer, 2015).

Legs with all usual segments and setae present. Leg I (foreleg) 24 (22–25), femur 8 (7–8), basiventral femoral seta bv 5 (5–7), genu 4 (3–5), antaxial genual seta l'' 16 (13–18), tibia 5 (4–5), short paraxial tibial seta l' located at $\frac{2}{5}$ ($\frac{1}{3}-\frac{1}{2}$) from dorsal base, 4 (3–4), very thin, tarsus 5 (4–5), unguinal tarsal seta u' 2 (1–2), solenidion ω 5 (5–6), distally rounded or with a minute knob, slightly curved, empodium simple, bilaterally symmetrical, 5 (4–5), 5-rayed, each ray of three basal pairs with additional secondary branches. Leg II (rear leg) 22 (22–23), femur 7 (7–8), basiventral femoral seta bv 7 (6–7), genu 3 (no range), antaxial genual seta l'' 3 (3–5) very thin, tibia 4 (3–4), tarsus 4 (4–5), unguinal tarsal seta u' 1 (1–2), solenidion ω 6 (6–7), distally rounded or with a minute knob, slightly curved, empodium simple, bilaterally symmetrical, 5 (5–6), 5-rayed, each ray of three basal pairs with a minute knob, slightly curved, empodium simple, bilaterally symmetrical, 5 (5–6), 5-rayed, each ray of three basal pairs with a minute knob, slightly curved, empodium simple, bilaterally symmetrical, 5 (5–6), 5-rayed, each ray of three basal pairs with a minute knob, slightly curved, empodium simple, bilaterally symmetrical, 5 (5–6), 5-rayed, each ray of three basal pairs with small subdivisions.

Coxigenital area with 5–7 microtuberculate semiannuli. Coxisternae I with many granules, coxisternae II with less granules; anterior seta on coxisternum I, *lb* 6 (5–7), tubercles setae *lb* 11 (10–12) apart, proximal seta on coxisternum I, *la* 12 (11–14), tubercles setae *la* 8 (7–9) apart, proximal seta on coxisternum II, *2a* 30 (22–35), tubercles setae *2a* 19 (17–20) apart. Subcapitular plate with granules. In 50% of the examined specimens the prosternal apodeme not well defined, in the others 9 (8–10).

Opisthosoma with 61 (58–65) dorsal, 63 (59–66) microtuberculate ventral semiannuli. Microtubercles oval dorsally and ellipsoid ventrally. Last 4–6 dorsal annuli with

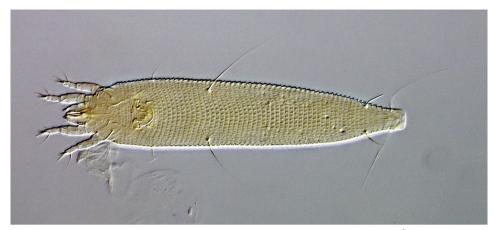


Fig. 1. Digital micrograph of Aceria bambusae, female in ventral view. (Photo: Árpád Szabó)

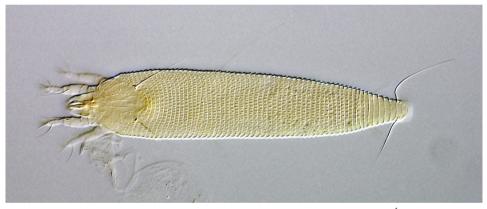


Fig. 2. Digital micrograph of Aceria bambusae, female in dorsal view. (Photo: Árpád Szabó)

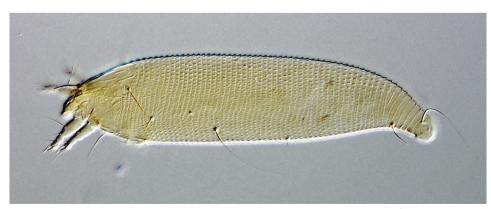


Fig. 3. Digital micrograph of *Aceria bambusae*, female in lateral view. (Photo: Árpád Szabó) *Acta Phytopathologica et Entomologica Hungarica*

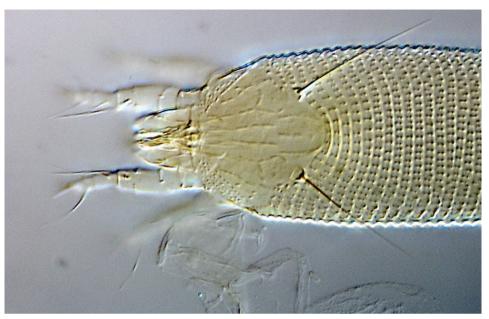


Fig. 4. Digital micrograph of *Aceria bambusae*, prodorsal shield and anterior opisthosoma of female. (Photo: Árpád Szabó)



