

Quantifying temporal variability in the metacommunity structure of stream fishes: the influence of non-native species and environmental drivers

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Abstract

Most studies characterize metacommunities based on a single snapshot of the spatial structure, which may be inadequate for taxa with high migratory behaviour (e.g., fish). Here, we applied elements of metacommunity structure to examine variations in the spatial distributions of stream fishes over time and to explore possible structuring mechanisms. Although the major environmental gradients influencing species distributions remained largely the same in time, the best-fit pattern of metacommunity structure varied according to sampling occasion and whether or not we included non-native species in the analyses. Quasi-Clementsian and Clementsian structures were the predominant best-fit structures, indicating the importance of species turnover among sites and the existence of more or less discrete community boundaries. The environmental gradient most correlated with metacommunity structure was defined by altitude, area of artificial ponds in the lowlands, and dissolved oxygen content. Our results suggest that the best-fit metacommunity structure of the native species can change in time in this catchment due to seasonal changes in distribution patterns. However, the distribution of non-native species throughout the landscape homogenizes the temporal variability in metacommunity structure of native species. Further studies are necessary from other regions to examine best-fit-metacommunity structures of stream fishes within relatively short environmental gradients.

Keywords: metacommunities, elements of metacommunity structure, streams, fish assemblages, temporal variation, non-native species

Introduction

The metacommunity concept substantially advanced ecological research by providing an opportunity to examine how spatial dynamics and local niche-based interactions influence community structure and how populations of different species are distributed across the landscape (Leibold *et al.*, 2004; Holyoak *et al.* 2005). The shift in focus from local to landscape or regional scale patterns was followed by the development of evaluation frameworks about between site species distributions (i.e., site-by-species distributions). In fact, several different constructs have been proposed by ecologists to identify patterns in species distributions (e.g., nested subsets or checkerboard distributions). However, these models have been tested separately and, in many cases, even without determining whether the spatial distribution was significantly different than random (Leibold & Mikkelsen, 2002). Two of the earliest models reflected differences in how species respond to environmental gradients; Clementsian distributions arise when groups of species show similar responses to environmental gradients and therefore can be classified into well defined, distinctive community types and Gleasonian distributions reflect individualistic responses that yield a continuum of gradually changing composition without clumping. Evenly-spaced gradients can occur in systems with intense interspecific competition in which trade-offs in competitive ability result in spatial distributions with evenly dispersed populations (Tilman, 1982). Alternatively, intense competition may manifest as mutually exclusive spatial distributions, resulting in checkerboard patterns (Diamond, 1975). Metacommunities with nested structure are associated with predictable patterns of species loss in which species-poor communities are proper subsets of more speciose communities; the resulting pattern of species loss is based often on species-specific characteristics such as dispersal ability, habitat specialization, tolerance to abiotic conditions (Patterson & Atmar, 1986; Ulrich *et al.*, 2012).

The elements of metacommunity structure (EMS) approach of Leibold & Mikkelsen

(2002) is useful when trying to characterize the overall pattern of species distributions from a regional perspective (e.g., Clementsian, Gleasonian, nested distributions) by assessing aspects of coherence, species turnover, and boundary clumping. These components (Fig. 1, see methods for more detail) coupled with the additions of Presley *et al.* (2010) identifies patterns in the spatial distribution of populations across the region and allows for the exploration of relationships between species distributions and environmental gradients. Previous pattern identification methods mostly tested for the existence of a single spatial distribution (e.g., nested or checkerboard patterns), whereas the EMS approach of Leibold & Mikkelsen (2002) tests for multiple distributions simultaneously by discriminating among a set of idealized patterns and their Quasi-structures in a single set of analyses (Presley *et al.*, 2010).

Examining spatial and temporal patterns in how species are spatially distributed using EMS can be fruitful for our generalizations about the diversity and relative frequency of community patterns in nature, especially in regards to the effects of human perturbation (e.g., habitat modifications, climate change, introduction of non-native species). From an applied perspective, a comprehensive understanding of how species are spatially distributed within and among fragmented habitats (i.e., metacommunity structure), and how that structure changes through time is required to establish effective conservation policy. For example, a nested structure may permit the prioritization of just a small number of the richest sites, whereas a Clementsian or Gleasonian structure require devoting conservation efforts to several different sites, not necessarily the richest ones (Baselga, 2010). The identification of idealized distribution patterns is also at the heart of applied stream ecology because management usually requires well-defined assemblage types for conservation purposes (Aarts & Nienhuis, 2003; Heino *et al.*, 2003; Hermoso & Linke, 2012).

Freshwater assemblages have been associated with a variety of non-random species distribution patterns (Jackson *et al.*, 2001; Heino, 2011). Several studies examined whether

they show discrete assemblage types or a continuum in individualistic species replacement to environmental gradients along the longitudinal profile of streams and rivers (Matthews, 1998; Statzner & Higler, 2006; Lasne *et al.* 2007). Nested distribution patterns due to selective extinction and/or colonization events or changes in the diversity of habitats have also been identified along the longitudinal continuum (Taylor, 1997; Erős & Grossman, 2005). Although much is known about the spatial distribution of stream assemblages in regards to longitudinal zonation within a river (Aarts & Nienhuis, 2003; Ibarra *et al.*, 2005; Statzner & Higler, 2006;), few studies have tested the idealized spatial distribution in regards to a network of smaller streams and shorter environmental gradients (but see Heino, 2005 for a test on stream macroinvertebrates).

