

Contents lists available at ScienceDirect

Agriculture, Ecosystems and Environment

journal homepage: www.elsevier.com/locate/agee



# Seed dispersal between aquatic and agricultural habitats by greylag geese

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ARTICLE INFO

Keywords: Agriculture Anser anser Endozoochory GPS tracking Habitat connectivity Pasture Weed dispersal

#### ABSTRACT

Waterbirds disperse plant seeds within and between aquatic and terrestrial ecosystems in their faeces. However, seed dispersal distances, connectivity among habitat types, and implications for dispersal of weeds remain unquantified in agricultural landscapes. Therefore, we GPS-tagged 31 greylag geese Anser anser and collected 300 faecal samples from feeding flocks in seven agricultural habitats (four cereals, hayfields, pasture, and strawberries) across two landscapes in southern Sweden. We identified intact seeds, determined key plant traits, and tested three hypotheses: (1) geese ingest, transport, and egest seeds from a wide range of terrestrial and aquatic plants, including weeds and alien species; (2) the community and trait composition of plant seeds in faeces vary between habitat types; and (3) seed dispersal by geese is a directional dispersal mechanism that connects some habitat types more than others. We recovered 131 seeds from 41 plant species (19 families), including nine agricultural weeds and one alien species. Many seeds were from aquatic plants (45%), dispersed into terrestrial habitats. A connectivity network formed between habitat types (as nodes) and direct flights (as links) revealed that all agricultural habitats were directly connected with each other, although 66% of flights were between aquatic and agricultural habitats. Geese spent most time at lakes (34%), pastures (14%), barley (10%) and wheat (8%) fields, which were also the most interconnected habitats, with high seed species richness and seed abundance in faecal samples. Combining waterfowl movement data with faecal analysis provided support for all three hypotheses. Geese may contribute to previously overlooked agricultural conflicts through weed dispersal. Proximity to aquatic habitats suitable for roosting may increases the use of agricultural habitats, and potentially the seed dispersal into them.

#### 1. Introduction

Understanding seed dispersal in agricultural landscapes allows for the prediction of how plant species may adjust spatially to habitat destruction, land use change and climate change, or – alternatively – how alien and weed species will spread (Baguette et al., 2012; van Leeuwen, 2018). Waterbirds act as key vectors of seeds in aquatic and terrestrial ecosystems, allowing plants to reach new habitats through their daily and seasonal movements (Kleyheeg et al., 2017; Green et al., 2016, 2023). However, the importance of waterbirds for seed dispersal has been less studied than the role of many other animal vectors (van Leeuwen et al., 2020, 2022).

Waterbirds disperse seeds because a fraction of ingested seeds survives gut passage, and is egested elsewhere (i.e. endozoochory, van Leeuwen et al., 2022). Smaller numbers of seeds can also be carried externally (i.e. epizoochory, Green et al., 2023). An increasing number of studies based on faecal analyses has revealed that many plants with a wide range of seed morphologies and other plant traits are regularly dispersed by gulls, herons, ducks, shorebirds, and other waterbirds (Green et al., 2022; Martín-Vélez et al., 2021a, 2021b; Navarro-Ramos et al., 2022; Urgyán et al., 2023). However, faecal sampling alone does

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https://doi.org/10.1016/j.agee.2023.108741

Received 8 June 2023; Received in revised form 10 August 2023; Accepted 11 September 2023 0167-8809/© 20XX

not demonstrate where seeds are ingested, and what are the dispersal distances or trajectories.

Spatiotemporal aspects of seed dispersal can be studied by combining GPS tracking of waterbirds with faecal samples collected from habitats frequented by waterbirds. However, faecal analysis and GPS tracking of waterbirds has to date been carried out independently, with the exception of a study of gulls wintering in ricefields (Martín-Vélez et al., 2021a, 2021b). The combination of both methods can be used to identify how waterbirds may contribute to connectivity of plant communities in different habitats (Green et al., 2023) and therefore to identify habitats that may be affected by the presence of waterbirds.

The majority of waterbird endozoochory studies have focused on the role of dabbling ducks such as mallards (Anas platyrhynchos), which are abundant, widely distributed, and suitable for captive experiments (Green et al., 2016; Urgyán et al., 2023). Fewer studies have focused on geese, and we focus on the greylag goose (Anser anser), a large, abundant waterfowl widely distributed in Eurasia. The European population is estimated at 950,000 individuals (Wetlands International, 2023). The agricultural impacts of this species through grazing are well investigated (Powolny et al., 2018; Buitendijk et al., 2022; Zhang et al., 2022), but its role in seed dispersal and habitat connectivity is poorly understood. Greylag geese gather in flocks and feed during the day in terrestrial habitats (e.g. grasslands, agricultural fields) near aquatic habitats where they roost at night, safe from terrestrial predators (Avé et al., 2017; Fox et al., 2017). In Sweden, agricultural habitats are frequently used by breeding populations of several goose species (Olsson et al., 2017). Faecal analyses in Swedish Baltic archipelagos have revealed the ability of greylag geese to disperse at least 97 plant species by endozoochory (Hattermann et al., 2019). Compared to dabbling ducks, greylag geese disperse relatively more terrestrial plants and fewer aquatic plants (Almeida et al., 2022).

It is important to evaluate the role of increasing geese populations as vectors of plant dispersal in agricultural habitats, and consider possible implications for spread of weeds. Our main objectives were to quantify seed dispersal by greylag geese in agricultural landscapes, identify key traits of plant species dispersed, and to determine how waterbirdmediated dispersal connects different habitats in such landscapes. Therefore, we collected goose faeces in seven agricultural habitat types within two areas in southern Sweden, and analysed movements of adult geese within these areas. We combined tracking data with the simultaneous collection of faeces in habitats visited by the GPS-tracked individuals. We hypothesised that (1) greylag geese ingest, transport and egest a wide range of terrestrial and aquatic plant seeds, including weeds, native and alien plant species, and seeds with a variety of sizes and morphological dispersal syndromes. Owing to limited research effort to date, we expected to find plant species not previously known to be transported by waterfowl (Anatidae); (2) the community and trait composition of seeds in faeces is unique to each habitat, and related to the level of connectedness of that habitat with other habitat types; and (3) seed dispersal by geese is not a random but a directional dispersal mechanism that connects some habitat types more than others.

# 2. Material and methods

## 2.1. Study species

The greylag goose has expanding populations in northern Europe that often cause agricultural conflicts by reducing yields of grass, cereals and other crops (van Eerden et al., 2005; Buitendijk et al., 2022). The flyway population that includes Sweden has 710,000–780,000 individuals (Wetlands International, 2023). Greylag geese are herbivores feeding mainly in terrestrial habitats, including post-harvest residual crops (van Eerden et al., 2005). They consume up to one third of their body mass daily (~3.3 kg body weight). The proportion of birds wintering at more southern latitudes has declined markedly over the past three decades due to changes in land-use and climate, leading to increases in overall geese numbers and shifts towards northern Europe (Ramo et al., 2015; Fox and Madsen, 2017; Powolny et al., 2018). In Sweden, the breeding population has also increased, and some of these birds now overwinter in the southernmost parts of the country (Nilsson, 2018; Månsson et al., 2022).

## 2.2. Study areas

This study was carried out in two agricultural landscapes: an area close to the city of Kristianstad in the south (56°01'N, 14°09 'E") and an area close to the city of Örebro in south-central Sweden (Kvismaren, 59°10 'N", 15°22 'E") (Fig. 1). Both study landscapes consist of a similar mosaic of shallow lakes, wetlands, forests, grasslands and agricultural fields, according to the CORINE land use categories (2012, CLC; https://land.copernicus.eu/). Both areas are dominated by cereal crops, mainly wheat, barley, rye and oats, but there are also natural grasslands, as well as other crops (Fig. 1). Generally, most crops are harvested from mid-August through October, and some of the cereal crops are re-sown the same autumn in the southernmost parts of the country. The Kristianstad area has a longer growing season and a higher variety of crops, including sugar beet, potatoes, rapeseed, maize and a wide range of vegetables, fruits and other root crops. In both areas, wetlands and lakes are important breeding and roosting sites for geese (Månsson et al., 2022; Nilsson et al., 2022). There are no irrigation canals, drainage ditches or other waterbodies within agricultural habitats.

In both areas, there are three types of grass-dominated fields: 1) pastures, that are grazed by livestock at least part of the year; 2) hayfields, mown several times annually for feed production; and 3) natural, unmanaged grasslands. All three field types are typically composed of a mixture of plant species (e.g. *Trifolium repens, Phleum pratense, Festuca pratensis*), but grasslands and pastures have a higher species richness than hayfields, which are sown for the sole purpose of feed production.

Geese usually visited fields post-harvest as they preferred to feed on short swards of grasslands (Durant et al., 2004; Fox et al., 2017) and postharvest residuals (Olsson et al., 2017). We collected goose faeces in two grassland types (pastures and hayfields) plus wheat and barley fields. Additional habitats were sampled both in Kristianstad (rye and strawberries) and Örebro (oats). In September 2019, a total of 5300 greylag geese were present in Kristianstad and 6000 in Örebro according to ground counts made by experienced personnel.

#### 2.3. Greylag goose movements

Goose movements and connectivity among different habitats were analyzed among the terrestrial habitat types in which it was possible to collect faecal samples, and additionally across lakes, wetlands and islands used for roosting. Movements of 31 greylag geese were monitored between the 1st of June and September 30th 2019 (121 days). In June 2017-2019, individual geese were captured when foraging in fields and equipped with a solar powered GPS tracking device (Ornitela OT-N35 or OT-N44) that sent a position every 15 min (i.e modus of 15) to the Global System for Mobile Communications (GSM) network with an accuracy of  $\leq 10$  m. For further details of the catching procedures and tracking equipment, see Månsson et al. (2022). All catching and handling of geese was performed in 2017-2019 under permit from the Animal Ethics Committee of Central Sweden (# 5.8.18-03584/2017). GPS coordinates were overlaid with cropland maps based on the CORINE Land Cover 2012 and the Swedish Mapping Cadastral and Land Registration Authority.



Fig. 1. : Habitat types of the two study locations, showing fields and habitats visited by GPS-tracked geese. (a) Örebro and (b) Kristianstad and around lake Ivö. The background images show other habitats (including other agricultural habitats) that were not visited by our GPS-tracked geese during summer 2019. All polygons coloured according to the legend belong to habitats used by geese. Orthophoto © Swedish Mapping Cadastral and Land Registration Authority, Dnr 2012/892. Insert map shows location of the study areas in Sweden (Örebro northern diamond, Kristianstad southern).

