1	How to spot a stranger's egg? A mimicry-specific
2	discordancy effect in the recognition of parasitic eggs
3	
4	Csaba Moskát ¹ , Anikó Zölei ² , Miklós Bán ³ , Zoltán Elek ¹ , Lainga Tong ⁴ ,
5	Nikoletta Geltsch ⁵ and Márk E. Hauber ⁴
6	
7	
8	¹ MTA-ELTE-MTM Ecology Research Group, Biological Institute, Eötvös Lóránd
9	University, Pázmány Péter sétány 1/C., H-1117 Budapest, Hungary and Hungarian Natural
10	History Museum, Baross u. 13., H-1088, Hungary
11	² Department of Systematic Zoology and Ecology, Eötvös Loránd University, Pázmány Péter
12	sétány 1/C, H-1117 Budapest, Hungary
13	³ MTA-DE "Lendület" Behavioural Ecology Research Group, Department of Evolutionary
14	Zoology, University of Debrecen, Egyetem tér 1, H-3010 Debrecen, Hungary
15	⁴ Department of Psychology, Hunter College and The Graduate Center of the City University
16	of New York, NY 10065, USA
17	⁵ Department of Ecology, University of Szeged, Közép fasor 52, Szeged, H-6726, Hungary
18	
19 20 21 22 23	
24	Word count: 6,386 incl. References + 965 (tables)

- 25 Abstract
- 26

27 Egg discrimination by hosts is an antiparasitic defence to reject foreign eggs from the nest. 28 Even when mimetic, the presence of brood parasitic egg(s) typically alters the overall 29 similarity of all eggs in a clutch, producing a discordant clutch compared to more 30 homogenous clutches of composed only of hosts' own eggs. In multiple parasitism, the more 31 foreign eggs are laid in the nest, the more heterogeneous the overall clutch appears. 32 Perceptual filters and recognition templates cannot explain the known pattern of lower 33 rejection rates of foreign eggs in multiple vs. single parasitism. We therefore assessed the role 34 of clutch homogeneity and manipulated the colour of one or more eggs in the clutches of great 35 reed warbler (Acrocephalus arundinaceus) hosts of common cuckoos (Cuculus canorus). Varying the colours of both the majority and the minority eggs caused predictable shifts in the 36 rejection of the focal egg(s), and ejection rates of the minority egg colour consistently 37 38 increased but only when it belonged to a more mimetic egg colour, relative to the less 39 mimetic colour of majority eggs. The results imply that in addition to sensory filters, and 40 template-based cognitive decision rules, discordancy-based rejection is affected by the overall 41 clutch appearance, and interacts with specific colours varying in the extent of mimicry, to 42 contribute to the recognition decisions of hosts to reject parasitic eggs. 43 44 45 Keywords: brood parasitism, cuckoo, great reed warbler, egg discrimination, clutch

46 47

48 Short title: Clutch variation and egg discrimination

characteristics, discordancy

- 49 Introduction
- 50

51 Rejecting a foreign egg in the nest is one of most effective steps in eliminating interspecific 52 avian brood parasitism by hosts (Davies 2011). However, egg discrimination is a rather 53 general phenomenon, not restricted to cases of interspecific brood parasitism, as similar 54 mechanisms also occur in intraspecific parasitism (e.g. Yom-Tov 1980; Lyon 2007; Hoi et al. 55 2010). Egg discrimination is a complex cognitive task (de la Colina et al. 2012), especially in the face of egg colour and pattern mimicry of the hosts' own eggs by the parasites (Stoddard 56 57 and Stevens 2010; 2011), and only some host species and certain individuals do so with 58 consistent success (Samaš et al. 2011). 59 Hosts may recognize a foreign egg based on differences in its appearance from the rest

60 of the eggs in the clutch, through the rule of discordancy (Rothstein 1975, Sealy & 61 Underwood 2012), so that eggs in the majority are regarded as own eggs, to be accepted, 62 whereas egg(s) in the minority are regarded as parasitic, to be eliminated (Stokke et al. 1999; 63 Moskát et al. 2008a). Alternatively, hosts may have a perceptual filter or a recognition 64 template, coupled with a discrimination threshold, for own eggs (Rothstein 1975; Lyon 2007; 65 Petrie et al. 2009), so that they do not rely on inspecting own eggs during discrimination to 66 make rejection decisions (Hauber & Sherman 2001; Bán et al. 2013). However, even rejecters might experience perceptual and cognitive constraints limiting the success of egg recognition 67 68 (Krüger 2011), depending on intrinsic factors (age: Lotem et al. 1995; clutch inspection 69 behaviour: Požgavová et al. 2011; Polačiková et al. 2013), or extrinsic factors (e.g., variation 70 in the extent of host-parasite mimicry: Cherry et al. 2007a; ambient light conditions: Honza et 71 al. 2011; or the number of parasitic eggs in the nest: Stevens et al. 2013).

72 Both discordancy and threshold-based recognition mechanisms are efficient because 73 eggs laid by the same female bird (the host) typically show greater similarity to each other 74 compared to eggs laid by different females, including brood parasites (Stokke et al. 2007; 75 Cassey et al. 2011; Honza et al. 2012). Accordingly, even in brood parasitism with mimetic 76 eggs, the foreign egg is typically more dissimilar from the others, disrupting the visual 77 homogeneity of the clutch (Stokke et al. 1999; Moskát et al. 2008a). Thus, in most hosts, 78 resulting in discordant clutches in parasitized nests (Moskát et al. 2010; Stevens et al. 2013). 79 Here we evaluated whether and how in great reed warblers (Acrocephalus 80 arundinaceus), a host locally heavily impacted by single or multiple parasitism of the 81 common cuckoo (*Cuculus canorus*) (Moskát et al. 2009), egg discrimination behaviours are 82 altered by characteristics of the whole clutch, depending on the relative proportions of