The lack of studies that focus on the temporal variability in metacommunity structure in stream systems is surprising given that streams are dynamic ecosystems both spatially and temporally (Resh *et al.*, 1988; Lake, 2000). In terms of temporal, or seasonal variation, stream fish often migrate between feeding habitats, spawning grounds, and refugia (Schlosser, 1991). The longitudinal movement up and downstream should alter local patterns of diversity and, consequently, metacommunity structure. Temporal changes in the water regime can also substantially influence the dynamics of species occurrences in streams (Resh *et al.*, 1988; Grossman *et al.*, 2010), which could alter the spatial structuring of populations across the stream network. Additionally, patterns of biodiversity are increasingly affected by the introduction of non-native species that can potentially impact native populations by altering habitat, increasing predation pressure and/or interspecific competition (e.g., for food or shelter), and hybridizing with native species (Fridley *et al.*, 2007). The impact of non-native species on spatial and temporal patterns of metacommunity structure, however, remains largely unknown.

The overall objective of this study was to examine temporal variability in metacommunity structure of stream-fishes in the catchment of Lake Balaton, Hungary. First, we wanted to characterize the spatial relationship of species across the landscape by determining which pattern of metacommunity structure best fit the data. Because we wanted to disentangle the effects of non-native species on metacommunity structure, we analysed the data at two assemblage levels (i) that of the entire assemblage and (ii) the native assemblage (excluding non-native species from the analysis). Second, we wanted to examine temporal variation in the spatial distribution of species and its impact on metacommunity structure. Third, we wanted to determine which environmental variables were most likely responsible for producing the observed metacommunity structure.

Material and Methods

Study area and stream surveys

We sampled a total of 40 sites across 22 wadable streams in the catchment of Lake Balaton, Hungary (5775 km²) from Spring 2008 through Autumn 2010. A map of the stream network and a complete description of the study area can be found in the work of Sály *et al.* (2011), but will be reiterated here, briefly. The dominant land use type in the catchment is agricultural (mainly arable lands, vineyards, and orchards) and comprises about 40% of the total area. Deciduous forests (28%) as well as pastures and grasslands (12%) are the other characteristic land cover types. The proportion of stagnant water bodies, watercourses, and wetlands is in combination 14%, and that of the human inhabited area is 6%. The highland and lowland streams in the catchment provide heterogeneous environmental conditions for fish, ranging from well-shaded stream sections to more open, weed or macrophyte dominated channels. The dominant substrates are typically gravel or silt-sand. Streams are usually less

than 5 m wide, and they are fairly modified with dikes along the banks, especially in the most lowland sections. Ponds used for aquaculture and recreational fishing can be also found in the catchment. Some of these artificial ponds maintain dense populations of non-native species, which may regularly escape into the streams.

Fish surveys

The 40 sites were surveyed three times in each year (spring, summer, and autumn) with a standardized sampling protocol, resulting in a total of 360 samples ($40 \text{ sites} \times 3 \text{ years} \times 3 \text{ seasonal samples}$). The sampling sites were randomly selected from potential candidate sites, which were selected after preliminary investigation in 2006 and 2007 to be representative of the segment (i.e., stretch with similar instream habitat features and riparian characteristics) itself based on land use and in-stream habitat characteristics, and accessibility constraints. At each site, we surveyed a 150 m long reach by wading, single pass electrofishing using a backpack electrofisher (IG200/2B, PDC, 50–100 Hz, 350–650 V, max. 10 kW; Hans Grassl GmbH, Germany). This amount of sampling effort was found to yield representative samples of fish assemblages in this study area for between-site assemblage comparisons (Sály *et al.*, 2009) and is also comparable with those routinely used elsewhere for the sampling of fish in wadeable streams (Magalhães *et al.*, 2002; Schmutz *et al.*, 2007; Hughes & Peck, 2008). Fish were stored in aerated containers filled with water while fishing, then identified to species level, counted, and released back to the stream.

Environmental variables

We measured a number of local environmental and landscape-level variables (Appendix I) that have been shown to structure fish assemblages in this catchment (Sály *et al.*, 2011) and elsewhere (e.g., Wang *et al.*, 2003; Hoeinghaus *et al.*, 2007). At each sampling site, 6–15 transects (depending on the complexity of the habitat) were placed perpendicular to