Fig. 5. Digital micrograph of Aceria bambusae, anterior part of female, in lateral view. (Photo: Árpád Szabó)



Fig. 6. Digital micrograph of *Aceria bambusae*, coxi-genital region and genitalia of female, in lateral view. (Photo: Árpád Szabó)

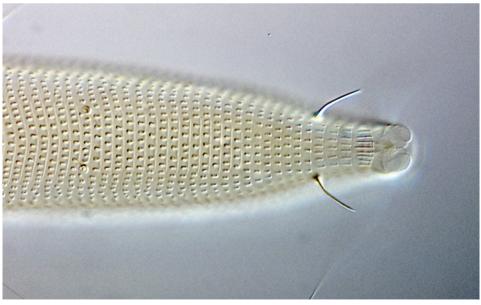


Fig. 7. Digital micrograph of Aceria bambusae, posterior opisthosoma of female, in ventral view. (Photo: Árpád Szabó)

minute microtubercles on rear annular margin. Last 4–5 ventral annuli with linear microtubercles. Opisthosomal setae c2 12 (10–15), on annulus 9 (9–10), 36 (33–38) apart; opisthosomal setae d 53 (48–59), on annulus 22 (20–23), 28 (26–29) apart; opisthosomal setae e 7 (5–10), on annulus 34 (31–36), 14 (12–15) apart; opisthosomal setae f 14 (13–15), on annulus 58 (54–61), or 5 (4–5) from the rear, 13 (10–14) apart. Opisthosomal setae h2 48 (43–55), very thin at apex, 9 (9–10) apart; opisthosomal setae h1 1 (1–2), 6 (5–7) apart. Anal lobes normal.

Genital plate 12 (10–12), 17 (16–18) wide. Female genital coverflap with 8 (7–9) longitudinal ridges; coxisternal III setae 3a 14 (13–15) apart, 7 (6–9), very thin (Figs 1–7).

MALE – Similar to female, white, 140–153 (n= 5), 35 wide, 35–38 thick. Gnathosoma 13–15, projecting obliquely downwards; dorsal palp genual setae d 2–3, unbranched; pedipalp coxal setae ep 1-2. Prodorsal shield 25-28, 28-30 wide, oval, ornamentation similar to female. Tubercles of scapular setae sc on rear shield margin, 17–19 apart, diverging, scapular setae sc 20-22, directed to the rear. Legs with all usual segments and setae present. Leg I 20–22, femur 7–8, basiventral femoral setae by 4–5, very fine, genu 3 (no range), antaxial genual setae l'' 14–20, tibia 3–4, paraxial tibial setae l' located at $\frac{1}{3}-\frac{2}{5}$ from dorsal base, 3–4, very fine, tarsus 4–5, unguinal tarsal seta u' 2–3, solenidion ω 5 (no range), slightly curved, distally rounded or with a minute knob, empodium simple, bilaterally symmetrical, 4 (no range), 5-rayed. Leg II 17–18, femur 6–7, basiventral femoral setae bv 6-7, very fine, genu 2-3, antaxial genual setae l" 5-6, very fine, tibia 2-3, tarsus 4–5, unguinal tarsal seta u' 1–2, solenidion ω 5–7, slightly curved, distally rounded or with a minute knob, empodium simple, bilaterally symmetrical, 4-5, 5-rayed. Coxisternae I and II with granules; setae 1b 5 (no range), 9–10 apart, setae 1a 10–11, 7–8 apart, setae 2a 26-27, 14-15 apart, all very fine. Subcapitular plate with granules. In 60% of the examined specimens prosternal apodeme not well defined, in the others 9 (8-10). Coxigenital area with 3-4 microtuberculate semiannuli. Opisthosoma with 55-61 dorsal, 56-58 ventral annuli. Microtuberculate dorsal and ventral annuli. Posterior 4 dorsal annuli with only a few indistinct microtubercles dorsally, almost smooth. Ellipsoid microtubercles on rear annular margin, except for 4-5 ventral annuli of anal lobes, which are elongate and linear. Caudal lobe normal in size and shape. Setae c_2 14–20, on ventral annulus 9–10, 31-32 apart; setae d 34-39, on ventral annulus 18-21, 18-26 apart; setae e 3-5, on ventral annulus 29–35, 12–14 apart; setae f 10–13, on ventral annulus 52–58, or 4–5 from rear, 11-12 apart. Setae h2 33-40, 9-10 apart; setae h1 2 (no range), 5-7 apart. Genitalia 11–14, 15–16 wide, posterior ¼ with minute granules, setae 3a 5–6, 13–15 apart (Fig. 8).