83	differently mimetic egg colours in the clutch. Based on past research (Bán et al. 2013), we
84	hypothesized that if a clutch contains at least two different egg phenotypes, it is the extent of
85	each phenotypes difference in egg appearance from the hosts' own eggs, and not their relative
86	status (i.e. majority or minority eggs), that determines the likelihood of egg rejection.
87	
88	
89	Material and Methods
90	
91	Study site and species
92	
93	The study was conducted in the surroundings of Apaj (47°07'N; 19°06'E), ~50 km south of
94	Budapest, Hungary, where common cuckoos commonly parasitize great reed warblers
95	(Moskát & Honza 2002). We searched for nests of hosts in 2-4 m wide reed beds of small
96	channels between May 15 and June 15, in 2009, 2011 and 2013 (for more details on the field
97	work, see Moskát and Hauber 2007). Pseudoreplication of our unbanded subjects, within and
98	across years, was likely low because we did not use nests in the same territory repeatedly and
99	because breeding philopatry is known in this population (Moskát et al. 2008b).
100	
101	Experimental treatments and controls
102	
103	Observations of cases of natural parasitisms by cuckoos, involving typically just one parasitic
104	egg and several host eggs in a clutch, do not permit researchers to explore the underlying
105	mechanism of egg discrimination in hosts of brood parasites. This task needs focal
106	experiments, where the proportion of parasitic eggs in a clutch, as well as the appearance of
107	the parasitic eggs, are manipulated (Moskát & Hauber 2007; Moskát et al. 2010). As the
108	shape of the egg in a clutch may affect egg recognition and rejection decisions in the great
109	reed warbler (Bán et al. 2011; Zölei et al. 2012), we simulated brood parasitism by painting
110	(dyeing) hosts' own eggs with highlighter pens, generating foreign eggs which resemble host
111	eggs in size, shape, and maculation pattern, but differ in background colour (Bán et al. 2013).
112	We manipulated eggs on the day when the clutch was predicted to reach the modal clutch size
113	of five eggs in this population (e.g. Moskát et al. 2011). As these warblers lay one egg per day

114 (Leisler & Schulze-Hagen 2011), this was the fourth day following the appearance of the first

115 host egg in a clutch. In order to generate a representative sample of egg discrimination

116 decisions in our great reed warbler population, we also used nests for experiments with four

117 and six eggs (4 eggs: 10 clutches, 5 eggs: 111 clutches, 6 eggs: 6 clutches). When hosts laid

an extra (6th) egg, it was left unmanipulated or painted with the colour of the majority egg

119 type in experiment (see below).

120 Altogether, we applied one of six treatments to 95 nests (Fig. 1a):

121 "1 green and 4 natural eggs" treatment: One egg in clutch was painted green, and all other122 eggs in clutch remained natural (unmanipulated).

"1 green and 4 blue eggs" treatment: One egg was painted green and the rest were paintedblue.

"1 green and 4 orange eggs" treatment: One egg was painted green and the rest were paintedorange

"1 orange and 4 natural eggs" treatment: One egg in clutch was painted orange, and the othereggs remained natural.

"1 orange and 4 blue eggs" treatment: One egg was painted orange and the rest were paintedblue.

"1 orange and 4 green eggs" treatment: One egg was painted orange and the rest were paintedgreen.

133 We used Stabilo Boss, non-toxic highlighter pens for dying, types 70/33 (green) and 134 70/54 (orange). We compared nests with natural eggs and a dyed ('parasitic') egg (green or 135 orange) with treatments of the same parasitic egg accompanied with parasitic eggs of the 136 other colour (green-orange and orange-green egg types in the minority and majority of the 137 clutches). We chose these egg types as differently mimetic artificial egg colours, based on the 138 calculated avian perceptual model of these eggs against natural eggs (see below), and on the 139 previously known patterns that great reed warblers respond to green vs. orange dyed eggs 140 differently (14% vs. 77% rejection rates toward the green and the orange eggs, respectively, in 141 single parasitism, Bán et al. 2013).

142 As host individuals had the chance to learn the appearance of each of their own, 143 unmanipulated eggs before our experiments started, i.e. during egg laying, we used the 144 treatments with light blue eggs (Stabilo Boss 70/31; treatments "1 green and 4 blue eggs" and 145 "1 orange and 4 blue eggs") to control for this potential learning effect in the "1 green and 4 146 natural eggs" and "1 orange and 4 natural eggs" treatments. . We chose this egg type as 147 previously we reported the lowest rejection rate toward this among differently dyed host own 148 eggs (7% rejection rate in single parasitism, Bán et al. 2013). This means that hosts had four 149 days to view and learn about their natural eggs before the experiment with dyed eggs started 150 on the fifth day after the onset of laying (5d). We dyed eggs blue on 5d, to parallel the

151 experimental methods for all dyed egg colours, so that all subjects are not exposed to the

- 152 manipulated egg colours until the same day of the laying cycle, across the different
- 153 treatments.

As unmanipulated controls, we monitored with the same frequency several nonparasitized clutches in the same years at our study population, and no desertions or egg losses were observed in any of them (n = 21).

157 The treatment to dye eggs did not cause any failure on host reproduction. We tabulated 158 the results our experiments when the nest was monitored at least until hatching, including 159 results of a previous study when different colours of Stabilo Boss highlighter pens were 160 applied (blue, green, yellow, red and orange; Bán et al. 2013 and the present study). Egg 161 hatching rates (%) per clutch did not differ between control nests (mean = 91.24 ± 2.188 SE) 162 and experimental nests (mean = 91.32 ± 2.314 SE) where one or more eggs were dyed (Mann-163 Whitney $U_{41.22} = 429$, P = 0.708).