the main channel of the stream to characterize physical features of the environment. Wetted width was measured once along each transect, whereas water depth and current velocity (at 60% depth) were measured at 3–6 (varied according to the width) equally spaced points along each transect. Visual estimates of percentage substratum cover were made at every transect point as well (see Appendix I for categories). Percentage substratum data of the transect points were later pooled and overall percentage of substrate categories were calculated for each site. Conductivity, dissolved oxygen content, and pH were measured with an OAKTON Waterproof PCD 650 portable handheld meter before fish sampling, and the content of nitrogen forms (i.e., nitrite, nitrate, ammonium) and phosphate were measured using field kits (Visocolor ECO, Macherey-Nagel GmbH & Co. KG., Germany). Coefficient of variation (CV) of depth, velocity, and width data were also calculated to characterize temporal variability in flow regime. Land cover variables were quantified based on their proportion (%) in the catchment area above each sampling site. Digital land cover information was obtained from the CORINE Land Cover 2000 database (CLC2000; European Environmental Agency, <http://www.eea.europa.eu>) (see Appendix I). We quantified the variable “pond area” as the total area of ponds located within the upstream catchment of each sample site. The longitudinal position of each sample site was measured as the stream-line distance from each site to its upstream source and to the upstream mouth of the stream at a scale of 1:80 000 using the National GIS Database of Hungary (Institute of Geodesy, Cartography and Remote Sensing, Hungary). The variables altitude, stream-line distances, and land cover descriptors were measured only once, whereas instream physical and chemical variables were measured during each sampling occasion.

Data analysis

Following Leibold & Mikkelsen (2002) and Presley *et al.* (2010), we analysed aspects of coherence, species turnover, and boundary clumping (EMS analysis) to characterize the seasonal metacommunity structure of stream-fish assemblages over time. We used reciprocal averaging (also called correspondence analysis, CoA), an unconstrained ordination method, to arrange the sampling sites so that sites with similar species composition are adjacent and to arrange the order of species so that species with similar spatial distributional range (i.e., spatial occurrence patterns) are closer together. One of the advantages of using this ordination technique is that one does not have to *a priori* specify which environmental variables to include because the first axis is based on maximum association between site scores and species scores (Leibold & Mikkelsen, 2002). That is, the primary axis represents the strongest relationship between species composition within a site and spatial distribution of species among sites. Any environmental variables significantly correlated with that primary axis of variation, or latent environmental gradient, would obviously be an important factor in determining a species' distributional pattern

After rearranging the data matrix, we tested for coherence in species occurrences along the environmental gradient defined by the first ordination axis (CoA1). We counted the number of embedded absences (gaps in species distributions) and compared that number to a null distribution created from a null model with 1000 iterations. The null model constrained simulated species richness of each site to equal empirical richness, with equiprobable occurrences for each species (Presley *et al.*, 2010). If the number of embedded absences was significantly different from random with more embedded absences than that expected by chance, we considered coherence to be negative. This suggests that trade-offs in competitive ability between species may manifest as a “checkerboard” like spatial distribution (Diamond, 1975). If the number of embedded absences was significantly less than that expected by chance, we considered the coherence within the metacommunity to be positive. Positive

coherence indicates that a majority of the species are responding similarly to a latent environmental gradient defined by the primary axis of variation (Leibold & Mikkelsen, 2002).

For metacommunities that were positively coherent, an additional aspect (species turnover) was considered. Species turnover was measured as the number of times one species replaced another between two sites (i.e., number of replacements) for each possible pair of species and for each possible pair of sites. A replacement between two species (e.g., species A and B) occurs when the range of species A extends beyond that of species B at one end of the gradient and the range of B extends beyond that of A at the other end of the gradient. The observed number of replacements in a metacommunity is compared to a null distribution that randomly shifts entire ranges of species (Leibold & Mikkelsen, 2002). Significantly low (negative) turnover is consistent with nested distributions, and significantly high (positive) turnover is consistent with Gleasonian, Clementsian, or evenly spaced distributions, requiring further analysis of boundary clumping to distinguish among them. Boundary clumping quantifies the geographic distribution of all species, determining whether the metacommunity is clumped, evenly-spaced, or random with respect to the spatial distribution of species across the region (Leibold & Mikkelsen, 2002). We quantified the degree of boundary clumping using Morisita's index, which is typically viewed as a statistical measure of dispersion of individuals in a population (Morisita, 1971). However, this index can be extrapolated to include the dispersion of species in a metacommunity (Leibold & Mikkelsen, 2002). Index values significantly greater than 1 indicated substantial boundary clumping (i.e., Clementsian distribution), values significantly less than 1 indicated evenly spaced boundaries, and values not significantly different from 1 indicated randomly distributed species boundaries (i.e., Gleasonian distribution).

We performed the EMS analysis for each seasonal survey separately (i.e., nine occasions). We conducted the analyses at the entire assemblage (which included both native

and non-native species) and the native assemblage (containing only native species) levels for each seasonal dataset. This resulted in a total of 18 EMS analyses (9 occasions \times 2 assemblage levels). Rare species (i.e. species representing $< 0.1\%$ relative abundance and/or species that occurred only at one site) were removed prior to analyses to reduce their disproportional effect on the results (Legendre & Legendre, 1998; Presley *et al.*, 2009; Keith *et al.*, 2011). Analyses of coherence, species turnover, and boundary clumping (i.e., elements of metacommunity structure, EMS) were conducted with algorithms written in Matlab 7.5 (Presley *et al.*, 2010; available at <http://faculty.tarleton.edu/higgins/metacommunity-structure.html>).