#### 2.4. Faecal sampling

Faecal samples were collected on 28 days between August 6th and September 13th 2019 in fields selected using greylag goose movements. As geese forage and move in flocks (Scheiber et al., 2013), we assumed that GPS-tagged individuals indicate the position of flocks of geese. Based on the position of GPS-tagged individuals, each sampling day started by selecting the fields for faecal sampling, using exclusively the daily movement data for the 24 h before. Fields were selected where target geese had been present within the past 24 h, and GPS-positioning did not measure instantaneous speeds of > 3 km/h or > 100 m altitudes. This exclusion criterion ensured that fixes in flight were eliminated, allowing us to focus on geese that remained stationary in the selected fields. After identifying fields used by 5–10 target geese, we ran-

domly selected some fields to sample faeces, beginning in early morning. Fresh faeces were collected, with at least 2 m between samples to avoid repeated sampling of the same individual. Each faecal sample was placed in a plastic bag and refrigerated at 4 °C until seed extraction. We used one GPS-tagged goose to identify each field sampled, although sometimes more than one tagged individual was present in the same field. No field was visited more than once in the same week. In total, 123 faecal samples were collected across six habitats in Kristianstad, and 177 samples across five habitats in Örebro (Table S2).

#### 2.5. Seed sample processing

Faecal samples were weighed using a precision balance (Sartorius MSE225P-100-DA, d = 0.01 mg), then washed with tap water over a 100 µm sieve. Sieve contents were examined for plant seeds in petridishes under a stereomicroscope (Stemi 2000-C, Carl-Zeiss). Intact retrieved seeds were photographed with ZEN 2–2.0 software (Carl-Zeiss) and identified to the lowest possible taxonomic level (Bojnanský and Fargašová, 2007; Cappers et al., 2012). When species identification was not possible, genus or family was recorded. All samples were processed within 8 months of collection, with an average delay of 19 weeks.

Mean seed length, Ellenberg F habitat indicator for soil moisture, and morphological dispersal syndromes were obtained as traits for each plant species from the BASEFLOR database (Julve, 1998), LEDA traitbase (Kleyer et al., 2008) or ECOFACT database (Hill et al., 1999). Alien and weed status were determined according to the Euro + Med Plantbase (Euro + Med, 2006 +) and the Agroatlas of Russia (Afonin et al., 2008). Ellenberg F-values indicate the soil moisture preference of each plant species, and range from 1 to 12 (1–2 represents very dry, 3–6 dry to moist, 7–9 moist to wet and 10–12 wet or submerged habitats). We considered aquatic plants to be those with  $F \ge 7$ .

#### 2.6. Germination and viability tests

Intact seeds were placed in Petri-dishes with 1% of bacteriological agar in a germination chamber (Binder KBW400) with a 12 h light (22 °C) and 12 h dark (18 °C) cycle. Germination tests were run for three months and checked on almost daily basis for germination. For ungerminated seeds not infected by fungi during germination trials, a Tetrazolium Chloride Test (TZ) was later used to assess viability (following Brochet et al., 2010; Farmer et al., 2017). Prior to the TZ test, seeds were soaked in water for 24 h and a longitudinal cut was made in the seed coat with a scalpel. Seeds were then placed in multidish wells with a 1% tetrazolium solution at 25 °C in the dark. Seeds that turned red or pink within 48 h were considered viable. These viability tests are highly conservative, as they were conducted over one year after samples were collected, and seed viability can decline rapidly after gut passage (Martín-Vélez et al., 2021a, 2021b).

#### 2.7. Movement analysis

Greylag goose movement data were analysed using the following parameters from the GPS devices: individual bird (ID), date, time of day, latitude, longitude, altitude and instantaneous speed. We applied three filters to the raw data related with longitude, latitude, horizontal dilution of precision and data position frequency (see supporting information S1). After filtering, all 31 individuals were retained in the movement database. As well as identifying fields used for faecal sampling (see above), GPS data from June to September (94  $\pm$  13 days, mean  $\pm$  SD) were used to create a connectivity network to analyse how geese moved among different habitat types.

We distinguished 12 habitat types for the connectivity network, which formed the nodes of a general network for the two study areas combined (details in Table S1). As well as habitats where faeces were collected, five additional habitats were included in the analysis because

they were considered hotspots in terms of goose activity, based on the movement data (see supporting information S1). The network was established on direct flights (DF) of geese between nodes, i.e. non-stop flights from one habitat to another when speed > 3 m/s. The nodes represent all the habitat types visited by the tagged geese, and the links represent direct flights in the directions indicated by arrows. The importance of each habitat (i.e. node) was assessed based on calculating two centrality measures: "strength" and "betweenness". The "strength" measure was an indication of the total number of direct flights (i.e. links) connecting a habitat type/node, and the "betweenness" identified which habitat type/node was used most as the shortest path between two other habitat types/nodes (Brandes, 2001). We used the iGRAPH package in R to build the network and calculate centrality measurements (Csardi and Nepusz, 2006). We also calculated mean distances covered between habitats by geese.

#### 2.8. Statistical analyses

Seed data from faecal samples were analysed using two Generalized Linear Mixed-effects Models (GLMMs). We used negative binomial distributions in the package glmmTMB (Brooks et al., 2017) in R to control for zero-inflated data (only 18% of samples contained seeds). Dependent variables were (1) the number of intact seeds per sample (hereafter "abundance"), and (2) the number of taxa per sample (hereafter "richness"). We also performed separate models for aquatic and terrestrial plant taxa. Habitats present in only one study area were excluded from the analyses to allow models to converge, thus removing three habitats (strawberries, rye, and oats). The remaining four habitat categories (pastures, hayfield, wheat, and barley) were included as levels of a fixed factor. Study area (Kristianstad or Örebro) was included as a random intercept ('Site'), although the plant distributions in both areas were similar (www.artfakta.se/artbestamning). Variation in sample mass was controlled with a continuous predictor variable. Differences among habitat types were assessed with Tukey's HSD post hoc tests (Abdi and Williams, 2010). To identify outliers and to assess the distribution of residuals, quantile-quantile plots were compared with fitted values, but no outliers were found.

The number of faecal samples collected differed among the habitat types, potentially biasing comparisons among habitat types in plant species richness. To correct for this sampling error effect, we carried out a rarefaction analysis using the R package "iNEXT" (Sanders, 1968; Hsieh et al., 2016). To identify differences in the plant community composition among the habitats, we used non-metric multidimensional scaling. We applied the metaMDS function in the package "vegan" using a Bray-Curtis distance matrix (Oksanen et al., 2010). Data were transformed using Wisconsin double standardization, and convergence was reached after 20 iterations (stress score < 0.1). All statistical analyses were performed in R software version 3.6.3 (R Core Team, 2023).

#### 3. Results

#### 3.1. Plant species dispersed into different habitats

Faeces were collected within seven habitat types (Table 1, Fig. 1), and intact plant seeds were recovered in faeces from all of these habitats except oat fields. Faecal sample mass was  $8.36 \pm 4.79$  g (mean  $\pm$  SD, range = 1.77–35.04 g, n = 300). In total, 131 intact seeds were recovered from 41 plant species and 19 families (Table 1). The mean abundance of seeds per sample was  $0.40 \pm 1.32$  SD and the mean number of plant species was  $0.24 \pm 0.71$  SD. Although we only sampled terrestrial habitats, seed abundance and richness were equally divided between aquatic and terrestrial plants. For aquatic plants, mean seed abundance was  $0.20 \pm 0.88$  and mean richness was  $0.12 \pm 0.49$ . For terrestrial plants, mean abundance was  $0.20 \pm 0.83$  and mean richness was  $0.12 \pm 0.37$ . Overall, 10% of the samples contained a single intact

#### Table 1

Details of intact seeds recovered in greylag goose droppings collected on agricultural lands (n = 300). Information is provided on dispersal syndromes (Julve, 1998), alien/native status, Ellenberg F values (3–6 represent dry to moist, 7–9 moist to wet and 10–12 wet or submerged habitats), seed length, crop type (W = Wheat, B = Barley, R = Rye, H = Hayfield, P = Pasture, S = Strawberries), total number of intact seeds (TI) and the number of samples with each taxon (NS). V identifies cases where viability was confirmed:  $\dagger$  = germinated after *Anser anser* gut passage by Jerling et al. (2001); Hattermann et al. (2019);  $\ddagger$  = seeds germinated in this study; \* = viability confirmed with tetrazolium in this study. Species marked in bold and italic indicate alien and cultivated, respectively. <sup>a</sup>species considered agricultural weeds (*sensu* Afonin et al., 2008) and <sup>b</sup>species with herbicide resistance population (*sensu* Heap, 2009).