- 164 Nests were monitored daily for six consecutive days after treatment (e.g. Moksnes et 165 al. 1991; Moskát et al. 2009). If an egg remained in the nest after the monitoring period, the 166 result was regarded as acceptance; missing eggs were regarded as ejected. Only one nest was 167 deserted (in the "1 green and 4 natural eggs" treatment) and we omitted it from the analyses. 168 In one nest of the "1 green and 4 blue eggs" treatment, the green parasitic egg was not ejected 169 on the first day of monitoring, but all of the accompanying four blue eggs were removed, and 170 then the nest with the remaining single green egg was deserted. We categorized it as post-171 ejection nest desertion (sensu Moskát et al. 2011), where desertion was probably caused by 172 the reduced clutch size. As "post ejection nest desertion" was a quick response here (observed 173 after one day the experiment started), reducing potential response period of host from six days 174 to only one, we omitted this nest from the data set used for statistical analyses of single 175 (green) parasitic eggs. However, we could use the ejection of the blue eggs as response to the 176 majority egg type.
- 177

178 Visual modelling of egg types

179

Representative reflectance spectra of natural and experimental eggs were measured by Ocean
Optics USB 2000 spectrometer and taken from Bán et al. 2013 (with the averages shown in
Fig. 1b). Reflectance data were tabulated for the avian perceivable 300-700 nm range at 3

183 different points/egg. We carried out avian visual modelling and calculated just noticeable

184 chromatic differences (JNDs) between randomly chosen pairs of host natural and dyed egg

colours, with the program Avicol 6.0 (Gomez 2006), following parameters and specifications
of Igic et al. (2012), based on a broken canopy nest light environment and an ultraviolet
sensitive oscine, the European blackbird (*Turdus merula*) (Aidala et al. 2012). Although the
absolute values of JNDs from perceptual modelling depend on the parameterization of the
particular sensory model (Igic et al. 2010), their relative values could be informative about the
extent of visual mimicry regarding how hosts' perceptual system can recognize and
distinguish foreign egg colors from own eggs (Avilés 2008; Spottiswoode & Stevens 2010).

192

193 Statistical analyses

194

195 Generalized linear models were used to analyse the relationship between the assumed 196 explanatory variables and the ejection event of the single eggs as a binary response variable 197 (where 0 denotes the acceptance, and 1 is for rejection) and the number of eggs ejected (as 198 count data). Two classes of null distribution were used: "binomial" for occurrence data and 199 "Poisson" for count data. For selecting the most parsimonious model, we applied the above 200 mentioned multi-model inference. The differences between the levels of the tested factor 201 (treatment) were evaluated by multiple comparisons (with Tukey computed contrast matrices 202 for several multiple comparison procedures). The goodness of the models' fit (compared to the 203 null model) was tested by a likelihood ratio test. The analyses were carried out in R 3.0.1 (R 204 Core Team 2013), using the package MuMIn for multi-model inference (Barton 2013), 205 package multcomp (Hothron et al. 2008) for multiple comparisons and the package lrtest for 206 likelihood tests (Zeileis & Hothorn 2002).

207 The model parameters were investigated by multi-model inference (Burnham and 208 Anderson 2002) in order to explore the effect of the explanatory variables such as treatment, 209 year, clutch size, number of days until the first egg laying and the latency in the egg rejection 210 on the response variables such as occurrences of ejection (binary) and the number of eggs 211 ejected (count). This approach was applied on the dataset for the rejection rate of the minority 212 egg type, and the dataset of the majority egg type. For each explanatory variable, we used an 213 information criterion (AICc) to rank the single-argument models in terms of their ability to 214 explain the frequency of ejection or number of eggs ejected (Burnham & Anderson 2002; 215 STable 1). In this way, a "best approximating" model of parameters was selected for the most 216 parsimonious explanation of the data. In the case of both dataset (minority and majority) the 217 latency were excluded from further analyses due to high number of missing values which 218 might distort the model fit. The clutch size was added as a correction term for dispersion

219	parameter (Bolker et al. 2009) to control for the potential effect of different clutch sizes on the
220	response variables (Moskát et al. 2011). For the dataset of the majority egg type, the year was
221	not considered as previous studies on this population did not reveal any year effect (e.g.
222	Moskát & Hauber 2007, Moskát et al. 2009, 2010; for details see STable 1 in the Supporting
223	Material).
224	
225	
226	Results
227	
228	Visual modelling of egg types
229	
230	Perceptual modelling revealed that dyeing host eggs with the three different colours generated
231	a significant variation in the extent of perceivable differences from the hosts' own eggs ($F_{8,24}$
232	= 8.0, $P = 0.007$). Plotting the JND values against those generated by comparisons of host
233	eggs against other conspecifics, revealed that blue eggs were most similar, green eggs
234	intermediate, and orange eggs least similar to the host eggs (Fig. 1c).
235	
233	
235 236	Ejection rates toward the experimental eggs
	Ejection rates toward the experimental eggs
236	<i>Ejection rates toward the experimental eggs</i> Great reed warblers showed a broad range of variation in ejection rates toward the minority
236 237	
236 237 238	Great reed warblers showed a broad range of variation in ejection rates toward the minority
236 237 238 239	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56%
236 237 238 239 240	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in
236 237 238 239 240 241	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in
 236 237 238 239 240 241 242 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of
 236 237 238 239 240 241 242 243 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green
 236 237 238 239 240 241 242 243 244 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green and 4 orange eggs" treatment, i.e. when a green egg was accompanied with four orange eggs,
 236 237 238 239 240 241 242 243 244 245 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green and 4 orange eggs" treatment, i.e. when a green egg was accompanied with four orange eggs, 93.3% of all eggs were ejected from nests where ejection occurred ($n = 9$). In the "1 orange
 236 237 238 239 240 241 242 243 244 245 246 247 248 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green and 4 orange eggs" treatment, i.e. when a green egg was accompanied with four orange eggs, 93.3% of all eggs were ejected from nests where ejection took
 236 237 238 239 240 241 242 243 244 245 246 247 248 249 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green and 4 orange eggs" treatment, i.e. when a green egg was accompanied with four orange eggs, 93.3% of all eggs were ejected from nests where ejection cocurred (n = 9). In the "1 orange and 4 green eggs" treatment 47.8% of all eggs were ejected from nests where ejection took place.
 236 237 238 239 240 241 242 243 244 245 246 247 248 	Great reed warblers showed a broad range of variation in ejection rates toward the minority egg type in response to the different treatments (11-75%; Fig. 2a). We detected 25-56% ejection rates (ejection/clutch) when the two experimental egg types (green or orange) were in the majority in the clutch. Ejection rates toward the two other egg types (natural and blue) in the majority of the clutch were 0-7% (Fig. 2b). Consequently, we observed high numbers of multiple egg ejections from clutches containing both green and orange eggs. In the "1 green and 4 orange eggs" treatment, i.e. when a green egg was accompanied with four orange eggs, 93.3% of all eggs were ejected from nests where ejection took

For the dataset on the minority eggs we tested two candidate models (see STable 2) and we found that the frequency of rejection and the number of eggs rejected were best described by the models where the treatment and the laying date were included as explanatory variables. For the majority eggs, we also tested two models, and we found that the reduced model was the most parsimonious one for both response types (STable 2).