Modelling metacommunity structure with environmental data

We used multiple linear regression to assess the importance of environmental variables in influencing metacommunity structure, with the first corresponding axis serving as the dependent (i.e., response) variable (see e.g., Presley & Willig, 2010; Keith *et al.*, 2011; Willig *et al.*, 2011 for a similar approach). We performed the analyses separately for each season and for the entire and the native assemblage levels, which yielded 18 multiple regression analyses (9 seasons \times 2 assemblage levels). Before data analyses, the environmental variables were transformed depending on their scale of measurement to improve normality and reduce heteroscedasticity (see Appendix I). Strongly collinear variables ($r > 0.7$) were omitted from further analyses. The explanatory variables were then first screened via a forward selection procedure with Monte Carlo randomization tests (10 000 runs) to obtain a reduced set of significant variables (variables retained at $p < 0.05$) for the final regression models (Blanchet *et al.*, 2008). Regression models were fitted on the standardized dependent and independent variables (i.e., variables with 0 mean and 1 standard

deviation) to yield standardized partial regression coefficients (i.e., beta coefficients) from the models (Quinn & Keough, 2002). Standardized partial regression coefficients are directly comparable with each other, and indicate the relative importance of the independent variables in explaining the variability of the dependent variable.

Results

Altogether we collected 39 species and 71 291 specimens during the three year study (Appendix II). Of the 39 species 15 were regarded as rare species (for definition see methods) and were omitted from the analyses. Hence 24 species of which 5 were non-native were retained for further analyses. EMS revealed the existence of different patterns of metacommunity structure depending on time period and the assemblage level (entire assemblages or non-natives excluded). At the entire assemblage level (Table 1), species were distributed in a pattern consistent with a Gleasonian structure in the spring of 2008. Beginning in the summer of 2008, the metacommunity structure shifted to one that was more consistent with a Quasi-Clementsian pattern. In fact, the spatial distributions of populations were Quasi-Clementsian for eight of the nine sampling seasons, suggesting metacommunity structure changed little over time (Fig. 2.). However, the variance explained by the first CoA axis was relatively low in each occasion and varied between 17.7% and 24.0%. Exclusion of non-natives influenced the results markedly (Table 2). After removing non-native species from the analyses, we observed Clementsian, Quasi-Clementsian, Gleasonian, and random metacommunity structures. Although there was no clear trend in changes in metacommunity structure over time, the only random structures were in Autumn 2009 and Autumn 2010. Similar to the analyses at the entire assemblage level, the variance explained by the first CoA axis was relatively low in each season and varied between 18.2% and 25.9%.

Regression analyses indicated that the gradient in fish assemblage composition (i.e., CoA1) was well modelled with environmental variables (Table 3 and 4). Adjusted R^2 -values varied between 0.479 and 0.774 at the entire assemblage level analyses (Table 3). The main environmental variables selected by the modelling procedure were not the same for each season (i.e., occasion). The most important ones were as follows: altitude, pond area, and oxygen content. Exclusion of non-natives from the analyses did not influence these results (Table 4). Adjusted R^2 -values increased slightly after removing non-native species and ranged from 0.489 to 0.802, but the most influential variables remained the same (altitude, pond area, and oxygen content).

Discussion

The metacommunity structure of stream fishes in the catchment of Lake Balaton changed temporally and differed when non-native species were included in the analyses. At the entire assemblage level, the metacommunity structure was consistent with a Quasi-Clementsian structure for every season except Spring 2008 in which case a Gleasonian distribution best fit the data. On the contrary, a variety of metacommunity structures, including even random distribution pattern characterized the native assemblage level dataset, although Quasi-Clementsian and Clementsian structures were dominant. These results show that species distributions were generally coherent, which indicates that species responded similarly to an environmental gradient. In our study, the environmental gradient that correlated the most with the primary axis scores of the CoA was predominantly defined by altitude, pond area, and dissolved oxygen content. Because the temporal extent of our study covered only three years, we discount water-basin level extinctions during the three years as being influential in these changes (Erős *et al.* unpublished results). Instead, we hypothesize that the temporal changes in metacommunity structure were attributable to changes in within

and among site occupancy patterns of fishes driven largely by migration dynamics and their responses to the environmental gradients.

Metacommunities with positive coherence and non-significant turnover have a non-random (i.e., quasi) structure (Fig. 1.). These Quasi-structures can emerge due to weaker structuring forces than those effecting characteristic structures (e.g., Clementsian, Gleasonian) in which turnover is significant (Presley *et al.*, 2010). The most frequently occurring metacommunity structure and the only quasi-structure we observed was Quasi-Clementsian. It was indicated by positive coherence, non-significant positive turnover, and positive boundary clumping. This structure emerged because the distribution of many species spanned the entire environmental gradient, whereas other species were restricted to one end or the other of the CoA primary axis. For example, the minnow and the stone loach always occupied only one half of the gradient, whereas the mud loach, the perch, and some rare species typically occupied the other side of the gradient (Fig. 2.). Both the stone loach and the minnow are characteristic species of higher altitude streams, whereas the many rare species occurring in the other side of the gradient are typical of lowland streams that have a more diverse fish assemblage composition than highland ones (Erős, 2007). On the contrary, the most common fishes, such as the chub, bitterling, gudgeon, and roach, were relatively abundant along the whole gradient. These results suggest that these fish are responding to an environmental gradient, but some species groups are responding differently to variation along that gradient.