Peace     Agnotic gigantic*     Barcchory     6     1.75     1     1     1     4     P       Alisma planugo-quartic**     Barcchory     5     2.66     3     2     7     P       Caryophi/lacce     Aramaria serpit/loch     Anemochory     5     2.66     3     2     7     P       Betulacces     Built penhla     Anemochory     5     2.66     3     2     7     P       Poace     Garce selant     Hydrochory     4     7.84     1     1     V     P       Cyperacea     Carce selant     Hydrochory     8     1.55     1     1     V     P       Cyperacea     Carce selant     Hydrochory     3     0.45     1     1     V     P       Cyperacea     Elochoris alustris     Anemochory     1     1.87     1     1     V     P       Cyperacea     Elochoris partula     Hydrochory     10     1.83     1     1     V     P       Cyperacea	Family	Species	Dispersal syndrome	Ellenberg F	Length (mm)	TI	NS	v	Crop type
Alisma creace   Alisma planage-aquatica <sup>2+h</sup> Hydrochory   10   1.1   2   1   1   V   Recent planage     Caryophyllacee   Arman'a serplifolia <sup>1</sup> Anenochory   3   0.4   2   1   V   W     Caryophyllacee   Brands pendia   Anenochory   3   0.4   2   1   V   W     Caryophyllacee   Brands seculiars <sup>3</sup> Anenochory   4   7.4   7.4   1   1   V   W     Cyperaceae   Carex chinaa   Hydrochory   0   1.55   1   1   V   P     Cyperaceae   Carex china   Hydrochory   3   0.46   1   1   V   P     Cyperaceae   Carex china   Hydrochory   3   0.46   1   1   V   P     Cheropolianciae   Cheropolianciae   Anenochory   4   1.19   4   V   P     Cyperaceae   Eleochoris parvia   Anenochory   10   0.83   1   1   V   P     Cyperaceae   Eleochoris naminita   Anenochory   10	Poaceae	Agrostis gigantea <sup>a</sup>	Barochory	6	1.75	1	1	*	Р
Paceae     Abgecurus pretensis     Barchory     5     2.36     3     2     7     P       Caryophyllaces     Arenochory     5     3.2     9     8     7     W.B.P.S       Betula carophyllaces     Betula scollus     Arenochory     5     3.2     9     8     7     W.B.P.S       Carophyllaces     Garax equalis     -     10     1.98     3     1     V     P       Cyperaceae     Carox chinata     Hydrochory     8     1.55     1.3     1     V     P       Carophyllaces     Carax chinata     Arenochory     3     0.45     1     V     P       Carophyllaces     Carophyllaces     Carophyllaces     Carophyllaces     P     P       Chenopolaces     Epizochory     10     1.87     1.8     V     P       Cyperaceae     Elsocharis parvila     Hydrochory     10     1.83     1.8     V     P       Cyperaceae     Elsocharis parvila     Barachory     0     3.5     V	Alismataceae	Alisma plantago-aquatica <sup>a,b</sup>	Hydrochory	10	1.1	2	1	*	R
Carryophyllaceae   Arenaria seryilfifiale <sup>2</sup> Arenaria seryilfifiale <sup>2</sup> Arenaria seryilfifiale <sup>2</sup> Arenaria seryilfifiale <sup>2</sup> M   P     Poaceae   Browns secolnaus <sup>3,6</sup> Epizochory   4   7.84   1   1   V   W   N, P, P, S     Cyperaceae   Carex exhinati   Hydrochory   8   1.55   1   1   V   P     Cyperaceae   Carex exhinati semidiecandrum   Anenochory   3   0.45   1   1   V   P     Chenopodiaceae   Chenopodiaceae   Chenopodiaceae   Chenopodiaceae   Chenopodiaceae   Anenochory   4   1.19   4   V   V   P     Cyperaceae   Eleochris parula   Hydrochory   10   0.87   1   1   V   P     Cyperaceae   Eleochris parula   Hydrochory   10   0.83   1   1   V   P     Coperaceae   Eleochris parula   Anenochory   10   0.83   1   1   V   P     Coperaceae   Eleochory   7   0.33   11   1   V   P	Poaceae	Alopecurus pratensis	Barochory	5	2.36	3	2	t	Р
Betula pendulaAnemochory53.2987W, 8, 7PoaceaeBrazochory47.41.81VWCyperaceaeCarex aquatifis-101.9831VPCyperaceaeCarex elataHydrochory102.461.81VPCaryophyllaceaeCarassium senidecandrumAnemochory30.451.81VPCarnophyllaceaeCarassium senidecandrumAnemochory41.1942PPChenopodiaceaeEducondrumBarcchory101.871VPPAsteraceaeEducoriti paluarisEpisochory100.9103*WPCyperaceaeElecharis panulatBarcchory100.8222VPPCyperaceaeElecharis panulatBarcchory100.822VNNPCopaceaeElecharis manufataBarcchory100.821VNNN <td< td=""><td>Caryophyllaceae</td><td>Arenaria serpyllifolia<sup>b</sup></td><td>Anemochory</td><td>3</td><td>0.4</td><td>2</td><td>1</td><td><b>†</b>*</td><td>W</td></td<>	Caryophyllaceae	Arenaria serpyllifolia <sup>b</sup>	Anemochory	3	0.4	2	1	<b>†</b> *	W
Paceace Cyperaccae Cyperaccae Cyperaccae Carex echinataEpizochory Hydrochory47.841.511VCyperaccae Carex elataHydrochory Hydrochory81.55111VPCyperaccae Carex elataCarex elataHydrochory Hydrochory102.46111VPChenopodianea Chenopodianea Chenopodianea Chenopodianea Elacoharis parvialAnenochory Hydrochory101.8711VPCyperaccae Coperaccae Elacoharis parvial Cyperaccae Elacoharis parvial DurationaAnenochory Hydrochory101.8711VPCyperaccae Coperaccae Elacoharis parvial Durationa Cyperaccae Elacoharis parvial Durationa CoperaccaeHydrochory Elacohory101.8811VPCyperaccae Coperaccae Elacoharis munitata Coperaccae Elacoharis munitata Durationa Dura	Betulaceae	Betula pendula	Anemochory	5	3.2	9	8	ŧ	W, B, P, S
Cyperaceae     Carce aduatilis     -     10     1.98     3     I     P       Cyperaceae     Carce clata     Hydrochory     10     2.46     1     1     V     P       Cyperaceae     Carce solatum sendecandrum     Anemochory     3     0.45     1     1     V     P       Chenopodiace     Cengodian adumrs <sup>10</sup> Barcehory     3     0.45     1     1     V     P       Chenopodiace     Cancyolyllaceae     Conva canadensis <sup>6,4</sup> Anemochory     10     1.87     1     1     V     P       Cyperaceae     Eleocharis panvila     Anemochory     10     0.92     1.8     V     P       Cyperaceae     Eleocharis manvilati     Ratochory     10     0.82     2     V     P     S       Cyperaceae     Epizoochory     10     1.85     1     1     V     P       Concae     Helicotrichon pratins     Epizoochory     7     0.53     11     4     P       Danceaee     <	Poaceae	Bromus secalinus <sup>a,b</sup>	Epizoochory	4	7.84	1	1		W
Cyperaceae CyperaceaeCarce echnanHydrochory81.5511 $\mathbb{F}$ PCaryophyllaceaeCarcatium semidacandrumAnemochory30.4511 $\mathbb{F}$ PChenopodiaceaeChenopodiamalbune <sup>b</sup> Barochory51.2911 $\mathbb{F}$ PChenopodiaceaeChenopodiaceaChenopodiaceaeChenopodiaceaeNo11 $\mathbb{F}$ PCyperaceaeEleocharis parviaBarochory101.8711 $\mathbb{F}$ PCyperaceaeEleocharis parviaBarochory101.3811 $\mathbb{F}$ PCongarceaeEleocharis parviaAnemochory100.8222 $\mathbb{F}$ PCongarceaeEleocharis parviaAnemochory101.8811 $\mathbb{F}$ PCongarceaeFestica pratensisEpizoochory63.6511 $\mathbb{F}$ PPaaceaeHilpuris vulgarisEpizoochory70.3711 $\mathbb{F}$ PJanaceaeJuncus onglomerutusEpizoochory70.3711 $\mathbb{F}$ PJanaceaeJuncus onglomerutusEpizoochory70.3711 $\mathbb{F}$ PJanaceaeJuncus onglomerutusEpizoochory70.3711 $\mathbb{F}$ PJanaceaeJuncus onglomerutusEpizoochory70.3711 $\mathbb{F}$ PJanaceae	Cyperaceae	Carex aquatilis	-	10	1.98	3	1		Р
CyperaceaeCarx elanHydrochory30.4511 $-$ PChanopodiaceaeChenopodian albane <sup>s,b</sup> Barochory51.2911 $+$ PChenopodiaceaeConga canadenis <sup>s,b</sup> Anemochory11.3741 $+$ PAsteraceaeCongra canadenis <sup>s,b</sup> Anemochory101.3711 $+$ PCyperaceaeElecharis pauriaHydrochory100.4311 $+$ PCyperaceaeElecharis namilataBarochory101.3811 $+$ POnagraceaeEpilobium partiforumAnemochory100.8222 $-$ P, SPoaceaeEpilobium partiforumAnemochory100.8222 $-$ P, SPoaceaeEpilobium partiforumAnemochory100.822 $ -$ P, SPoaceaeHelictorichon pratense-45.411 $  -$ PoaceaeHelictorichon pratense-45.411 $  -$ <	Cyperaceae	Carex echinata	Hydrochory	8	1.55	1	1		Р
CaryophyllaceaeCarsaium semidecandrumAnemochory30.45111FPChenopodum album*^hBarochory11.1942500CyperaceaeElecharis parulaHydrochory100.371115PCyperaceaeElecharis parulaHydrochory100.38115PPCoparaceaeElecharis mamilataBarochory100.82225PPConagraceaeElecharis mamilataBarochory63.65115HPPConagraceaeFestuca pretensisEpizoechory51.5122*W, PPPoaceaePagaria x ananasa (Strawberry)Endozoochory70.351148PPJuncaceaeHippur's vigarisHydrochory101.8511*PPJuncaceaeJunceaeHydrochory70.331148PPJuncaceaeMyosofis scorpioliesBarochory70.331148PPJuncaceaePlaum pretenseEpizoechory70.331148PPJuncaceaeMyosofis scorpioliesBarochory103.292228PPoaceaePlaum pretenseEpizoechory122.63111W <td>Cyperaceae</td> <td>Carex elata</td> <td>Hydrochory</td> <td>10</td> <td>2.46</td> <td>1</td> <td>1</td> <td></td> <td>Р</td>	Cyperaceae	Carex elata	Hydrochory	10	2.46	1	1		Р
Chenopodiaceae   Chenopodian album <sup>6,b</sup> Barcchory   4   1.29   1   1   1   1   1   1   1     Asteraceae   Conysa canadensis <sup>6,b</sup> Anemochory   10   1.87   1 <td>Caryophyllaceae</td> <td>Cerastium semidecandrum</td> <td>Anemochory</td> <td>3</td> <td>0.45</td> <td>1</td> <td>1</td> <td>t</td> <td>Р</td>	Caryophyllaceae	Cerastium semidecandrum	Anemochory	3	0.45	1	1	t	Р
AsteraceaeCoryza canadensist*Anemochory11.1942PCyperaceaeEleocharis parvulaHydrochory100.9710311*PCyperaceaeEleocharis mannulataBarochory100.822*PSDoagraceaeEleocharis mannulataBarochory100.822**PSDoagraceaeFestuca pratensisEpizochory63.6511**HRosaceaeRegrafin a canadoss (Strawberry)Endozoochory70.531.11**BPaaceaeHigpuris vidgarisHydrochory70.531.11**PJuncaceaeJuncies conglometransEpizochory70.531.11**PJuncaceaeJuncies biojonius*Epizochory70.531.11**PJuncaceaeJuncies biojonius*Epizochory70.531.11**PJuncaceaeMaunics biojonius*Barochory103.292***PPoaceaePhilum pratenseEpizochory71.5311***PPoaceaePhilum genites aurila*Amemochory122.5311*********PPotamogetonaceaePatangeton pe	Chenopodiaceae	Chenopodium album <sup>a,b</sup>	Barochory	5	1.29	1	1	‡	Р
Cyperaceae   Eleocharis palvaria   Epizochory   10   1.87   1   1   P     Cyperaceae   Eleocharis parvula   Hydrochory   10   0.9   10   3   ‡   W, PC     Cyperaceae   Eleocharis marnilata   Barochory   10   0.82   2   2   P, S     Onagraceae   Epizoachory   6   3.55   1.1   1   H     Rosaceae   Fregoria x ananasaa (Strawberry)   Endozoochory   5   1.5   12   2   *   W, S     Poaceae   Helletorichon pratene   -   4   5.4   1   1   *   B     Ilantaginaceae   Highpris vulgaris   Hydrochory   7   0.33   11   4   *   P     Lamiaceae   Juncaces de missis   Epizoochory   7   0.33   11   *   P     Lamiaceae   Myosoti surgionarias   Barochory   9   1.71   1   1   *   P     Lamiaceae   Myosoti surgionarias   Hydrochory   12   2.25   3   3   *   W <tr< td=""><td>Asteraceae</td><td>Conyza canadensis<sup>a,b</sup></td><td>Anemochory</td><td>4</td><td>1.19</td><td>4</td><td>2</td><td></td><td>Р</td></tr<>	Asteraceae	Conyza canadensis <sup>a,b</sup>	Anemochory	4	1.19	4	2		Р