NYMPH – White, vermiform, 135–157 (n = 5), 37 wide, 35–38 thick. Gnathosoma 14–16; dorsal palp genual setae *d* 3 (2–3), unbranched; chelicerae 13–14. Prodorsal shield 24–26, 31 wide, circular. Setae *sc* 25–27, 17 apart, pointing rear. Leg I 18–21, femur 5–7, basiventral femoral seta *bv* 2 (no range), genu 2–3, antaxial genual setae *l*" 13–15, tibia 2–3, paraxial tibial setae *l*' located at $\frac{1}{3}-\frac{2}{5}$ from dorsal base, 2–3, very fine, tarsus 3–4, solenidion ω 4–5, slightly curved, distally blunt, empodium simple, bilaterally symmetrical, 3–4, with 5 paired rays. Leg II 14–16, femur 5–6, basiventral femoral setae *bv* 3–5, very fine, genu 2–3, antaxial genual setae *l*" 4–5, very fine, tibia 1–2, tarsus 3–4, solenidion ω 5–6, slightly curved, distally blunt, empodium simple, bilaterally symmetrical, 3–4, with 5 paired rays. Setae *lb* 3–4, 10 apart, setae *la* 6–7, 7 apart, setae *2a* 13–16, 17 apart. Prosternal apodeme absent. Opisthosoma with 53–58 dorsal, 55–57 ventral semiannuli. Dorsal and ventral semiannuli with minute microtubercles. Setae *c2* 5–10, on annulus 10–

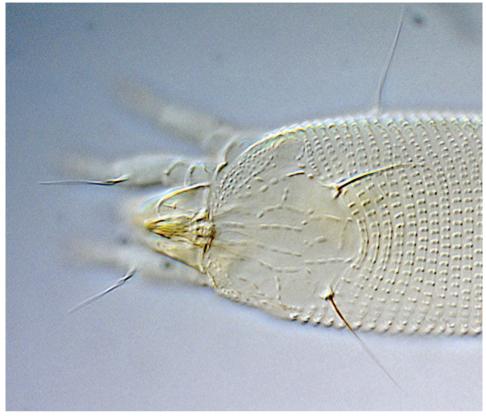


Fig. 8. Digital micrograph of *Aceria bambusae*, prodorsal shield and anterior opisthosoma of male. (Photo: Árpád Szabó)

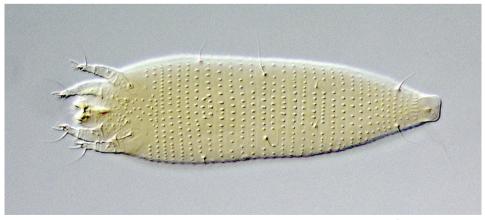


Fig. 9. Digital micrograph of Aceria bambusae, nymph in ventral view. (Photo: Árpád Szabó)

Acta Phytopathologica et Entomologica Hungarica

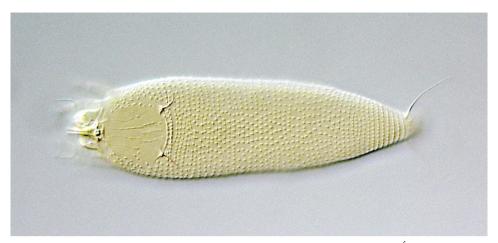


Fig. 10. Digital micrograph of Aceria bambusae, nymph in dorsal view. (Photo: Árpád Szabó)

11, 32 apart; setae d 28–29, on annulus 20–22, 25 apart; setae e 3–6, on annulus 30–33, 15 apart; setae f 10–13, on annulus 50–53, or 4–5 from rear, 14 apart. Setae h2 31–32, 9 apart; setae h1 1–2, 6 apart; setae 3a 2–4, 6 apart (Figs 9-10).

Host plant – Red-margin bamboo, *Phyllostachys rubromarginata* McClure (Poaceae: subfamily Bambusoideae: tribe Bambuseae). Red-margin bamboo is an ornamental bamboo species, native to temperate Central China, especially in Guangxi, Guangdong, Anhui, Zhejiang, Jiangsu and Henan provinces.