In a recent study on stream fish assemblages, Hermoso & Linke (2012) found that assemblage level predictions from a top-down (i.e., environmental classification based) approach were no different than random expectations; in fact, the bottom-up models also performed poorly as a result of high levels of within and among site variation. The larger the variation in composition the more likely the metacommunity will have a “Quasi” component. In this respect, our study supports this general conclusion in that species responded to the

environmental gradient, but did not have enough turnover in species composition along that gradient to be statistically different than random, a result which was further supported by the low explained variance in the first axis of the correspondence analysis. However, the significant clumping is indicative of a Clementsian pattern and is consistent with differences in species composition between upland and lowland regions. In our study, altitude and pond area proved to be the most stable variables with which fish assemblage composition (CoA1 axis scores) correlated in most occasions. Artificial ponds (reservoirs, fish ponds) are most frequent in the lowland areas in this catchment (Sály *et al.*, 2011; Erős *et al.*, 2012), and therefore it is not surprising that the composition of the assemblages in this lowlands showed opposite reaction to altitude. Therefore, this study confirms previous findings in which Erős *et al.* (2012) applied a different analytical procedure (variance partitioning in redundancy analysis) and highlighted the fact that relatively small variations in altitude can contribute to changes in fish assemblage composition.

Based on the studies that have examined EMS so far, a variety of idealized spatial patterns have been identified (e.g., Presley *et al.*, 2009; Presley & Willig, 2010; Hoverman *et al.*, 2011; López-Gonzalez *et al.*, 2012). However, to our knowledge only one study examined coherence, species turnover, and boundary clumping (i.e., EMS) as a means of characterizing metacommunity structure of stream organisms; results indicated that the spatial distributions of stream midges were most consistent with Gleasonian and nested patterns distributional pattern (Heino, 2005). Much of the emphasis on EMS has been spatial in nature with little focus on temporal variations. Of the few exceptions, Keith *et al.* (2011) observed no change in Clementsian structure of vascular plants in woodland patches approximately 70 years apart, despite a significant loss in beta diversity through taxonomic homogenization. For terrestrial gastropods of Puerto Rico, the spatial structure was least nested, or more random, immediately following a hurricane disturbance, becoming more nested as the forest recovered

during secondary succession reducing spatial heterogeneity (Bloch *et al.*, 2007). Nested distribution patterns have been found for both stream macroinvertebrates (Malmqvist & Hoffsten, 2000; Heino, 2011) and fishes (Taylor & Warren, 2001; Erős & Grossman, 2005). We did not find nested metacommunity structure in any occasion, although differences in species richness among sites were clearly important in this metacommunity. However, it is important to emphasize that EMS finds the best-fit pattern of metacommunity structure from a set of idealized patterns. In this catchment, positive turnover along the environmental gradient was a stronger structuring force than factors that cause richness differences among sites (e.g., changes in habitat complexity from source to mouth, Erős & Grossman, 2005).

We observed temporal changes in metacommunity structure at the native assemblage level, but the structure remained relatively stable at the entire assemblage level. Removal of the non-native species allowed three of the Quasi-Clementsian distributions observed at the entire-assemblage level to become statistically significant in which case the overall spatial distribution was changed to Clementsian. However, in two other occasions the removal of non-natives yielded random pattern; in one occasion Gleasonian structure was found. These results suggest that the dominant Clementsian and Quasi-Clementsian metacommunity structure of the native species can change in time in this catchment due to temporally variable species distribution patterns that may be due to migration dynamics of some species between sites and/or to the effect of seasonally differing environmental factors on species distributions. However, the distribution of non-native species in the landscape homogenizes this temporal variability in metacommunity structure. This is the first time, to our knowledge, that shows that non-native species can homogenize temporal patterns in metacommunity structure. Our study thus highlights that distribution pattern of non-natives should be separately evaluated from those of native species when seeking for the best-fit metacommunity structure in landscapes where non-natives are present.

In conclusion, mechanism-based (Erős *et al*, 2012) and our pattern-based approaches both show moderate responses (here turnover) of fish assemblages to environmental gradients in this landscape. Although we found Quasi-Clementsian structure to be the most dominant metacommunity structure, our analyses indicated temporal variability in the best-fit-metacommunity structure depending on which assemblage level was used in the analyses. The difference in species composition and associated distributions between highland and lowland streams likely accounts for a majority of the clustering of species, a hypothesis supported by the fact that altitude was one of the primary environmental factors. Since compositional changes of fishes along long environmental gradients are relatively well known, we believe that further studies are necessary from other regions to examine best-fit-metacommunity structures of stream fishes within relatively short environmental gradients. This could help to better understand the predictability of fish assemblages to subtle changes in environmental heterogeneity and the dominant ecological mechanisms.

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Table 1. Results of the elements of metacommunity structure (EMS) analysis at the entire assemblage level (i.e., both native and non-native species included). Abs: the number of embedded absences in the ordinated (First axis of a correspondence analysis) matrix. Re: Species replacements. M: Morisita's index. Mean and standard deviation (SD) values show the values calculated based on 1000 iterations of a null matrix (see methods for further details).