Cyperaceae   Eleocharis mammilata   Hydrochory   10   0.9   10   0.3   1   1   *   W     Cyperaceae   Eleocharis mammilata   Barochory   10   0.82   2   2   P   S     Daagraceae   Fistica pratensis   Epizoochory   6   3.65   1   1   W   N     Rosaceae   Fageraf a canaosas (Strawberry)   Endozoochory   5   1.5   1.2   2   V   W, S     Poaceae   Helictorichon pratense   -   4   5.4   1   1   *   B     Juncaceae   Hippuris vulgaris   Hydrochory   7   0.37   1   1   *   P     Lamiaceae   Lycopus europaeus   Epizoochory   7   0.37   1   1   *   P     Lamiaceae   Myosotis scorijoides   Barochory   9   1.71   1   1   *   P     Poaceae   Platem graines   Epizoochory   5   1.63   1   1   W     Poaceae   Platongo major <sup>4</sup> Barochory   10   3.2	Cyperaceae	Eleocharis palustris	Epizoochory	10	1.87	1	1		Р
CypenceaeEleochrismammiliataBarochory101.3811*POnagraceaeEpilobium parviforumAnemochory100.8222*P, SPoaceaeFestuca pratensisEpilozochory51.5122*W, SRosaceaeFragaria x ananasa (Strawberry)Endozochory51.5122*W, SPoaceaeHipuris vulgarisHydrochory101.85114*PJuncaceaeJuncus bufonitas*Epilzoochory70.53114*PJuncaceaeJuncus bufonitas*Epilzoochory70.53114*PJuncaceaeJuncus bufonitas*Hydrochory81.4411*PLamiaceaeLycopus curopaeusHydrochory103.2922PPPoaceaePhragmise australis*Anemochory103.2922PPPotamogetonaceaePhataga major*Barochory122.6311*WPotamogetonaceaePotamogeton perchtoldiiHydrochory122.6311*WPotamogetonaceaePotamogeton perchtoldiiHydrochory122.6311*PPotamogetonaceaePotamogeton perchtoldiiHydrochory123.5111*WPotamogetonaceaePotamogeton perchtoldii <td< td=""><td>Cyperaceae</td><td>Eleocharis parvula</td><td>Hydrochory</td><td>10</td><td>0.9</td><td>10</td><td>3</td><td>‡</td><td>W, P</td></td<>	Cyperaceae	Eleocharis parvula	Hydrochory	10	0.9	10	3	‡	W, P
OnagraceaeEpilobium parviforumAnenochory100.8222P, SPoaceaeFragaria x ananasa (Strawberry)Endozochory53.65122*W, SRosaceaeHalcotrichon pratense-45.411*BPlantaginaceaeHippuris vulgarisEpizochory70.53114*PJuncaceaeJuncies confiomeratusEpizochory70.3711*PJuncaceaeJuncies confiomeratusEpizochory70.3711*PJuncaceaeMyosotis scorpioidesBarochory91.7111*PPoaceaePheum pratenseEpizochory51.6311*PPoaceaePatenganies australis*Anenochory103.39222*NPoaceaePotamogeton berchubdiiHydrochory122.2533*WPotamogetonaceaePotamogeton berchubdiiHydrochory123.542*WPotamogetonaceaePotamogeton patinatusHydrochory103.5111*WPotamogetonaceaePotamogeton patinatusHydrochory103.5111*WPotamogetonaceaePotamogeton patinatusHydrochory103.5111*WPotamogetonaceaePotamogeton patinatus	Cyperaceae	Eleocharis mammilata	Barochory	10	1.38	1	1	*	Р
PoaceaeFestuca pratensisEpizoochory63.6511IHRosaceaeFragaria x anansas (Strawberry)Endozoochory51.51.2288PlantaginaceaeHilpuris vulgarisHydrochory101.851.1188JuncaceaeJuncus bufonius*Epizoochory70.5311189JuncaceaeJuncus bufonius*Epizoochory70.371179LamiaceaeLycopus europaeusHydrochory81.4411*9BoraginaceaeMyosotis scorpioidesBarochory91.7111*9PoaceaePhragmites australis*Anenochory103.2922*9PoaceaePhragmites australis*Anenochory103.2922*8PoaceaePhragmites australis*Anenochory122.2533**WPotamogetonaceaePotamogeton bercholdiiHydrochory122.6311*WPotamogetonaceaePotamogeton petrinatusHydrochory122.553*WWPotamogetonaceaePotamogeton petrinatusHydrochory113.3111*WPotamogetonaceaePotamogeton petrinatusHydrochory113.1311*WPotamogetonaceaePotamogeton pet	Onagraceae	Epilobium parviflorum	Anemochory	10	0.82	2	2		P, S
RoscaeeFragaria x ananass (Strawberry)Endozochory51.5122*W, SPoaceaeHelicotrichon pratense-45.411-BPoaceaeHelicotrichon pratenseHydrochory101.8514*BJuncaceaeJuncus bufonius*Epizoochory70.53114*PJuncaceaeLycopus europeausHydrochory81.4411*PBoraginaceaeMyosofi scorpioidesBarochory91.7111*PPoaceaePlatam gratenseEpizoochory51.6311*PPoaceaePlataginaceaePlotanogetonaceaeAmemochory103.292*WPotanogetonaceaePlotanogetonaceaePlotanogetonaceaeAmemochory122.2533*WPotanogetonaceaePotanogeton berkholdiHydrochory122.6311*WPotanogetonaceaePotanogeton berkholdiHydrochory123.542*WPotanogetonaceaePotanogeton berkholdiHydrochory123.542*WPotanogetonaceaePotanogetonaceaePotanogetonaceaePotanogetonaceaeFizoochory72.5311*WPotanogetonaceaeRotanogetonaceaRotanogetonaceaFizoochory70.381	Poaceae	Festuca pratensis	Epizoochory	6	3.65	1	1		Н
PoaceaeHelicotrichon pratense·45.411*BPlantaginaceaeHippuris vulgarisHippuris vulgarisEpizoochory70.53114*8JuncaceaeJuncus bufonius*Epizoochory70.37111*9LamiaceaeLycopus europaeusHiydrochory81.4411*8BoraginaceaeNyosotis scorpioldesBarochory91.7111*8PoaceaePhiragmites ausralis*Anemochory103.2922PPPoaceaePhantaginaceaePoamogeton aceaePotamogeton aceae151.6311WWPotamogeton aceaePotamogeton barchtoldiiHydrochory122.6311WWPotamogeton aceaePotamogeton barchtoldiiHydrochory123.548W, BPotamogeton aceaePotamogeton barisfoliusHydrochory123.511*WPotamogeton aceaePotamogeton aduutus repers*Epizoochory70.3811*WPotamogeton aceaePotamogeton aduutus repers*Epizoochory70.3811*WPotamogeton aceaePotamogeton aduutus repers*Epizoochory70.3811*WPotamogeton aceaeRoumeshisfoliumHydrochory113.1311* <td>Rosaceae</td> <td>Fragaria x ananassa (Strawberry)</td> <td>Endozoochory</td> <td>5</td> <td>1.5</td> <td>12</td> <td>2</td> <td>*</td> <td>W, S</td>	Rosaceae	Fragaria x ananassa (Strawberry)	Endozoochory	5	1.5	12	2	*	W, S
PlantaginaceaeHippuris vulgarisHydrochory101.85111*BJuncaceaeJuncus conglomeratusEpizoochory70.53114*PLamiaceaeJuncus bufonius <sup>a</sup> Epizoochory70.53114*PLamiaceaeLycopus europaeusHydrochory81.4411*BBoraginaceaeMyosoti scorpioidesBarochory91.7111*BPoaceaePhum pratenseEpizoochory51.6311*PPoaceaePhung nites australis <sup>a</sup> Amemochory103.2922*PPotanogetonaceaePotanogeton gramineusHydrochory122.6311*WPotanogetonaceaePotanogeton pectinatusHydrochory122.6311*WPotanogetonaceaePotanogeton pectinatusHydrochory123.542**W, BPotanogetonaceaePotanogeton pectinatusHydrochory123.542*W, BPotanogetonaceaePotanogeton pectinatusHydrochory123.5342*W, BPotanogetonaceaeRaunucular epens <sup>4</sup> Epizoochory70.3811*PPotanogetonaceaeSagran andosaBarochory70.3811*W, BPolyphilaceae </td <td>Poaceae</td> <td>Helictotrichon pratense</td> <td></td> <td>4</td> <td>5.4</td> <td>1</td> <td>1</td> <td></td> <td>В</td>	Poaceae	Helictotrichon pratense		4	5.4	1	1		В
JuncaceaeJuncus conglomeratusEpizochory70.53114*PJuncaceaeJuncus bufonius*Epizochory70.3711*PJuncaceaeJucous europeausHydrochory81.4411*PBoraginaceaeMyosotis scorpioidesBarochory91.7111*PPoaceaePhragmites custralis*Amenochory103.2922*PPlantag inaceaePantago major*Barochory51.5811*WPoaceaePotanogeton berchtoldiiHydrochory122.6311*WPotamogetonaceaePotamogeton berchtoldiiHydrochory123.542**WPotamogetonaceaePotamogeton potinatusHydrochory123.541*WPotamogetonaceaePotamogeton potinatusHydrochory124.0553*W, BPolygonaceaeRanunculus repens*Epizoochory70.3811*WCaryophyllaceaeSagan andosaBarochory70.3811*WCaryophyllaceaeSellaria mediaAnemochory81.021811*WPabaceaeTriftiam andiaAnemochory51.622W, S1WCaryophyllaceaeSellaria mediaAnemochory <td< td=""><td>Plantaginaceae</td><td>Hippuris vulgaris</td><td>Hydrochory</td><td>10</td><td>1.85</td><td>1</td><td>1</td><td>*</td><td>В</td></td<>	Plantaginaceae	Hippuris vulgaris	Hydrochory	10	1.85	1	1	*	В
JuncaceaeJuncus bufonius <sup>a</sup> Epizoochory70.37111 $\uparrow$ PLamiaceaeLycopus europaeusHydrochory81.4411*PBoraginaceaeMyosotis scorpioidesBarochory91.7111‡8PoaceaePhleum pratenseEpizoochory51.6311*PPoaceaePlantago majoraBarochory103.2922PPPlantaginaceaePotamogeton berchtoldiiHydrochory122.2533‡*W,BPotamogetonaceaePotamogeton berchtoldiiHydrochory122.6311*WPotamogetonaceaePotamogeton pectinatusHydrochory123.542\$*W,BPotamogetonaceaePotamogeton pectinatusHydrochory123.511*WPotamogetonaceaePotamogeton pectinatusHydrochory103.5111*WRanunculaceaeRanunculus repens <sup>a</sup> Epizoochory70.3811*WCaryophyllaceaeSgang anodosaBarochory51.02181*WCaryophyllaceaeSgengularia mediaAnemochory51.021811*WPoaceaeVirtica dioicaEpizoochory51.021811*WCaryophyllaceaeSgengularia media<	Juncaceae	Juncus conglomeratus	Epizoochory	7	0.53	11	4	*	Р
LamiaceaeLycopus europaeusHydrochory81.4411IPBoraginaceaeMysoxis scorpicidesBarochory91.71111\$BarochoryPoaceaePhleum pratenseEpizoochory51.6311\$PPoaceaePhragnites australis*Anemochory103.2922PPPlantaginaceaePotamogeton berchtoldiiHydrochory122.2533\$*WPotamogetonaceaePotamogeton berchtoldiiHydrochory122.6311VWPotamogetonaceaePotamogeton obusifoliusHydrochory123.542\$*W, BPotamogetonaceaePotamogeton percinatusHydrochory124.0553*W, BPotamogetonaceaeRunnuclus repens*Epizoochory72.5311*PPolygonaceaeRunnuclus repens*Epizoochory70.3811*W, BPolygonaceaeSagnanium angustifoliumHydrochory113.1311*W, BPolygonaceaeSagnanium angustifoliumHydrochory133.111*W, BPolygonaceaeSagnanium angustifoliumHydrochory113.1311*W, BPolygonaceaeSagnanium angustifoliumHydrochory51.02181*W, B	Juncaceae	Juncus bufonius <sup>a</sup>	Epizoochory	7	0.37	1	1	ŧ	Р
BoraginaceaeMyosotis scorpioidesBarochory91.71111‡BPoaceaePhlum pratenseEpizochory51.6311*PPoaceaePhragnites australis <sup>a</sup> Anemochory103.29222PPlantaginaceaePlantogo major <sup>a</sup> Barochory103.2911*WPotamogetonaceaePotamogeton berchtoldiiHydrochory122.2533\$*W.BPotamogetonaceaePotamogeton obtusifoliusHydrochory123.542\$*W.BPotamogetonaceaePotamogeton obtusifoliusHydrochory123.53*W.BPotamogetonaceaePotamogeton pectinatusHydrochory124.0553*W.BPotamogetonaceaeRumucular repenseEpizocchory72.5311*PPolygonaceaeRumucular repenseEpizocchory70.3811*W.BCaryophyllaceaeSarganium angustifoliumHydrochory113.1311*W.B, HCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria mediacabBarochory51.16228W, B, HPoaceaeTrifolium repenseEpizochory51.661.2148BCaryophyllaceae<	Lamiaceae	Lycopus europaeus	Hydrochory	8	1.44	1	1		Р
PoaceaePhleum pratenseEpizoochory51.6311*PPoaceaePhragmites australis*Anemochory103.2922PPlantag inaceaePlantago major*Barochory51.5811VVPotamogeton berchtoldiiHydrochory122.2533‡*V, BPotamogeton aceaePotamogeton gramineusHydrochory122.6311VWPotamogeton aceaePotamogeton pectinatusHydrochory123.542‡*V, BPotamogeton aceaePotamogeton pectinatusHydrochory123.542‡*V, BPotamogeton aceaeRamunculas repens*Hydrochory103.5111*PPolygonaceaeRamunculas repens*Epizoochory70.3811*PCaryophyllaceaeSagina nodosaBarochory70.3811*PCaryophyllaceaeSpergularia mediaAnemochory51.0452*PFabaceaeTrifolium repens*Epizoochory51.1622*PCaryophyllaceaeStellaria media*bBarochory51.021811*PFabaceaeTrifolium repens*Epizoochory555.7522*PPoaceaeHordeum vulgare (Barley)FEpizoocho	Boraginaceae	Myosotis scorpioides	Barochory	9	1.71	1	1	‡	В