Relationship to the host – This mite was found in high number in the leaf sheaths of the host and caused no discernible symptoms.

Hungarian locality – Gödöllő (Pest county, Central Hungary), in the Botanical Garden of Szent István University, 241 m elev., 47°35'37,99" N, 19°21'59,95" E.

Material examined – The re-described and illustrated female among 5 females, 4 males and 7 nymphs, and one female of *Abacarus korosicsomai* Ripka on one slide, 2 July 2019, slide # 1465a, coll. Géza Ripka. Other materials: slide # 1465b was prepared from this material containing 8 females, 2 males and 2 nymphs from the leaf sheaths of the host; slide # 1469a from *Phyllostachys tianmuensis* one female, and 5 females and 3 males of *Rhyncaphytoptus longipalpis*, and one female of a *Cecidophyes* species, and one female of *Adventacarus turulae* Ripka, 2 July 2019, coll. Géza Ripka. Slides are in the corresponding author's collection and deposited in the National Food Chain Safety Office, Directorate of Plant Protection, Soil Conservation and Agri-environment, Budapest, Hungary.

Discussion

Out of 42 known bamboo-inhabiting eriophyoid mite species only four Aceria species are known, viz. Aceria bambusae ChannaBasavanna, 1966, Aceria gilloglii Keifer, 1963, Aceria havenensis (Keifer, 1979) and Aceria niitakayamensis Wang et Huang, 2011 described from Bambusa vulgaris in India, Sasa pygmaea in California, USA, Bambusa vulgaris in Florida, USA and Yushania niitakayamensis in Taiwan, resp. (ChannaBasavanna, 1966; Keifer, 1963, 1979; Elhalawany, 2015; Wang and Huang, 2011). According to Amrine and Stasny (1994) and Sukhareva (1994) Aceria havenensis (Keifer) is a junior synonym of Aceria bambusae ChannaBasavanna.

The re-described species shows similarities with the four nominal Aceria species living on bamboo hosts. The re-described species is close to Aceria gilloglii because it shares numerous features, particularly body shape, the number of dorsal and ventral annuli, the ornamentation of prodorsal shield and coxi-genital region, length of several setae, but can be differentiated by having single row of microtubercles on annuli, whereas A. gilloglii has double rows of microtubercles on each annulus laterally and dorsally. There are some morphometric differences between the examined Hungarian Aceria bambusae specimens and both the holotype of Aceria bambusae and the re-described specimens of A. bambusae given by Elhalawany (2015). We especially draw attention to the length of setae sc: which is 18-20 in holotype of A. bambusae, whereas 30-39 in specimens of A. bambusae collected in Hungary. There are two other marked differences: in the number of ridges on genital coverflap, which is 4-6 in holotype of A. bambusae, whereas 7-8 in the examined A. bambusae specimens; and the number of empodial rays, which is 7 in A. bambusae, versus 5 in the Hungarian A. bambusae specimens (Table 1). Nevertheless the examined specimens belong to A. bambusae because the vast majority of the morphometric data for female matches the original description by ChannaBasavanna.

Further eriophyoid species were collected from different bamboo species during this survey:

Abacarus korosicsomai Ripka from Phyllostachys fimbriligula T. H. Wen (Poaceae), Gödöllő (Pest county, Central Hungary), Botanical Garden of Szent István University, 13 October 2016 and 1 December 2016, coll. Enikő Kiss; from Phyllostachys rubromarginata McClure (Poaceae), Gödöllő (Pest county, Central Hungary), Botanical Garden of Szent István University, 24 August 2017, coll. Enikő Kiss; same locality, 2 July 2019, coll. Géza Ripka; from Phyllostachys angusta McClure (Poaceae), Debrecen (Hajdú-Bihar county, East Hungary), Diószegi Sámuel Botanical Garden, 19 October 2017, coll. András Neményi; from Phyllostachys edulis (Carrière) J. Houzeau (Poaceae), Szada (Pest county, Central Hungary), private garden, 11 September 2017, coll. András Neményi.

Adventacarus turulae Ripka from Phyllostachys edulis (Carrière) J. Houzeau (Poaceae), Szada (Pest county, Central Hungary), private garden, 11 September 2017, coll. András Neményi; from Phyllostachys aureosulcata McClure (Poaceae), Egerág (Baranya county, South Hungary), private garden, 11 September 2017, coll. András Neményi; from Phyllostachys nigra var. henonis (Mitford) Stapf ex Rendle (Poaceae), Egerág (Baranya county, South Hungary), private garden, 5 July 2019, coll. Jenő Kontschán.