Date	Abs	Coherence			Species turnover			Boundary clumping		Coherence	Turnover	Clumping	Best fit Structure
		P	Mean	SD	Re	P	Mean	SD	M	P			
Sp-2008	333	<0.001	418.5	24.5	13320	0.013	8069.2	2121.6	0.98	0.504	Positive	Positive	NS
Su-2008	292	0.002	385.8	30.4	9215	0.826	8774.7	1997.0	1.85	<0.001	Positive	NS+	Positive
Au-2008	243	<0.001	381.5	31.8	11180	0.261	9118.1	1833.9	1.60	0.006	Positive	NS+	Positive
Sp-2009	267	<0.001	416.7	26.3	13609	0.169	1069.7	2115.9	2.39	<0.001	Positive	NS+	Positive
Su-2009	271	<0.001	403.3	32.7	14541	0.110	10738.1	2379.2	1.59	0.008	Positive	NS+	Positive
Au-2009	344	0.0387	403.4	28.7	10109	0.286	7983.9	1991.8	1.52	0.013	Positive	NS+	Positive
Sp-2010	287	<0.001	424.1	36.9	14155	0.386	12090.2	2382.4	1.77	0.003	Positive	NS+	Positive
Su-2010	280	<0.001	415.9	33.9	17707	0.182	13937.4	2827.1	2.06	<0.001	Positive	NS+	Positive
Au-2010	290	0.0029	396.4	35.7	12230	0.100	9104	1897.6	1.65	0.002	Positive	NS+	Positive

Sp, spring; Su, summer; Au, autumn;

Table 2. Results of the elements of metacommunity structure (EMS) analysis at the native assemblage level (i.e., non-native species excluded). Abs: the number of embedded absences in the ordinated (First axis of a correspondence analysis) matrix. Re: Species replacements. M: Morisita's index. Mean and standard deviation (SD) values show the values calculated based on 1000 iterations of a null matrix (see Methods for more details.)

Date	Abs	Coherence			Re	Species turnover			Boundary clumping		Coherence	Turnover	Clumping	Best fit Structure
		P	Mean	SD		P	Mean	SD	M	P				
Sp-2008	238	0.0016	308.2	22.3	9316	0.0195	5509.4	1629.1	1.0053	0.4625	Positive	Positive	NS	Gleasonian
Su-2008	202	0.0317	268.5	30.9	6435	0.1343	4456.3	1321.6	1.9432	0.0017	Positive	NS+	Positive	Quasi-Clementsian
Au-2008	156	<0.001	267.7	29.5	8343	0.0519	5693.9	1362.7	1.7992	0.0054	Positive	NS+	Positive	Quasi-Clementsian
Sp-2009	201	<0.001	301.7	27.5	11363	0.006	6871.3	1635.7	2.2984	<0.001	Positive	Positive	Positive	Clementsian
Su-2009	192	<0.001	298.2	29.1	8949	0.0307	5806.1	1454.8	1.5114	0.0398	Positive	Positive	Positive	Clementsian
Au-2009	240	0.1483	289.9	34.5	6159	0.165	4416.1	1255.2	1.471	0.0611	NS	NS+	NS	Random
Sp-2010	209	0.0028	307.6	33	10209	0.1187	7542	1709.1	1.9218	0.0041	Positive	NS+	Positive	Quasi-Clementsian
Su-2010	217	0.0035	304.5	29.9	12881	0.0051	7639.9	1871.9	1.6598	0.0027	Positive	Positive	Positive	Clementsian
Au-2010	224	0.0762	278.8	30.9	8020	0.0134	4801.5	1301.3	2.2452	<0.001	NS	Positive	Positive	Random

Sp, spring; Su, summer; Au, autumn;

Table 3. Summary of results of the regression analyses between the environmental variables and the main fish assemblage gradient (i.e., first CoA axis) at the entire assemblage level (i.e. both native and non-native species included). Note, that we calculated standardized regression coefficients (i.e., beta coefficients [Quinn & Keough 2002]) from the *a priori* zero mean and one standard deviation standardized variables, in order that the importance of each variable could be directly compared.

Date	Model	F	P	R ² adj
Sp-2008	$y = 0.569 \text{ (altitude)} - 0.396 \text{ (pa)} + 0.223 \text{ (dissolved oxygen content)}$	45.41	<0.001	0.774
Su-2008	$y = -0.581 \text{ (altitude)} + 0.397 \text{ (pa)}$	40.40	<0.001	0.669
Au-2008	$y = -0.702 \text{ (altitude)}$	36.89	<0.001	0.479
Sp-2009	$y = -0.518 \text{ (altitude)} + 0.471 \text{ (pa)}$	41.41	<0.001	0.675
Su-2009	$y = -0.516 \text{ (altitude)} + 0.325 \text{ (pa)} - 0.296 \text{ (dissolved oxygen content)}$	25.80	<0.001	0.656
Au-2009	$y = -0.764 \text{ (altitude)}$	53.33	<0.001	0.573
Sp-2010	$y = -0.492 \text{ (altitude)} + 0.442 \text{ (pa)}$	29.97	<0.001	0.598
Su-2010	$y = -0.557 \text{ (altitude)} + 0.322 \text{ (pa)} - 0.288 \text{ (dissolved oxygen content)}$	32.51	<0.001	0.708
Au-2010	$y = 0.489 \text{ (altitude)} - 0.179 \text{ (CV depth)} + 0.366 \text{ (silt)} - 0.279 \text{ (% wetland)}$	16.64	<0.001	0.616

Sp, spring; Su, summer; Au, autumn; pa, pond area.