PoaceaePhragmites australisaAnemochory103.29222PPlantaginaceaePlantago majoraBarochory51.5811WPotamogetonaceaePotamogeton berchtoldiiHydrochory122.2533‡*W, BPotamogetonaceaePotamogeton obusifoliusHydrochory123.542‡*W, BPotamogetonaceaePotamogeton obusifoliusHydrochory123.542‡*W, BPotamogetonaceaePotamogeton pectinatusHydrochory124.0553*W, BRanunculaceaeRanunculus repensaEpizoochory72.5311*PPolgogonaceaeSagian nodosaBarochory70.3811*WCaryophyllaceaeSpargminu angustifoliumHydrochory113.1311*WCaryophyllaceaeStellaria mediaa-bBarochory51.021811†‡W, B, HFabaceaeTirfolium repensaEpizoochory51.1622*PPoaceaeHrideun vulgare (Barley)4411*BPoaceaeHrideun vulgare (Barley)Epizoochory56.7522W, SJuncaceaeHordeun vulgare (Barley)Epizoochory56.7522W, SPoaceaeNoruca sp.11*	Poaceae	Phleum pratense	Epizoochory	5	1.63	1	1	*	Р
PlantaginaceaePlantago majoraBarochory51.5811WPotamogetonaceaePotamogeton berchtoldiiHydrochory122.2533\$*W, BPotamogetonaceaePotamogeton gramineusHydrochory122.6311WPotamogetonaceaePotamogeton obusi foliusHydrochory123.542\$*W, BPotamogetonaceaePotamogeton obusi foliusHydrochory124.0553*W, BPotamogetonaceaePotamogeton pectinatusHydrochory124.0553*W, BRanunculaceaeRanunculus repensaEpizoochory72.5311*WCaryophyllaceaeSgaina nodosaBarochory70.3811<	Poaceae	Phragmites australis <sup>a</sup>	Anemochory	10	3.29	2	2		Р
PotamogetonaceaePotamogeton berchtoldiiHydrochory122.25333‡*W, BPotamogetonaceaePotamogeton obusifoliusHydrochory122.6311WPotamogetonaceaePotamogeton obusifoliusHydrochory123.542‡*W, BPotamogetonaceaePotamogeton pectinatusHydrochory123.542‡*W, BRanunculaceaeRanunculus repensaEpizoochory72.5311*PPolygonaceaeRumex hydrolapathumHydrochory103.5111*WCaryophyllaceaeSagina nodosaBarochory70.3811*WCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.162*W, B, HFabaceaeTrifolium repensaEpizoochory51.162*PVurticaceaeUrtica dioicaEpizoochory56.7522W, SPoaceaeTrifolium nulgare (Barley)41*B1*BPoaceaeJuncus sp.422W, S34%BPoaceaePoaceaePoaceaeF11*BB1*BPoaceaePoaceaePoaceaeS6.752<	Plantaginaceae	Plantago major <sup>a</sup>	Barochory	5	1.58	1	1		W
Potamogeton aceaePotamogeton gramineusHydrochory122.6311WPotamogeton aceaePotamogeton obtusifoliusHydrochory123.542‡*W, BPotamogeton aceaePotamogeton pectinatusHydrochory124.0553*W, BRanuculaceaeRanunculus repensaEpizoochory72.5311*PPolygonaceaeRumex hydrolapathumHydrochory103.5111*WCaryophyllaceaeSagina nodosaBarochory70.3811*WCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeDrifolium repensaEpizoochory51.021811†‡W, B, HFabaceaeUtrica dioicaEpizoochory51.1622*PPoaceaeHordeum vulgare (Barley)411*B1*BPoaceaeJuncus sp.456.7522W, S11*B1*B1*B1*B1*B1*B1*B1*NNNNNNNNNNNNN <td< td=""><td>Potamogetonaceae</td><td>Potamogeton berchtoldii</td><td>Hydrochory</td><td>12</td><td>2.25</td><td>3</td><td>3</td><td><b>‡</b>*</td><td>W, B</td></td<>	Potamogetonaceae	Potamogeton berchtoldii	Hydrochory	12	2.25	3	3	<b>‡</b> *	W, B
Potamogeton aceaePotamogeton obtusifoliusHydrochory123.542‡*W, BPotamogeton aceaePotamogeton pectinatusHydrochory124.0553*W, BRanunculaceaeRanunculus repensaEpizoochory72.5311*PPolygonaceaeRumex hydrolapathumHydrochory103.5111*WCaryophyllaceaeSagina nodosaBarochory70.3811*WCaryophyllaceaeSpergularia mediaAnemochory113.1311*WCaryophyllaceaeStellaria media.bBarochory51.021811†‡W, B, HFabaceaeTrifolium repensaEpizoochory51.1622*PUrtica ceaeUrtica dioicaEpizoochory51.1622*PPoaceaeHordeum vulgare (Barley)Epizoochory56.7522W, SJuncaceaeJuncus sp.4Epizoochory56.7522W, SPoaceaePoaceaeJuncus sp.11*BPPoaceaePoaceaePoaceae22W, S, PPoaceaePoaceaePoaceae22W, S, P, PPoaceaePoaceaePoaceae56.7522EPoaceaePoaceaePoaceaeS4\$ </td <td>Potamogetonaceae</td> <td>Potamogeton gramineus</td> <td>Hydrochory</td> <td>12</td> <td>2.63</td> <td>1</td> <td>1</td> <td></td> <td>W</td>	Potamogetonaceae	Potamogeton gramineus	Hydrochory	12	2.63	1	1		W
Potamogeton aceaePotamogeton pectinatusHydrochory124.0553*W, BRanunculaceaeRanunculus repensaEpizoochory72.5311*PPolygonaceaeRumex hydrolapathumHydrochory103.5111WWCaryophyllaceaeSagranium angustifoliumHydrochory113.1311WWCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeTrifolium repnsaEpizoochory51.1622*PUrtica ceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)Epizoochory56.7522W, SJuncaceaeJuncus sp.Epizoochory56.7522W, SPoaceaePoaceaePoaceaeIncus sp.11*BPoaceaePoaceaePoaceae22W, S, R, PPoaceaePoaceaePoaceae56.7522B, PPoaceaePoaceaePoaceae56.7522B, PPoaceaePoaceaePoaceae56.7522B, PPoaceaePoaceaePoaceaeSS55	Potamogetonaceae	Potamogeton obtusifolius	Hydrochory	12	3.5	4	2	<b>‡</b> *	W, B
RanunculaceaeRanunculus repensaEpizoochory72.5311*PPolygonaceaeRumex hydrolapathumHydrochory103.5111WCaryophyllaceaeSagina nodosaBarochory70.3811PTyphaceaeSparganium angustifoliumHydrochory113.1311WCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeTrifolium repnsaEpizoochory51.1622*PUrtica ceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BBBPoaceaeJuncus sp.22W, SS11*BPoaceaePoaceaePoaceaePoaceae22W, SSJuncaceaePoaceaePoaceaeS22W, SSPoaceaePoaceaePoaceae22B, PSPoaceaePoaceaePoaceae22B, PSPoaceaePoaceaePoaceae22B, PSPoaceaePoaceaePoaceaeS22B, PPoaceaePoaceaePoaceaeSS4 <td>Potamogetonaceae</td> <td>Potamogeton pectinatus</td> <td>Hydrochory</td> <td>12</td> <td>4.05</td> <td>5</td> <td>3</td> <td>*</td> <td>W, B</td>	Potamogetonaceae	Potamogeton pectinatus	Hydrochory	12	4.05	5	3	*	W, B
PolygonaceaeRumex hydrolapathumHydrochory10 $3.51$ 11WCaryophyllaceaeSagina nodosaBarochory7 $0.38$ 11PTyphaceaeSparganium angustifoliumHydrochory11 $3.13$ 11WCaryophyllaceaeSpergularia mediaAnemochory8 $1.04$ 52B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory5 $1.02$ 18 $11$ $^{\dagger}\pm$ W, B, HFabaceaeTrifolium repens <sup>a</sup> Epizochory5 $1.16$ 22*PUrtica ceaeHordeum vulgare (Barley)6 $1.21$ 21 $^{\dagger}$ WPoaceaeIrticum aestivum (Wheat)Epizochory5 $6.75$ 22W, SJuncaceaeJuncus sp.1 $1$ *B $^{\bullet}$ $1$ $^{\bullet}$ $1$ PoaceaePoaceaePoaceaeIncus sp.1 $1$ * $8$ $8$ $8$ $1$ <td>Ranunculaceae</td> <td>Ranunculus repens<sup>a</sup></td> <td>Epizoochory</td> <td>7</td> <td>2.53</td> <td>1</td> <td>1</td> <td>*</td> <td>Р</td>	Ranunculaceae	Ranunculus repens <sup>a</sup>	Epizoochory	7	2.53	1	1	*	Р
CaryophyllaceaeSagina nodosaBarochory70.3811PTyphaceaeSparganium angustifoliumHydrochory113.1311WCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeStellaria media <sup>a,b</sup> Barochory51.1622*PUrticaceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BBPoaceaeJuncus sp.56.7522W, SJuncaceaeJuncus sp.11*BPoaceaePoaceaePoaceae22W, S, PPoaceaePoaceaeFoaceae56.7522B, PPoaceaeJuncus sp.11*B, PPoaceaePoaceaeFoaceae54\$W, S, R, PPoaceaePoaceaeFoaceae54\$\$B, PPoaceaePoaceaeFoaceae54\$\$B, PPoaceaePoaceaeFoaceae54\$\$B, PPoaceaePoaceaeFoaceae54\$\$B, PPoaceaePoaceaeFoaceae54\$\$ <td>Polygonaceae</td> <td>Rumex hydrolapathum</td> <td>Hydrochory</td> <td>10</td> <td>3.51</td> <td>1</td> <td>1</td> <td></td> <td>W</td>	Polygonaceae	Rumex hydrolapathum	Hydrochory	10	3.51	1	1		W
TyphaceaeSparganium angustifoliumHydrochory113.1311WCaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeTrifolium repens <sup>a</sup> Epizoochory51.1622*PUrticaceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BBPoaceaeTrificum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.11*BBPoaceaePoaceaePoaceae22W, S, SPoaceaeDageFoaceae22W, S, PPoaceaePoaceaeFoaceae56.7522B, PPoaceaeDage BaceaeSSS55	Caryophyllaceae	Sagina nodosa	Barochory	7	0.38	1	1		Р
CaryophyllaceaeSpergularia mediaAnemochory81.0452B, PCaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeTrifolium repens <sup>a</sup> Epizoochory51.1622*PUrticaceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BPoaceaeTriticum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.11*BPoaceaePoaceaePoaceae2%B, PPoaceaePoaceaePoaceae2%B, PPoaceaePoaceaeSe, P2%B, PPoaceaePoaceaeSe, P2%B, PPoaceaePoaceaeSe, P54\$W, B, R, PTOTALTotATotATotASe\$Se	Typhaceae	Sparganium angustifolium	Hydrochory	11	3.13	1	1		W
CaryophyllaceaeStellaria media <sup>a,b</sup> Barochory51.021811†‡W, B, HFabaceaeTrifolium repens <sup>a</sup> Epizoochory51.1622*PUrticaceaeUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BPoaceaeTriticum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.11*BPoaceaePoa sp.11*B, PPoaceaePoaceaeCoaceae54\$B, PPoaceaePoaceaeTorALS4\$W, B, R, P	Caryophyllaceae	Spergularia media	Anemochory	8	1.04	5	2		В, Р
FabaceaeTrifolium repensaEpizoochory51.1622*PUrtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BPoaceaeTriticum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.11*BPoaceaePoa sp.22B, PPoaceaePoaceaeSoceae54\$W, S, R, PTOTALTOTALTotalTotalTotal1*1	Caryophyllaceae	Stellaria media <sup>a,b</sup>	Barochory	5	1.02	18	11	†‡	W, B, H
Urtica dioicaEpizoochory61.2121†WPoaceaeHordeum vulgare (Barley)411BPoaceaeTriticum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.1118BPoaceaePoa sp.22B, PPoaceaePoaceae56.7522B, PPoaceaePoaceae54\$\$TOTALTOTALTT111	Fabaceae	Trifolium repens <sup>a</sup>	Epizoochory	5	1.16	2	2	*	Р
PoaceaeHordeum vulgare (Barley)41BPoaceaeTriticum aestivum (Wheat)Epizoochory56.7522W, SJuncaceaeJuncus sp.11*BPoaceaePoa sp.22B, PPoaceaePoaceae54\$W, B, R, PTOTALIIIIII	Urticaceae	Urtica dioica	Epizoochory	6	1.21	2	1	ŧ	W
Poaceae     Triticum aestivum (Wheat)     Epizoochory     5     6.75     2     2     W, S       Juncaceae     Juncus sp.     1     1     *     B       Poaceae     Poa sp.     2     2     B, P       Poaceae     Poaceae     5     4     ‡     W, B, R, P       TOTAL	Poaceae	Hordeum vulgare (Barley)		4		1	1		В
Juncaceae   Juncus sp.   1   1   *   B     Poaceae   Poa sp.   2   2   B, P     Poaceae   Poaceae   5   4   ‡   W, B, R, P     TOTAL   131	Poaceae	Triticum aestivum (Wheat)	Epizoochory	5	6.75	2	2		W, S
Poaceae     Poa sp.     2     2     B, P       Poaceae     Poaceae     5     4     ‡     W, B, R, P       TOTAL     131	Juncaceae	Juncus sp.				1	1	*	В
Poaceae     5     4     ‡     W, B, R, P       TOTAL     131     1	Poaceae	Poa sp.				2	2		В, Р
TOTAL 131	Poaceae	Poaceae				5	4	‡	W, B, R, P
	TOTAL					131			