257

258 Estimating discordance by the most parsimonious models

259

260 In the cases of the different minority egg colours, there was significant difference among the 261 treatments, but laying date had no effect on the frequencies of ejections (Table 1). The 262 multiple model comparisons revealed that most of the differently mimetic colour 263 combinations ("1 green and 4 natural eggs" vs. "1 orange and 4 natural eggs" or "1 green and 264 4 blue eggs" vs. "1 orange and 4 blue eggs" treatments") resulted in significantly different 265 rejection rates (Table 2). However we detected no statistical differences in the numbers of 266 eggs rejected neither between the treatments nor for laying date (Tables 1 and 2; 95% 267 confidence intervals are shown in STable 3).

For the majority egg types, the likelihood test revealed that the binary model was not significantly different from the null model (STable 4), thus the estimates from this model are not reported here. The model for the number of eggs ejected revealed significant differences among the treatments (Table 1), so that combinations of differently mimetic egg colours improved the ability of birds to recognize differently-coloured parasitic eggs (Table 2).

- 273
- 274

275 **Discussion**

276

277 Exploring the sensory and cognitive mechanism of how hosts of brood parasites discriminate 278 among own vs. foreign eggs in their clutches is at the core of testing coevolutionary arms 279 races between host defences in recognition systems and parasitic counteradaptations of egg 280 phenotypes, and it has been a great challenge for researchers because most experiments rely 281 on egg rejection behaviour to detect perceptual discrimination (Davies 2000; Rothstein & 282 Robinson 1998; but see Antonov et al. 2009; Požgavová et al. 2011; Moskát et al. 2014). 283 Recent studies have confirmed the most prevalent prediction for the evolution of parasitic egg 284 colour and pattern mimicry, initially tested by human observers (Davies 2000), that less

mimetic eggs, when seen by the avian eye, are rejected more often (Cassey et al. 2008; Avilés
et al. 2010; Spottiswoode and Stevens 2010; Croston & Hauber 2014; Stevens et al. 2013).

287 In our study, we focused on the perceptual mechanisms of egg recognition, which is 288 the first step in the egg discrimination process (Soler et al. 2012). Previous results on the great 289 reed warbler, a frequently parasitized host species of the common cuckoo, already revealed 290 that these birds primarily use template-based egg recognition mechanism in order to recognize 291 the parasitic egg(s) in clutch, so they can discriminate the parasitic eggs even when there is no 292 own egg in the clutch (Moskát & Hauber 2007; Moskát et al. 2010; Bán et al. 2013). 293 However, in addition to template recognition, these hosts can also rely on discordancy, as 294 when they recognize and reject the egg phenotype presenting in the minority of the clutch 295 (Moskát et al. 2010). However, discordancy is error prone, especially in multiple parasitism, 296 as it might lead to the ejection of single own eggs if they are accompanied by several parasitic 297 eggs, as may be possible nests with multiple parasitisms (Rothstein 1974; Moskát et al. 2010).

298 The present study revealed a new aspect of egg recognition, and a complexity of the 299 discordancy-based rejection mechanism, by demonstrating that the majority egg type's 300 coloration, especially when it differs in mimetic appearance from hosts' own eggs, affects the 301 rejection of the minority egg types in discordant clutches. This result suggests that 302 discordancy is a contributing and interactive process with other decision rules involved in egg 303 recognition, and not necessarily a stand-alone mechanism as previously thought by some 304 (reviewed by Moskát et al. 2010). Our new data set up the need for further research to reveal 305 the presence of the role of discordancy in egg recognition in other species that discriminate 306 and reject the parasitic egg. In addition, discordancy-based mechanisms may also have 307 relevance for cognitive processes in intraspecific brood parasitism, where the parasitic 308 (conspecific) eggs are typically a close match ("mimetic") of the host's own eggs, yet several 309 hosts respond to intraspecific parasitism by ejecting foreign eggs, burying the foreign eggs, or 310 deserting the parasitized clutches, (Davies 2000); with sometimes many of them rejected from 311 a clutch (e.g. up to six parasitic eggs in the bearded tit, *Panurus biarmicus*; Hoi et al. 2010). 312 Variation in the colour, size and shape of eggs, or in general, foreign objects in the nest, may 313 motivate ejections in conspecific parasitism (Poláček et al. 2013), similarly to cases in 314 interspecific brood parasitism (see for review in Guigueno & Sealy 2012).

Rejection rates of green and orange eggs, whether in the minority or in the majority of the clutch, were similar when combined with other eggs (natural or blue). However, when in the presence of each other, the rejection rates for the green eggs were higher than expected from the other combinations for single and green eggs, but not for the orange eggs (Fig. 2). This similarity of rejection rates of both the minority green and the majority orange eggs implies that the particular colours of majority vs. minority eggs interact in a previously unknown manner to generate egg rejection behaviours. Results such as these may form the basis of future physiological studies on the hypotheses about the opponency-based process of the avian tetrachromatic vision (Vorobyev & Osorio 1998) in hosts' egg discrimination.

324 The most typical case of parasitized clutches is when the nest contains several host 325 eggs and one parasitic egg, generating a discordant clutch (e.g. in *Molothrus* cowbirds; 326 Hauber 2001; in *Cuculus* cuckoos: Moskát & Hauber 2007). Nevertheless, parasitic eggs 327 could also be the majority egg type in a clutch in areas with locally high rates of multiple 328 parasitism (Moskát et al. 2009; Stevens et al. 2013). Our experiments tested the influence of 329 majority egg colours on the rejection of minority egg colours, and showed that the ejection of 330 the minority egg was increased when it belonged to a less mimetic colour relative to the 331 majority eggs. A previous experimental study, also on the great reed warbler, already showed 332 that the successful ejection of an easily recognisable (i.e. non-mimetic) egg type helped the 333 ejection of another egg type (moderately mimetic) which was otherwise more difficult to 334 recognize by these hosts (Hauber et al. 2006). Egg rejection behaviours may have been due to 335 observational learning to recognise foreign eggs during egg laying in that previous study, but 336 here we excluded this possibility by starting our treatments with dyed eggs at clutch 337 completion. Future studies should clarify how different host species solve the task of own vs. 338 foreign egg discrimination in the face of intrinsic vs. extrinsic sources of variation in the 339 homogeneity of (non-) parasitized clutch appearance (e.g. egg arrangement in the nest: 340 Polačiková et al. 2013).