Table 4. Summary of results of the regression analyses between the environmental variables and the main fish assemblage gradient (i.e., first CoA axis) at the native assemblage level (i.e., non-native species excluded). Note, that we calculated standardized partial regression coefficients (i.e., beta coefficients [Quinn & Keough 2002]) from the *a priori* zero mean and one standard deviation standardized variables, in order that the importance of each variable could be directly compared.

Date	Model	F	P	R ² adj
Sp-2008	$y = +0.521 \text{ (altitude)} - 0.473 \text{ (pa)} + 0.275 \text{ (dissolved oxygen content)} + 0.249 \text{ (\% inhabited area)}$	40.37	<0.001	0.802
Su-2008	$y = -0.523 \text{ (altitude)} + 0.356 \text{ (pa)} - 0.279 \text{ (dissolved oxygen content)}$	30.30	<0.001	0.693
Au-2008	$y = -0.477 \text{ (altitude)} + 0.428 \text{ (pa)}$	25.50	<0.001	0.557
Sp-2009	$y = 0.709 \text{ (pa)}$	38.30	<0.001	0.489
Su-2009	$y = -0.506 \text{ (altitude)} + 0.335 \text{ (pa)} - 0.292 \text{ (dissolved oxygen content)}$	25.39	<0.001	0.652
Au-2009	$y = -0.574 \text{ (altitude)} - 0.269 \text{ (dissolved oxygen content)} + 0.251 \text{ (pa)}$	22.90	<0.001	0.628
Sp-2010	$y = 0.482 \text{ (pa)} - 0.444 \text{ (altitude)}$	28.58	<0.001	0.586
Su-2010	$y = -0.526 \text{ (altitude)} + 0.422 \text{ (pa)}$	32.66	<0.001	0.619
Au-2010	$y = 0.159 \text{ (altitude)} - 0.296 \text{ (pa)} - 0.438 \text{ (silt)} - 0.279 \text{ (\% wetland)} - 0.428 \text{ (conductivity)} + 0.271 \text{ (\% inhabited area)}$	15.66	<0.001	0.704

Sp, spring; Su, summer; Au, autumn; pa, pond area.

Captions to figures

Fig. 1. A diagrammatic representation of how “elements of metacommunity structure” (EMS) can differentiate among six idealized patterns of metacommunity structure and their quasi-structures, adapted from Willig *et al.* (2011), and originally conceptualized in Leibold & Mikkelsen (2002) and Presley *et al.* (2010). Note that the dark grey ovals are the EMS and the light grey area highlights the "Quasi-" structures.

Fig. 2. An example for the most common best-fit-pattern: incidence matrix of spring 2009 at the entire assemblage level showing a Quasi-Clementsian metacommunity structure. Sites, in columns are ordered according to their position along the first CoA axis, whereas species are in the rows. Species name abbreviations can be found in Appendix II.

Appendix I. The median, minimum and maximum values of the environmental variables used in this study and their type of transformation for regression analyses.