Mucotergum nigrum Ripka from Phyllostachys meyeri McClure (Poaceae), Gödöllő (Pest county, Central Hungary), Botanical Garden of Szent István University, 19 September 2016, coll. Enikő Kiss; from Phyllostachys rubromarginata, Gödöllő (Pest county,

Table 1

Distinguishing characters between the females of A. bambusae, A. gilloglii, A. havenensis
and A. niitakayamensis $(n.d. = no data)$

Character	Aceria bambusae Hungarian specimens	Aceria bambusae holotype India	Aceria bambusae re-described Egyptian specimens	Aceria gilloglii holotype California, USA	Aceria havenensis holotype Florida, USA	Aceria niitaka- yamensis holotype Taiwan
Prodorsal shield shape	oval	broadly trun- cate in front	semicircu- lar, truncate anterior	subsemicir- cular	anteriorly truncate sub- semicircular	n.d.
Length of seta sc	30-39	20	18-20	29	11	20.8
Length of gnatho- soma	13–16	18	17–20	16	18	n.d.
Length of leg I	22–25	22	24–25	21	23	n.d.
Length of leg II	22-23	20	21-23	19	19	n.d.
Number of empo- dial rayes	5	7	7	5	6	5
Prosternal apo- deme	not well de- fined	present	absent	distinct	absent	present
Number of coxi-genital annuli	5–7	6	5-6	5	5	6
Number of ridges on genital cover- flap	7–8	4–6	4–6	8–10	5 (3 short and 2 long)	8
Number of opist- nosomal annuli	58-66	65–70	57–62	60	57	57
Length of seta c2	10-15	5	3-4	14	4	24.7
Length of seta d	48-59	55	33-40	51	36	34.7
Length of seta e	5-10	7	8-10	4,5	5	9.3
Length of seta f	13-15	19	18-21	11	7	20.6
Length of seta h2	43-55	45	33-40	n.d.	n.d.	n.d.
Length of seta 3a	6–9	6	5-6	12	7	10.7
Posterior dorsal opisthosomal and telosomal annuli	distinct elon- gate oval mi- crotubercles on dorsal an- nuli, tiny mi- crotubercles on last 4–5 dorsal annuli	obscure dor- sally and elon- gate ventrally	last 10 ven- tral annuli with linear microtuber- cles	double rows of microtu- bercles on most annuli	acuminate microtuber- cles on rear annular mar- gins	spine mi- crotuber- cles at posterior
Microhabitat; dam- age symptom	lives in the leaf sheaths, no damage	lives between stem and en- veloping leaf- sheath on terminal shoots, no damage	lives in the leaves sheath around stem	lives in the petiole bases, causing slight witch's broom	sheaths, may	vagrant or leaf under side, no damage

Central Hungary), Botanical Garden of Szent István University, 24 August 2017, coll. Enikő Kiss; from *Phyllostachys tianmuensis* McClure (Poaceae), Gödöllő (Pest county, Central Hungary), Botanical Garden of Szent István University, 24 August 2017, coll. Enikő Kiss.

Abacarus korosicsomai, Adventacarus turulae and Mucotergum nigrum are vagrant species, living on the underside of the leaves.