341 We conclude that great reed warblers' parasitic egg discrimination is affected by the 342 appearance of the whole clutch, not only by the phenotype of the parasitic egg(s) alone. This 343 finding probably has more relevance for systems where parasites repeatedly target or multiple 344 parasites lay in hosts nests (Hauber et al. 2012); for example, with multiple cuckoo eggs laid 345 by the same or different female cuckoos (Wyllie 1981), which is common in our great reed 346 warbler population (Moskát et al. 2009). Although these hosts primarily recognize foreign 347 eggs by a memory-template based mechanism (Bán et al. 2013), they also rely on a 348 discordancy mechanism (Moskát et al. 2010; this study). Here we manipulated the extent of 349 discordancy by using differently mimetic artificial egg colours, to understand this duality in 350 the hosts' decision mechanisms. In concordance with our previous results (Bán et al. 2013), 351 we revealed that discordancy in foreign egg recognition of this cuckoo host species functions 352 not as a primary recognition mechanism, but its effect is contextual, and induces or prevents

egg discrimination in the context of more (or less) mimetic egg colours present in the nest. These results suggest that non-mimetic parasitic eggs may help to recognize mimetic parasitic eggs in the clutch. In a population with high rates of multiple parasitism, often by different females, this recognition mechanism may provide a novel route to accelerate the evolution of egg colour mimicry of host eggs by its specialist parasites, so that differently mimetic parasite eggs are less likely to occur in the same nest (Brooker et al. 1990; Spottiswoode 2013).

359 Previous studies in our great reed warbler population revealed that multiple cuckoo 360 eggs were typically laid by different cuckoo females and cuckoos did not preferentially lay in 361 nests with already present cuckoo eggs; they also removed one already present egg from nest 362 at random (Moskát & Honza 2002). However, laying cuckoos may select host nests with 363 better match between host and parasitic eggs than expected by chance (Avilés et al. 2006; 364 Cherry et al. 2007b; Honza et al. 2014). This process may result in better mimicry of the host 365 eggs by the parasite (Igic et al. 2012), yielding less heterogeneous clutches, even in multiple 366 parasitism, and selecting for further fine-tuning of the sensory and cognitive rules used by 367 hosts to recognize foreign eggs, as seen in our results here. Future studies should aim to reveal 368 whether template-based recognition enhances the recognition of foreign eggs detected through 369 discordancy, and how these two processes interact with perceived mimicry and enhance or 370 interfere with each other egg to yield recognition decisions (Stevens et al. 2013), eventually, 371 to generate the naturally and experimentally elicited rejection behaviours at level of the 372 individual hosts.

373

374 Acknowledgements

The study was supported by the Hungarian National Science Fund (OTKA, No. 83217 to
CM). Additional funding was provided by the Human Frontier Science Program (to MEH),
and the McNair Scholars programs and a Raab Presidential Research Fellowship at Hunter
College (to LT). István Zsoldos and Antal Nagy kindly helped in field work. The MiddleDanube-Valley Inspectorate for Environmental Protection, Nature Conservation and Water
Management provided permission for research.

381

382 **References**

383

Aidala, Z., Huynen, L., Brennan, P. L. R., Musser, J., Fidler, A., Chong, N., Machovsky
Capuska, G. E., Anderson, M. G., Talaba, A., Lambert, D. & Hauber, M. E. 2012:

- Ultraviolet visual sensitivity in three avian lineages: paleognaths, parrots, and
 passerines. J. Comp. Physiol. A **198**, 495–510.
- Antonov, A., Stokke, B. G., Moksnes, A. & Røskaft, E. 2009: Evidence for egg
 discrimination preceding failed rejection attempts in a small cuckoo host. Biol. Lett. 5,
 169-171.
- Avilés, J. M. 2008: Egg colour mimicry in the common cuckoo *Cuculus canorus* as revealed
 by modelling host retinal function. Proc. R. Soc. B 275, 2345-2352.
- Avilés, J. M., Soler, J. J., Soler, M, & Møller, A. P. 2006: Rapid increase in cuckoo egg
 matching in a recently parasitized reed warbler population. J. Evol Biol., 19, 19011910.
- Avilés, J. A., Vikan, J. R., Fossøy, F., Antonov, A., Moksnes, A., Røskaft, E. & Stokke, B. G.
 2010: Avian colour perception predicts behavioural responses to experimental brood
 parasitism in chaffinches. J. Evol. Biol 23, 293-301.
- Bán, M., Barta, Z., Munoz, A. R., Takasu, F., Nakamura, H. & Moskát, C. 2011: The analysis
 of common cuckoo's egg shape in relation to its hosts' in two geographically distant
 areas. J. Zool. 284, 77-83.
- 402 Bán, M., Moskát., C., Barta., Z. & Hauber, M. E. 2013: Simultaneous viewing of own and
 403 parasitic eggs is not required for egg rejection by a cuckoo host. Behav. Ecol. 24,
 404 1014-1021.
- Barton, K. 2013: MuMIn: Multi-model inference. R package version 1.9.5. http://CRAN.Rproject.org/package=MuMIn
- Bolker, B. M., Brooks, M. E., Clark, C. J., Geange, S. W., Poulsen, J. R., Stevens, M. H. &
 White, J. S. 2009: Generalized linear mixed models: a practical guide for ecology and
 evolution. Trends Ecol. Evol. 24, 127-135.
- Brooker, L. C., Brooker, M. G. & Brooker, A. M. H. 1990: An alternative population genetics
 model for the evolution of egg mimesis and egg crypsis in cuckoos. J. Theor. Biol.
 146, 123-143.
- Burnham, K.P. & Anderson, D. R. 2002: Model Selection and Multimodel Inference: a
 Practical Information Theoretic Approach.Springer-Verlag, New York.
- 415 Cassey, P., Hauber, M. E., Maurer, G. & Ewen, J. G. 2011: Sources of variation in reflectance
 416 spectrophotometric data: a quantitative analysis using avian eggshell colours. Methods
 417 Ecol. Evol. 3, 450-456.