	Transformation	Median	Min.	Max.
Altitude (m)	$x' = \ln(x + 1)$	132.0	107.0	221.0
Distance from source (km)	$x' = \ln(x + 1)$	11.8	1.3	64.5
Distance from mouth (km)	$x' = \ln(x + 1)$	6.2	0.1	93.8
Catchment area above the sampling site (km ²)	$x' = \ln(x + 1)$	93.7	6.9	1165.4
¹ Land cover descriptors (CLC2000)				
% inhabited area (111, 112)	$x' = \arcsin(x^{0.5})$	1.3	0.0	51.9
² % artificial surface (121, 122, 124, 131, 132, 133, 141, 142)	$x' = \arcsin(x^{0.5})$	0.9	0.0	15.8
² % agricultural area (211, 212, 221, 222, 231, 241, 242, 243)	$x' = \arcsin(x^{0.5})$	50.1	13.25	92.1
% forest (311, 312, 313)	$x' = \arcsin(x^{0.5})$	21.3	2.3	58.8
% non-forest vegetation (321, 324, 333)	$x' = \arcsin(x^{0.5})$	10.6	0.0	42.2
% wetlands (411, 412)	$x' = \arcsin(x^{0.5})$	1.8	0.0	7.5
Pond area (km ²)	$x' = \ln(x + 1)$	0.1	0.0	3.1
Wet width (m)	$x' = \ln(x + 1)$	3.2	1.2	7.8
² Depth (cm)	$x' = \ln(x + 1)$	47.4	15.5	77.8
Current velocity (cm s ⁻¹)	$x' = \ln(x + 1)$	14.8	3.4	38.5
CV Width	$x' = \ln(x + 1)$	16.1	7.9	47.6
CV Depth	$x' = \ln(x + 1)$	29.2	6.5	68.4
² CV Current velocity	$x' = \ln(x + 1)$	58.5	24.4	130.2
% Silt (diameter 0–0.02 mm)	$x' = \arcsin(x^{0.5})$	2.4	0.0	71.2
² % Silty sand (diameter 0.02–0.2 mm)	$x' = \arcsin(x^{0.5})$	42.2	0.8	96.9
² % Sand (diameter 0.2–2 mm)	$x' = \arcsin(x^{0.5})$	7.6	0	52.9
% Gravel (diameter 2–60 mm)	$x' = \arcsin(x^{0.5})$	4.9	0.0	64.9
% Stone (diameter 60–300 mm)	$x' = \arcsin(x^{0.5})$	4.7	0.0	22.0
% Rock (diameter >300 mm)	$x' = \arcsin(x^{0.5})$	3.1	0.0	47.9
% Concrete	$x' = \arcsin(x^{0.5})$	0.0	0.0	59.5
pH	$x' = \exp(x)/100$	8.0	7.4	8.3
Conductivity (μS cm ⁻¹)	$x' = \ln(x + 1)$	823.8	170.6	1141.8
Dissolved oxygen (mg l ⁻¹)	$x' = \exp(x)^{0.5}$	7.3	2.7	8.9
Nitrite (mg l ⁻¹)	$x' = \ln(x + 1)$	0.07	0.03	0.23
Nitrate (mg l ⁻¹)	$x' = \ln(x + 1)$	3.81	1.61	17.44
Ammonium (mg l ⁻¹)	$x' = \ln(x + 1)$	0.13	0.08	0.50
Phosphate (mg l ⁻¹)	$x' = \ln(x + 1)$	0.52	0.24	1.70

¹Numbers in parentheses are the three-digit identifying numbers of the original CORINE2000 patch classes.

²Variables discarded due to collinearity.

Appendix II. List of species used in this	Common	Abbreviation	Status	Ind	Rare
<i>Abramis brama</i>	Common	abrbra	N	842	n
<i>Alburnus alburnus</i>	Bleak	albalb	N	4261	n
<i>Ameiurus melas</i>	Black	amemel	NN	427	n
<i>Anguilla anguilla</i>	European eel	angang	NN	31	y
<i>Aspius aspius</i>	Asp	aspasp	N	9	y
<i>Barbatula barbatula</i>	Stone loach	oribar	N	2249	n
<i>Barbus barbus</i>	Barbel	barbar	N	47	y
<i>Blicca bjoerkna</i>	Silver bream	blibjo	N	386	n
<i>Carassius carassius</i>	Crucian carp	carcar	N	32	y
<i>Carassius gibelio</i>	Gibel carp	cargib	NN	5994	n
<i>Cobitis elongatoides</i>	Spined loach	cobelo	N	663	n
<i>Ctenopharyngodon idella</i>	Grass carp	cteide	NN	22	y
<i>Cyprinus carpio</i>	Carp	cypcar	N	117	n
<i>Esox lucius</i>	Pike	esoluc	N	330	n
<i>Gobio gobio</i>	Gudgeon	gobgob	N	5049	n
<i>Gymnocephalus cernuus</i>	Ruffe	gymcer	N	390	n
<i>Hypophthalmichthys molitrix</i>	Silver carp	hypmol	NN	6	y
<i>Lepomis gibbosus</i>	Pumpkinseed	lepgib	NN	1243	n
<i>Leucaspis delineatus</i>	Sunbleak	leudel	N	193	n
<i>Leuciscus idus</i>	Ide	leuidu	N	1	y
<i>Misgurnus fossilis</i>	Mud loach	misfos	N	119	n
<i>Neogobius fluviatilis</i>	Monkey goby	neoflu	NN	243	n
<i>Onchorynchus mykiss</i>	Rainbow	oncmyk	NN	13	y
<i>Perca fluviatilis</i>	Eurasian	perflu	N	2918	n
<i>Perccottus glenii</i>	Chinese	pergle	NN	1	y
<i>Phoxinus phoxinus</i>	Minnow	phopho	N	5731	n
<i>Proterorhinus semilunaris</i>	Tubenose	prosem	NN	40	y
<i>Pseudorasbora parva</i>	Topmouth	psepar	NN	6356	n
<i>Rhodeus sericeus</i>	Bitterling	rhoser	N	8994	n
<i>Romanogobio albipinnatus</i>	Whitefin	romalb	N	80	n
<i>Rutilus rutilus</i>	Roach	rutrut	N	19273	n
<i>Salmo trutta m. fario</i>	Brown trout	saltru	NN	1	y
<i>Sander lucioperca</i>	Pikeperch	sanluc	N	57	y
<i>Sander volgensis</i>	Volga	sanvol	N	3	y
<i>Scardinius erythrophthalmus</i>	Rudd	scaery	N	1171	n
<i>Silurus glanis</i>	European	silgla	N	10	y
<i>Squalius cephalus</i>	Chub	squcep	N	3860	n
<i>Tinca tinca</i>	Tench	tintin	N	38	y
<i>Umbra krameri</i>	European	umbkra	N	91	n