seed and another 8% contained at least two, with a maximum of 11 seeds per sample. Overall, seed viability after gut passage was confirmed for 54% of plant species recorded either by our own germination or tetrazolium tests, or by previous field studies of greylag geese in Sweden (Table 1).

Mean seed length ranged from 0.37 (*Juncus bufonius*) to 7.84 mm (*Bromus secalinus*; Table 1). Morphological dispersal syndromes were identified for 38 species, and included epizoochory (n = 10), hydrochory (n = 12), barochory (n = 8) and anemochory (n = 7) syndromes. The strawberry was the only taxon with an endozoochory syndrome (Table 1).

*Conyza canadensis* is an alien species in Sweden and 11 taxa are considered agricultural weeds in Sweden (Table 1). Only 18 species were terrestrial, i.e. Ellenberg  $F \le 6$ . A total of 23 aquatic taxa were moved into terrestrial habitats, including four *Potamogeton* species (*P. berchtoldii*, *P. gramineus*, *P. obtusifolius*, and *P. pectinatus*). Overall, 45% of the plant seeds were from aquatic plants, but their proportion was

higher in rye and pasture habitats (Fig. 2). Pasture was the habitat with the highest number of plant taxa in faeces, representing 62% of those recorded, even though more samples were collected from barley and wheat (Table S2).

Seed abundance and seed richness did not differ significantly among habitats (Table 2). Similar results were obtained when analysing aquatic and terrestrial plants separately ( $p \ge 0.07$ , Table S4). However, rarefaction analysis for four habitats sampled in both study areas suggested that species richness was much higher in pastures, much lower in hayfields, and intermediate in wheat and barley (Fig. S3). Mean seed abundance and richness were high in the few samples we collected in the strawberry fields (Fig. 2).

Heavier faecal samples contained significantly more seeds and a higher species richness (Table 2). When analysed separately, sample weight had a significant effect for terrestrial plants (abundance: Z = 2.6, p = 0.01; richness: Z = 2.48, p = 0.01), but not for aquatic plants (Table S4). Taxonomic composition of the seeds differed among



**Fig. 2.** : (a) Number of direct flights arriving to each habitat for each of 31 tagged geese. The boxes in the plot represent the middle 50% of the data for each habitat type, with the line inside the box representing the median value. The top of the box represents the 75th percentile, and the bottom of the box represents the 25th percentile for the set of geese. Lower and upper whiskers represent maximum and minimum values of the data, excluding outliers that are represented by dots, (b) Mean values of seed abundance (A) and richness (R) of plant species according to Ellenberg F moisture values ( $F \ge 7$  for aquatic plants).

habitats, with only 27% of taxa recorded in at least two habitat types, and 41% of the species only recorded in pasture (Fig. 3).