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Literature

- Amrine, J. W., Jr. and Manson, D. C. M. (1996): Preparation, mounting and descriptive study of eriophyoid mites. In: E. E. Lindquist, M. W. Sabelis and J. Bruin (eds): Eriophyoid Mites – their Biology, Natural Enemies and Control. World Crop Pests, 6. Elsevier Scientific Publishing, Amsterdam, pp. 383–396.
- Amrine, J. W., Jr. and Stasny, T. A. (1994): Catalog of the Eriophyoidea (Acarina: Prostigmata) of the World. Indira Publishing House, West Bloomfield, pp. ix + 798.
- Amrine, J. W. Jr., Stasny, T. A. H. and Flechtmann, C. H. W. (2003): Revised Keys to World Genera of Eriophyoidea (Acari: Prostigmata). Indira Publishing House, West Bloomfield, iv + 244 p.
- Baker, E. W., Kono, T., Amrine, J. W., Jr., Delfinado-Baker, M. and Stasny, T. A. (1996): Eriophyoid mites of the United States. Indira Publishing House, West Bloomfield, USA. pp. ix + 394.
- ChannaBasavanna, G. P. (1966): A contribution to the knowledge of Indian eriophyid mites (Eriophyoidea: Trombidiformes: Acarina). University of Agricultural Sciences, Hebbal, Bangalore, India, ix + 154 p + 31 pls.
- Chetverikov, P. E. and Craemer, C. (2015): Gnathosomal interlocking apparatus and remarks on functional morphology of frontal lobes of eriophyoid mites (Acariformes, Eriophyoidea). Experimental and Applied Acarology, 66, 187–202. http://dx.doi.org/10.1007/s10493-015-9906-3
- Davis, R., Flechtmann, C. H. W., Boczek, J. H. and Barké, H. E. (1982): Catalogue of Eriophyid Mites (Acari: Eriophyoidea). Warsaw Agricultural University Press, Warsaw, 254 p.
- Elhalawany, A. (2015): Description of one new species and two first records of eriophyid mites (Prostigmata: Eriophyidae) on grasses in Egypt. Egypt J. Agric. Res., 93, 41–59.
- Huang, K.-W. (2001): Eriophyoid mites of Taiwan: description of eighty-six species from the Tengchih area (Acarina: Eriophyoidea). Bulletin of National Museum of Natural Science, 14, 1–84.
- Keifer, H. H. (1963): Eriophyid Studies B-9. Bureau of Entomology, California Department of Agriculture, 1–20.
- Keifer, H. H. (1975): Eriophyoidea Nalepa. In: L. R. Jeppson, H. H. Keifer and E. W. Baker (eds): Mites Injurious to Economic Plants. University of California Press, Berkeley, Los Angeles, London. pp. 327–533.
- Keifer, H. H. (1979): Eriophyid Studies C-16. Entomology Research Division, Agr. Res. Svc. USDA, 1–24.
- Lin, J.-Z., Zhang, Z.-Q., Zhang, Y.-X., Liu, Q.-Y. and Ji, J. (2000): Checklist of mites from moso bamboo in Fujian, China. Systematic and Applied Acarology Special Publications, 4, 81–92.
- Lindquist, E. E. (1996): External anatomy and notation of structures. In: E. E. Lindquist, M. W. Sabelis and J. Bruin (eds): Eriophyoid Mites – their Biology, Natural Enemies and Control. World Crop Pests, 6. Elsevier Scientific Publishing, Amsterdam, pp. 3–31.
- Ohrnberger, D. (2002): The Bamboos of the World: Annotated Nomenclature and Literature of the Species and the Higher and Lower Taxa. Elsevier, 596 pp (accessed 15 October 2018).

- Oldfield, G. N. (1996): Diversity and host plant specificity. In: E. E. Lindquist, M. W. Sabelis and J. Bruin (eds): Eriophyoid Mites – their Biology, Natural Enemies and Control. World Crop Pests, 6. Elsevier Scientific Publishing, Amsterdam, pp. 199–216.
- http://dx.doi.org/10.106/S1572-4379(96)80011-X
- Ripka, G. (2011): A new genus, Adventacarus and new Abacarus species from Hungary (Acari: Prostigmata: Eriophyoidea). Acta Phytopathol. et Entomol. Hung. 46, 139–149.
- Ripka, G., Kontschán, J. and Neményi, A. (2015): A new genus and species of eriophyoid mites (Acari: Eriophyoidea: Diptilomiopidae) on *Phyllostachys iridescens* (Poaceae) from Hungary. Acta Zoologica Academiae Scientiarum Hungaricae, 61, 47–56.
- Sukhareva, S. I. (1994): The four-legged mites (Acariformes, Tetrapodili) on the bamboos in Georgia. Vestnik Sankt-Peterburgskogo Universiteta 3, 33–38. [in Russian with English summary]
- The Plant List (2013): Version 1.1. Published on the Internet; http://www.theplantlist.org/ (accessed 11th September 2019).
- Wang, C.-F. and Huang, K.-W. (2011): Taxonomy of eriophyoid mites (Acari: Trombidiformes) infesting Yushania niitakayamensis (Poaceae: Bambuseae) in Taiwan. Formosan Entomologist, 31, 249–280.
- Xue, X.-F., Song, Z.-W., Amrine, J. W., Jr. and Hong, X.-Y. (2006): Eriophyid mites (Acari: Eriophyoidea) on bamboo from China, with descriptions of three new species from the Qinling Mountains. Ann. Entomol. Soc. Am., 99, 1057–1063.

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