- 418 Cassey, P., Honza, M., Grim, T. & Hauber, M. E. 2008: The modelling of avian visual
- 419 perception predicts behavioural rejection responses to foreign egg colours. Biol. Lett.
 420 4, 515-517.
- 421 Cherry, M. I., Bennett, A. T. D. & Moskát, C. 2007a: Host intra-clutch variation, cuckoo egg
 422 matching and egg rejection by great reed warblers. Naturwissenschaften 94, 441-447.
- 423 Cherry, M. I., Bennett, A. T. D. & Moskát, C. 2007b: Do cuckoos choose nests of great reed
 424 warblers on the basis of host egg appearance? J. Evol. Biol. 20, 1218-1222.
- 425 Croston, R. & Hauber, M. E. 2014: Spectral tuning and perceptual differences do not explain
 426 the rejection of brood parasitic eggs by American robins (*Turdus migratorius*). Behav.
 427 Ecol. Sociobiol. 68, 351-362.
- 428 Davies, N. B. 2000: Cuckoos, Cowbirds and Other Cheats. T & AD Poyser, London.
- 429 Davies, N. B. 2011: Cuckoo adaptations: trickery and tuning. J. Zool. 284, 1-14.
- 430 de la Colina, A. M., Pompilio, L., Hauber, M. E., Reboreda, J. C. & Mahler, B. 2012:
- 431 Different recognition cues reveal the decision rules used for egg rejection by hosts of a
 432 variably mimetic avian brood parasite. Anim. Cogn. 15, 881-889.
- Gomez D. 2006: AVICOL, a program to analyze spectrometric data. Last update 2011. Free
 executable available at <u>http://sites.google.com/site/avicolprogram/</u> or from the author
 at dodogomez@yahoo.fr.
- Guigueno, M. F. & Sealy, S. G. 2012: Nest sanitation in passerine birds: implications for egg
 rejection in hosts of brood parasites. J. Ornithol. 153, 35-52.
- Hauber, M. E. 2001. Site selection and repeatability in brown-headed cowbird (*Molothrus ater*) parasitism of eastern phoebe (*Sayornis phoebe*) nests. Can. J. Zool. **79**, 15181523.
- Hauber, M. E. & Sherman, P. W. 2001: Self-referent phenotype matching: theoretical
 considerations and empirical results. Trends Neurosci. 24, 609-616.
- Hauber, M. E., Strausberger, B. M., Feldheim, K. A., Lock, J. & Cassey, P. 2012. Indirect
 estimates of breeding and natal philopatry in an obligate avian brood parasite. J.
 Ornithol. 153, 467-475.
- Hauber, M. E., Moskát, C. & Bán, M. 2006: Experimental shift in hosts' acceptance threshold
 of inaccurate-mimic brood parasite eggs. Biol. Lett. 2,77-180.
- Hoi, H., Darolová, A. & Kristofik, J. 2010: Conspecific brood parasitism and anti-parasite
 strategies in relation to breeding density in female bearded tits. Behaviour 147, 15331549.

- Honza, M., Procházka, P., Morongová, K., Čapek, M. C. & Jelinek, V. 2011: Do nest light
 conditions affect rejection of parasitic eggs? A test of the light environment
 hypothesis. Ethology 117, 539-546.
- Honza, M., Procházka, P. & Požgayová, M. 2012: Within- and between-season repeatability
 of eggshell colouration in the great reed warbler *Acrocephalus arundinaceus*. J. Avian.
 Biol. 43, 91-96.
- Honza M, Šulc M, Jelínek V, Požgayová M & Procházka P. 2014: Brood parasites lay eggs
 matching the appearance of host clutches. Proc. R. Soc. B 281, 20132665.
- Hothorn, T., Bretz, F. & Westfall, P. 2008: Simultaneous inference in general parametric
 models. Biometrical J. 50, 346-363.
- 461 Igic, B., Cassey, P., Grim, T., Greenwood, D. R., Moskát, C., Rutila, J. & Hauber, M. E.
 462 2012: A shared chemical basis of avian host-parasite egg colour mimicry. Proc. R.
 463 Soc. B 279, 1068-1076.
- 464 Igic, B., Leuschner, N., Parker, K. A., Ismar, S. M. H., Gill, B. J., Lovegrove, T. G., Millar,
 465 C. D. & Hauber, M. E. 2010: Size dimorphism and avian-perceived sexual
 466 dichromatism in a New Zealand endemic bird, the whitehead *Mohoua albicilla*. J.
 467 Morph. 271, 697-704.
- 468 Krüger, O. 2011: Brood parasitism selects for no defence in a cuckoo host. Proc. R. Soc. B
 469 278, 2777-2783.
- 470 Leisler, B. & Schulze-Hagen, K. 2011: The reed warblers: diversity in a uniform bird family.
 471 KNNV Uitgeverij.
- 472 Lotem, A., Nakamura, H. & Zahavi, A. 1995: Constraints on egg discrimination and cuckoo473 host co-evolution. Anim. Behav. 49, 1185-1209.
- 474 Lyon, B. 2007: Mechanism of egg recognition in defenses against conspecific brood
 475 parasitism: American coots (*Fulica americana*) know their own eggs. Behav. Ecol.
 476 Sociobiol. 61, 455-463.
- 477 Moksnes, A., Røskaft, E., Braa, A. T., Korsnes, L., Lampe, H. M. & Pedersen, H. C. 1991:
 478 Behavioural responses of potential hosts towards artificial cuckoo eggs and dummies.
 479 Behaviour 116, 64-89.
- 480 Moskát, C. & Hauber, M. E. 2007: Conflict between egg recognition and egg rejection
- 481 decisions in common cuckoo (*Cuculus canorus*) hosts. Anim. Cogn. **10**, 377-386.
- 482 Moskát, C. & Honza, M. 2002: European Cuckoo Cuculus canorus parasitism and host's
- rejection behaviour in a heavily parasitized great reed warbler *Acrocephalus arundinaceus* population. Ibis **144**, 614-622.