#### 3.2. Habitat connectivity network

There were 12 main habitat types visited by greylag geese: 10 terrestrial (wheat, barley, rye, oats, strawberries, hayfields, pastures, grasslands, islands, and "other crops") and two aquatic habitats (lakes and wetlands, Fig. 1). A total of 16384 direct flights were performed over 121 days (7381 flights by 13 individual geese in Kristianstad, and 9003 flights by 19 individuals in Örebro (Table S3, Figs. S1, S2). An average goose therefore performed 5.61  $\pm$  3.39 (mean  $\pm$  SD) direct flights per day in late summer between different habitats. There were 1.77 ( $\pm$  1.45 SD) flights per day from an aquatic to a terrestrial habitat, 1.94 ( $\pm$  1.51 SD) flights from terrestrial to aquatic habitat, 1.75 ( $\pm$  2.24 SD) between two terrestrial habitats, and only 0.15 ( $\pm$  0.52 SD) between lakes and wetlands. For all flights, 80% were done during daylight hours, whereas 20% were night flights.

All pairs of habitats were interconnected by at least one direct flight, although four (pastures, lakes, wheat and barley) were more strongly interconnected and used as sources and/or sinks of most direct flights (Table 3, Fig. 4). Lakes and pastures were involved in the highest number of direct flights (Table S3, "node strength" in Table 3), and they formed important bridges between other habitats ("node betweenness" in Table 3). Overall, 45% of all flights departed from barley, wheat or pasture, and 34% of flights were towards these habitats (Table 3).

Those habitats with more direct flights generally had faeces that contained a higher diversity, richness and abundance of seeds (Figs. 3, 4). Strawberries were an exception, but there was very little of this habitat available (Table 3), and estimates of seeds in strawberry faeces are likely subjected to high sampling error.

Six individual geese spent more than 24 h continuously in the same habitat type on at least one occasion, and this happened in five different habitats: lakes, wetlands, pasture, natural grasslands, and wheat. Overall, geese spent most time in aquatic habitats (wetlands and lakes, 59% combined) (Table 3). During daylight hours, geese stayed in one habitat type for a mean of 2.21 h ( $\pm$  1.17,  $\pm$  SD) before moving to other habitats. They covered linear distances of 6.8 km daily ( $\pm$  1.54). Only 31% of direct flights were between different agricultural habitat types, whereas 66% connected aquatic and agricultural habitats (Fig. 4, Table S3).

#### 4. Discussion

We found greylag geese to be important vectors of seed dispersal in landscapes dominated by agriculture, providing connectivity among different habitat types, both aquatic and terrestrial. Geese dispersed a wide variety of angiosperm taxa with different traits via endozoochory, including weeds and an alien plant species. The combination of tracking individual geese and targeted faecal sampling revealed how different habitats are interconnected, including among terrestrial habitats, among aquatic habitats, and from aquatic to terrestrial habitats. This

#### Table 2

Results of generalized linear mixed models (GLMMs) using negative binomial error distribution to test effects of sample mass (weight) and habitat type on seed abundance and seed richness per sample (N = 300), giving estimates, standard errors of means, standardised effect sizes (Z- values), and the P-value for each explanatory variable (with significant values in bold). Factor level comparison refers to Tukey's range tests corrected for multiple comparisons between habitat pairs.

GLM	Response variable	Explanatory variable	Factor level comparison	Estimate	Error	Z-value	P-value
1	Seed abundance	Intercept		-1.55	0.36	-4.31	< 0.001
		Weight (g)		0.07	0.02	3.02	0.003
		Habitat type		1.40	0.76	1.83	0.07
			Hayfield – Barley	-1.35	0.77	-1.77	0.27
			Pasture – Barley	0.46	0.38	1.22	0.59
			Wheat – Barley	0.05	0.38	0.12	0.99
			Pasture –Hayfield	1.82	0.74	2.46	0.06
			Wheat – Hayfield	1.41	0.76	1.83	0.24
			Wheat – Pasture	-0.42	0.37	-1.12	0.66
2	Seed richness	Intercept		-3.30	0.76	-4.30	< 0.001
		Weight (g)		0.06	0.02	2.73	0.01
		Habitat type		1.37	0.76	1.81	0.07
			Hayfield – Barley	-1.37	0.77	-1.79	0.26
			Pasture - Barley	0.48	0.38	1.27	0.56
			Wheat - Barley	0.01	0.38	0.03	1.00
			Pasture - Hayfield	1.85	0.74	2.49	0.06
			Wheat - Hayfield	1.38	0.76	1.81	0.25
			Wheat - Pasture	-0.47	0.37	-1.28	0.56

study revealed how seeds of many aquatic plants are dispersed into agricultural fields, in which they may or may not establish depending on their timing and local conditions.

#### 4.1. Plant species dispersed by greylag geese, and their traits

A wide variety of plant species with different seed size, moisture requirements and dispersal syndromes were dispersed across different habitats. Twelve of these plant species (31%) have not previously been reported to be dispersed by any European waterfowl species (see Almeida et al., 2022), providing novel insights into how these plant species can be dispersed.

Geese may ingest the seeds they disperse in different ways. At the time of the study, many of the plants would have been at the seed ripening stage and perhaps seeds were deliberately or accidentally ingested along with foliage during grazing (Janzen, 1984; Jaroszewicz et al., 2023). Geese are often likely to ingest seeds from the parent plant when grazing on land and in the water (Amat, 1995; Green et al., 2016, 2018). However, unlike frugivorous birds (González-Varo et al., 2021), waterbirds do not limit the ingestion of seeds to the period of seed production, but can instead ingest seeds intentionally or otherwise from sediments or soil, or when floating (Alderton et al., 2017; Brochet et al., 2010a; Lovas-Kiss et al., 2018; Urgyán et al., 2023).

With the exception of strawberries, all plant species dispersed by geese lacked a fleshy fruit and were assigned to dispersal syndromes other than the "endozoochory syndrome", as previously shown for ducks and shorebirds (Green et al., 2022; Urgyán et al., 2023). In most cases, goose endozoochory will allow plants to move much greater distances than those predicted by other dispersal syndromes (Bullock et al., 2017; Green et al., 2022; Lovas-Kiss et al., 2023). Syndromes may still help explain how seeds are ingested by geese. Seeds with a hydrochory syndrome may be ingested when floating in water, as may those with an anemochory syndrome after reaching water by wind. For instance, silver birch seeds Betula pendula are often blown onto water where they can be ingested by ducks (Lovas-Kiss et al., 2018), and potentially geese. As yet, there is a near-total lack of research as to how plant traits such as buoyant structures, or hooks and hairs, promote endozoochory by non-frugivorous vectors. On the other hand, seed size is a critical trait for endozoochory (van Leeuwen et al., 2023), and smaller seeds may be ingested when attached to waste grain or other residual



Fig. 3. : Non-metric multidimensional scaling (NMDS) plot showing the relationship between plant seeds dispersed in greylag goose faeces collected from different habitat types in Sweden (stress value <0.2). NMDS1 and NMDS2 represent the coordinates obtained from Bray–Curtis matrix distances. Each plant species is assigned these two coordinates, to visualize possible associations or contrasts among plant species in different habitats. Plant species that overlap between habitats are indicated in black, whereas names of species unique to one habitat are given in the colours indicated in the legend.

#### Table 3

Information for the 12 main habitats visited by GPS-tracked greylag geese, in order of time spent there. Includes total surface area (km<sup>2</sup>) per habitat type, mean time spent per tagged goose (represented by percentage) from 1st of June to September 30th of 2019, centrality measures of betweenness (indicating the frequency with which a habitat was used as a node connecting two other nodes), and values of the link weights for each node (i.e. strength, indicating how many direct flights were leaving or arriving to the node, plus the percentage of all direct flights).

Habitat	Area (km²)	Time spent (%)	Node betweenness	Node strength (% DF)
Lakes	2802	34.5	0.96	9062 (28)
Wetland	49	19.5	0.11	2647 (8)
Pasture	15	13.9	0.96	5792 (18)
Barley	24	9.9	0.96	4576 (14)
Grassland	14	8.9	0.96	3325 (10)
Wheat	32	7.4	0.96	4071 (12)
Island	4	0.8	0	107 (<1)
Other crops	13	2.6	0.25	1823 (6)
Strawberries	1	0.5	0.11	226 (1)
Rye	3	1.0	0.96	440 (1)
Oat	3	0.8	0.54	581 (2)
Hayfield	2	0.3	0.25	118 (<1)



**Fig. 4.** : Network connectivity between different habitat types. The total number of direct flights between all polygons of a given habitat type is represented by the thickness and darkness of the links. See Table S3 for precise numbers of direct flights.

# crops, as is likely for weed endozoochory in Spanish ricefields (Martín-Vélez et al., 2021a, 2021b).

The diversity and viability of seeds dispersed by geese via endozoochory was likely underestimated in our study because a) our rarefaction analysis suggested that additional plant species may have remained undetected, b) we sampled a tiny fraction of the daily faecal production of geese in agricultural fields (Hahn et al., 2008), and c) long sample storage times led us to underestimate viability. Nevertheless, viability of the dispersed seeds was confirmed by us or in previous studies for 22 (54%) of the species detected.

# 4.2. Weed dispersal and potential implications for economic impacts

Eleven (28%) of the plant species recorded were weeds, including the widely distributed white goosefoot *Chenopodium album* and the common chickweed *Stellaria media*. These are among the most problematic weeds for Swedish agriculture, with documented herbicide resistance, and are common in our study areas (Afonin et al., 2008; CABI, 2022; Liljander, 2007). Weed seeds were found in all habitat types except strawberry and oat fields. For eight weed species (e.g. *Agrostis gigantean, Alisma plantago-aquatica*) we demonstrated their viability, or else their viability after goose gut passage was already demonstrated by previous research (Jerling et al., 2001; Hattermann et al., 2019). The smallest seed we recorded was of *Juncus bufonius*, a weed readily spread from ricefields into other habitats via gull endozoochory, over maximum distances exceeding 100 km (Martín-Vélez et al., 2021a, 2021b; Peralta-Sánchez et al., 2023). Rye bromegrass *Bromus secalinus*, a problematic weed in rye, wheat and other crops (CABI, 2022) has particularly large seeds (Table 1). Although larger seeds are more likely to be destroyed during gut passage (van Leeuwen et al., 2023), similarly sized seeds are dispersed by geese in North America (Farmer et al., 2017).

The dispersal of weed species and herbicide resistant varieties, by whatever mechanism, has costly impacts on agricultural crops and their yield (Varah et al., 2020; Vila et al., 2021). The potential role of waterfowl in such dispersal has been largely overlooked. More than 70% of weed species worldwide have been assigned a barochory (i.e. gravity) dispersal syndrome, implying that they have very limited ability to spread, except via human activity (Benvenuti, 2007). Geese may play a previously overlooked role in the spread of weeds and herbicide resistance within and between habitats, possibly resulting in increased costs through yield reduction and the need for more intense herbicide control for individual farmers (Farmer et al., 2017; Chauhan, 2021). Our results suggest geese may possibly have a significant role in spreading weeds, especially since ~10,000 greylag geese are present within our two study areas during the summer, and their numbers have increased in recent years (Månsson et al., 2022). However, we have not provided evidence that dispersal of weed seeds by geese leads to weed establishment, and this topic is worthy of further research. Furthermore, weeds may also have positive impacts on ecosystem functioning by maintaining populations of macrofauna and microbes that maintain soil quality for crops (Franke et al., 2009).

# 4.3. Dispersal of plant seeds across habitats and functional connectivity

As shown for dabbling ducks (Urgyán et al., 2023), greylag geese are likely to be important seed vectors during their long-distance migratory flights (Alsos et al., 2015; Ramo et al., 2015). Our study does not study goose migration or cover migratory periods, however it confirms that geese have a high potential to disperse seeds among habitat types within agricultural landscapes. Geese moved daily from roosting areas (usually islands, lakes and wetlands) to feeding sites in agricultural fields (see also Olsson et al., 2017). Considering the seed abundance in faecal samples, a mean dropping rate for greylag goose of  $1.2 \text{ h}^{-1}$  (Hahn et al., 2008), and the numbers of geese, approximately ~100,000 seeds were dispersed daily by geese in our study area. Geese covered mean linear distances of 7 km daily, and likely often dispersed seeds over similar distances. Furthermore, tagged geese occasionally flew non-stop between our two study areas (separated by 300 km), and likely dispersed seeds between them (Lovas-Kiss et al., 2023).

Previous field studies have quantified waterfowl endozoochory through analyses of gut contents or faeces (Green et al., 2016; Soons et al., 2016), but without demonstrating that seeds were actually dispersed between habitats. Within each of our two study areas, geese stayed a mean of 2.2 h before switching habitats, so most seeds may be dispersed into other habitats. Seeds are egested a mean of 7 h or more after ingestion, although median gut retention time is 3 h, and maxima exceed 48 h (García-Álvarez et al. (2015), so a large proportion of seeds ingested may be deposited in other habitats. Dispersal of seeds between habitats is clearly indicated by the abundance of aquatic seeds in faeces from agricultural fields, e.g. hoary willowherb Epilobium parviflorum in strawberry fields, water plantain Alisma plantago-aquatica in rye, or narrowleaf bur-reed Sparganium angustifolium in wheat. Dispersal of aquatic plants into non-irrigated agricultural fields is unlikely to be effective with the possible exception of moist soil plants with Ellenberg F = 7 (van Leeuwen et al., 2022), but effective dispersal is more likely when weed seeds are moved between different crops. A total of 28% of the plant taxa were recorded in at least two agricultural habitat types,

further supporting interchange of seeds between them. Strawberry seeds recovered from a wheat field provide further evidence. The exchange of aquatic species between wetlands or lakes is a major ecosystem service that facilitates effective dispersal, ensures gene flow between populations that are otherwise disconnected, and enables the colonization of new habitats (Green et al., 2016, 2023). This process plays a vital role in maintaining biodiversity and enhancing the overall resilience of ecosystems.

The networks we developed based on GPS tagging suggest an important connectivity role of greylag geese in agricultural landscapes, with possible implications for the spread of alien plants and weeds, and for dispersal across the terrestrial-aquatic interface. Although we here show dispersal of plant seeds by geese, our findings also have implications for possible vectoring of microbes and invertebrates (Green et al., 2023), contaminants (Martínez-Haro et al., 2013), nutrients (Dessborn et al., 2016), and microplastics (Coughlan et al., 2021). For all these elements, our movement analysis showed that goose create stronger and more directional connectivity among specific habitat types (e.g. between pasture and barley) than among other habitat types (e.g. between pasture and rye). These strong connectivity patterns, combined with variation in seed availability between habitats, may contribute to differences in plant communities in the studied habitats. Rarefaction analyses supported differences in seed richness in faeces between agricultural habitats, and community composition of plant seeds in faeces varied among habitats (confirming hypothesis 2). In addition, faeces from the most visited habitats generally presented higher seed abundance and a greater diversity of plant species per unit of faeces, perhaps because these habitats are both the most connected with major sources of seeds (notably lakes and wetlands), and because they provide a diversity of food plants. An important determinant for the number of visits, and therefore the potential impact of geese on agricultural landscapes, was proximity to aquatic roosting habitat. This is in line with a relatively large proportion of small-scale (<1 km) goose movements (Fig. S4) and previous reports in Sweden that showed how geese prefer grasslands, pastures and hayfields that are close to shorelines of lakes and wetlands (Axelsson, 2004; Tennfors, 2013).

#### 5. Conclusions and priorities for future research

Waterbirds provide important ecosystem services (Green and Elmberg, 2014) and the present study underlines their role as seed dispersal vectors. Greylag geese act as important connectors within agricultural habitats, potentially spreading weeds. They also connect aquatic and terrestrial ecosystems, which may share weeds such as J. bufonius or R. repens. Furthermore, nutrients imported into fields from aquatic ecosystems in goose faeces may potentially boost crop growth (Buij et al., 2017). This study suggests that geese may be important drivers of plant communities in agricultural landscapes, and important vectors that exchange plant seeds among aquatic habitats. The combination of GPS tools and faecal sampling identifies important aspects of functional connectivity between different habitats, and reveals how waterbirds disperse native, alien and weed species by local or longdistance movements. Like some other goose species (Buij et al., 2017), greylag geese have shown recent increases in population numbers and altering of migration patterns. For these species, movement studies can be key to reducing crop damage (Månsson et al., 2022), but also to understand seed dispersal. There is ample literature on conflicts caused by increasing goose populations through grazing (Fox et al., 2017), but potential benefits and impacts through seed dispersal have so far been largely ignored. Future studies are needed to compare the flora present in different agricultural habitats, so that seeds coming from elsewhere via geese can be more readily identified. There is also a need to investigate dispersal of plants by geese over longer distances during migrations, not least to understand whether geese can help plants compensate for climate change by moving to cooler latitudes, as shown for dabbling ducks (Viana, 2017; Urgyán et al., 2023; Lovas-Kiss et al., 2023).

Research into weed dispersal and its consequences needs to expand its focus to include geese and other waterbird vectors, as well as human vectors. Our results illustrate how waterbird endozoochory may greatly enhance the dispersal ability of weeds (and herbicide-resistance) across agricultural landscapes. Agricultural strategies for management of weeds and herbicide resistance may need to be improved if a role of geese as effective vectors is confirmed. Future research into the establishment success of weed seeds spread between fields and different crops by geese should be of high priority. In intensively used habitats, the implications of the fungi and other microbes transported in geese faeces (Briscoe et al., 2022) for soil ecology should also be investigated.

# **Uncited references**

Hahn et al., 2008; Isik et al., 2020; Kosztra et al., 2014.

# Funding

All of the sources of funding for the work described in this publication are acknowledged below:

This research was supported by Spanish Ministerio de Economía, Industria y Competitividad project CGL2016-76067-P (AEI/FEDER, EU), and Ministerio de Ciencia e Innovación project PID2020-112774GB-I00 / AEI / 10.13039/501100011033 (to AJG). MJN was supported by an FPI grant from the Spanish Ministerio de Ciencia, Innovación y Universidades (BES-2017-081599) and by the Phd Programme of Biología Integrada from the University of Seville. For the laboratory work, support was provided by staff of the Aquatic Ecology Laboratory (LEA-EBD), and for fieldwork in Sweden MJN was supported economically by the Academy Ecology Fund of the Royal Dutch Academy of Sciences (KNAW). Participation and field work in this study was co-funded by Kristianstad University (JE, CO), the Swedish University of Agricultural Sciences (JM) and through grants from the Swedish Environmental Protection Agency no. 16/71, 16/72, 19/128, and 19/129. ÁLK was supported by: János Bolyai Research Scholarship of the Hungarian Academy of Sciences; New National Excellence Programme of the Hungarian Ministry of Innovation and Technology ÚNKP-21-5-DE-457, NKFIH FK-138698 grants.

#### **Declaration of Competing Interest**

We wish to confirm that there are no known conflicts of interest associated with this publication.

#### **Data Availability**

Data will be made available on request.

# Acknowledgments

This research was supported by Spanish Ministerio de Economía, Industria y Competitividad project CGL2016–76067-P (AEI/FEDER, EU), and Ministerio de Ciencia e Innovación project PID2020–112774 GB-I00 / AEI / 10.13039/501100011033 (to AJG). MJN was supported by an FPI grant from the Spanish Ministerio de Ciencia, Innovación y Universidades (BES-2017–081599) and by the Phd Programme of Biología Integrada from the University of Seville. For the laboratory work, support was provided by staff of the Aquatic Ecology Laboratory (LEA-EBD), and for fieldwork in Sweden MJN was supported economically by the Academy Ecology Fund of the Royal Dutch Academy of Sciences (KNAW). Participation and field work in this study was co-funded by Kristianstad University (JE, CO), the Swedish University of Agricultural Sciences (JM) and through grants from the Swedish Environmental Protection Agency no. 16/71, 16/72, 19/128, and 19/129. ÁLK was supported by: János Bolyai Research Scholarship of the Hungarian Academy of Sciences; New National Excellence Programme of the Hungarian Ministry of Innovation and Technology ÚNKP-21–5-DE-457, NKFIH FK-138698 grants.

#### Intellectual property

We confirm that we have given due consideration to the protection of intellectual property associated with this work and that there are no impediments to publication, including the timing of publication, with respect to intellectual property. In so doing we confirm that we have followed the regulations of our institutions concerning intellectual property.

# Authorship

All listed authors contributed significantly to the creation of this manuscript. We confirm that the manuscript has been read and approved by all named authors. We confirm that the order of authors listed in the manuscript has been approved by all named authors.

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#### Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.agee.2023.108741.

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