- Moskát, C., Avilés, J. M., Bán, M., Hargitai, R. & Zölei, A. 2008a: Experimental support for
 the use of egg uniformity in parasite egg discrimination by cuckoo hosts. Behav. Ecol.
 Sociobiol. 62, 1885-1890.
- Moskát, C., Bán, M., Székely, T., Komdeur, J., Lucassen, R. W. G., van Boheemen, A. L. &
 Hauber, M. E. 2010: Discordancy or template-based recognition? Dissecting the
 cognitive basis of the rejection of foreign eggs in hosts of avian brood parasites. J.
 Exp. Biol. 213, 1976-1983.
- Moskát, C., Hansson, B., Barabás, L., Bártol, I. & Karcza, Z. 2008b: Common cuckoo *Cuculus canorus* parasitism, antiparasite defence and gene flow in closely located
 populations of great reed warblers *Acrocephalus arundinaceus*. J. Avian Biol. **39**, 663671.
- Moskát, C., Hauber, M. E., Avilés, J. M., Bán, M., Hargitai, R. & Honza, M. 2009: Increased
 host tolerance of multiple cuckoo eggs leads to higher fledging success of the brood
 parasite. Anim. Behav. 77, 1261-1290.
- Moskát, C., Hauber, M. E., Elek, Z., Gommers, M., Bán, M., Groenewoud, F., Versluijs, T. S.
 L., Hoetz, C. W. A. & Komdeur, J. 2014: Foreign egg retention by avian hosts in
 repeated brood parasitism: why do rejecters accept? Behav. Ecol. Sociobiol. 68, 403413.
- Moskát, C., Rosendaal, E. C., Boers, M., Zölei, A., Bán, M. & Komdeur, J. 2011: Postejection nest-desertion of common cuckoo hosts: a second defense mechanism or
 avoiding reduced reproductive success? Behav. Ecol. Sociobiol. 65, 1045-1053.
- 506 Petrie, M., Pinxten, R. & Eens, M. 2009: Moorhens have an internal representation of their
 507 own eggs. Naturwissenschaften 96, 405-407.
- Poláček, M., Griggio, M., Bartíková, M. & Hoi, H. 2013: Nest sanitation as the evolutionary
 background for egg ejection behaviour and the role of motivation for object removal.
 PLoS ONE 8, e78771.
- 511 Polačiková, L., Takasu, F., Stokke, B. G., Moksnes, A., Røskaft, E., Cassey, P., Hauber, M.
 512 E. & Grim, T. 2013: Egg arrangement in avian clutches covaries with the rejection of
 513 foreign eggs. Anim. Cogn. 16, 819-828.
- 514 Požgayová, M., Procházka, P., Polačiková, L. & Honza, M. 2011: Closer clutch inspection 515 quicker egg ejection: timing of host responses toward parasitic eggs. Behav. Ecol. 22,
 516 46-51.
- 517 R Core Team 2013: R: A language and environment for statistical computing. R Foundation
 518 for Statistical Computing, Vienna, Austria. URL <u>http://www.R-project.org/</u>.

- Rothstein, S. I. 1974: Mechanisms of avian egg recognition: possible learned and innate
 factors. Auk **91**, 796-807.
- Rothstein, S. I. 1975: Mechanisms of avian egg-recognition: Do birds know their own eggs?
 Anim. Behav. 23, 268-278.
- Rothstein, S. & Robinson, S. K. 1998: The evolution and ecology of avian brood parasitism.
 In Rothstein, S. I. & Robinson, S. K. (eds.) Parasitic Birds and Their Hosts: Studies in
 Coevolution: 3–56. Oxford University Press, New York.
- Samaš, P., Hauber, M. E., Cassey, P. & Grim, T. 2011: Repeatability of foreign egg
 rejection: testing the assumptions of co-evolutionary theory. Ethology 117, 606-619.
- Sealy, S. G. & Underwood, T. J. 2012: Egg discrimination by hosts and obligate brood
 parasites: a historical perspective and new synthesis. Chin. Birds 3, 274-294.
- 530 Soler, M., Fernandez-Morante, J., Espinosa, F. & Martín-Vivaldi, M. 2012: Pecking but
- accepting the parasitic eggs may not reflect ejection failure: the role of motivation.
 Ethology 118, 662-672.
- Spottiswoode, C. N. 2013: A brood parasite selects for its own egg traits. Biol. Lett. 9,
 20130573.
- Spottiswoode, C. N. & Stevens, M. 2010: Visual modeling shows that avian host parents use
 multiple visual cues in rejecting parasitic eggs. PNAS 107, 8672-8676.
- 537 Stevens, M., Troscianko, J, & Spottiswoode C. N. 2013: Repeated targeting of the same hosts
 538 by a brood parasite compromises host egg rejection. Nature Comm. 4, 2475.
- 539 Stoddard, M. C. & Stevens, M. 2010: Pattern mimicry of host eggs by the common cuckoo, as
 540 seen through a bird's eye. Proc. R. Soc. B. 277, 1387-1393.
- 541 Stoddard, M. C. & Stevens, M. 2011: Avian vision and the evolution of egg color mimicry in
 542 the common cuckoo. Evolution 65, 2004-2013.
- 543 Stokke, B. G., Takasu, F., Moksnes, A. & Røskaft, E. 2007: The importance of clutch
 544 characteristics and learning for antiparasite adaptations in hosts of avian brood
 545 parasites. Evolution 61, 2212-2228.
- 546 Stokke, B. G., Moksnes, A., Røskaft, E., Rudolfsen, G. & Honza, M. 1999: Rejection of
 547 artificial cuckoo (*Cuculus canorus*) eggs in relation to variation in egg appearance
 548 among reed warblers (*Acrocephalus scirpaceus*). Proc. R. Soc. Lond. B 266, 1483549 1488.
- Zeileis, A. & Hothorn T. 2002: Diagnostic Checking in Regression Relationships. R News
 2(3), 7-10. URL http://CRAN.R-project.org/doc/Rnews/

- Zölei, A., Hauber, M. E., Geltsch, N. & Moskát, C. 2012: Asymmetrical signal content of egg
 shape as predictor of egg rejction by great reed warblers, hosts of the common cuckoo.
 Behaviour 149, 391-416.
- 555 Vorobyev, M. & Osorio, D. 1998: Receptor noise as a determinant of colour thresholds. Proc.
 556 R. Soc B 265, 351-358.
- 557 Wyllie, I. 1981: The Cuckoo. Batsford, London.
- 558 Yom-Tov, Y. 1980: Intraspecific nest parasitism in birds. Biol. Rev. 55, 93-108.

559

560

561 Table 1. Results of the most parsimonious models for estimating the impact of colour-562 dependent discordancy on egg rejection by great reed warbler. $\alpha = 0.05$, significant P values 563 are underlined for added emphasis. 'Response' is the reaction of hosts to the experimental 564 parasitism, either acceptance or ejection. 'Treatment' refers to experimental manipulations, 565 when one egg in the clutch ("minority") was dyed green or orange, and the rest of the eggs 566 ("majority") dyed a different colour (either orange, green or blue) or were left in their natural 567 state. 'Laydate' denotes the number of days passed after the appearance of the first egg, 568 'clsize' is clutch size; and offset is the dispersion correction parameter.

569

Model (dataset, type of response)	Estimates	SE	Ζ.	Р
Minority, response				
~treatment+laydate+offset(log(clsize+1))				
Intercept ("1 green and 4 blue eggs")	-2.99	1.03	-2.09	0.003
"1 green and 4 natural eggs"	-0.1	1.09	-0.09	0.92
"1 green and 4 orange eggs"	1.93	0.97	1.98	0.047
"1 orange and 4 natural eggs"	2.84	0.94	3.02	<u>0.002</u>
"1 orange and 4 blue eggs"	2.42	0.97	2.49	0.012
"1 orange and 4 green eggs"	2.11	0.95	2.21	<u>0.026</u>
Laydate	-0.02	0.04	-0.72	0.47
Minority, no. of eggs ejected				
~treatment+laydate+offset(log(clsize+1))				
Intercept ("1 green and 4 blue eggs")	-3.43	0.85	-3.99	< 0.001
"1 green and 4 natural eggs"	-0.14	1.01	-0.14	0.88
"1 green and 4 orange eggs"	1.28	0.819	1.56	0.11
"1 orange and 4 natural eggs"	1.61	0.76	2.12	<u>0.03</u>
"1 orange and 4 blue eggs"	1.48	0.786	1.88	<u>0.05</u>
"1 orange and 4 green eggs"	1.35	0.796	1.7	0.088
Laydate	-0.013	0.02	-0.46	0.63
Majority, response				
~treatment+offset(log(clsize+1))				
Intercept ("1 orange and 4 green eggs")	-2.89	0.57	-5.02	< 0.001
"1 green and 4 orange eggs"	1.35	0.76	1.77	0.076

Majority, no. of eggs ejected

~treatment+offset(log(clsize+1))				
Intercept ("1 orange and 4 green eggs")	-2.009	0.27	-7.24	< 0.001
"1 green and 4 orange eggs"	1.008	0.32	3.08	0.002

 Table 2. Multiple comparisons for treatment effect (with Tukey contrast matrices) estimating the discordance by egg rejection for Reed Warbler; $\alpha = 0.05$, significant *P* values are underlined for added emphasis. 'Response' is the reaction of hosts to the experimental parasitism, either acceptance or ejection. 'Treatment' means experimental treatments, when one egg in the clutch ("minority") was dyed green or orange, and the rest of the eggs ("majority") to a different colour (either orange, green or blue) or were left in their original, natural state.

579

Comparisons (dataset, type of response)	Estimates	<i>S.E</i> .	Ζ	Р
Minority, response				
"1 green and 4 natural eggs" - "1 green and 4 blue eggs"	-0.1022	1.0994	-0.093	1.000
"1 green and 4 natural eggs" - "1 green and 4 orange eggs"	2.0395	0.9151	2.229	0.220
"1 green and 4 natural eggs" - "1 orange and 4 natural eggs"	2.9448	0.9207	3.198	<u>0.017</u>
"1 green and 4 natural eggs" - "1 orange and 4 blue eggs"	2.5245	0.9422	2.679	0.077
"1 green and 4 natural eggs" - "1 orange and 4 green eggs"	2.2137	0.9094	2.434	0.141
"1 green and 4 blue eggs" - "1 green and 4 orange eggs"	1.9374	0.9777	1.982	0.347
"1 green and 4 blue eggs" - "1 orange and 4 natural eggs"	2.8426	0.9412	3.020	<u>0.029</u>
"1 green and 4 blue eggs" - "1 orange and 4 blue eggs"	2.4223	0.9701	2.497	0.122
"1 green and 4 blue eggs" - "1 orange and 4 green eggs"	2.1115	0.9514	2.219	0.224
"1 green and 4 orange eggs" - "1 orange and 4 natural eggs"	0.9052	0.7483	1.210	0.829
"1 green and 4 orange eggs" - "1 orange and 4 blue eggs"	0.4849	0.7689	0.631	0.988
"1 green and 4 orange eggs" - "1 orange and 4 green eggs"	0.1742	0.7187	0.242	0.999
"1 orange and 4 natural eggs" - "1 orange and 4 blue eggs"	0.4203	0.7677	0.547	0.994
"1 orange and 4 natural eggs" - "1 orange and 4 green eggs"	0.7311	0.7326	0.998	0.917
"1 orange and 4 blue eggs" - "1 orange and 4 green eggs"	-0.3108	0.7581	-0.410	0.999
Minority, no. of eggs ejected				
"1 green and 4 natural eggs" - "1 green and 4 blue eggs"	-0.1496	1.0127	-0.148	1
"1 green and 4 natural eggs" - "1 green and 4 orange eggs"	1.4299	0.7924	1.804	0.441
"1 green and 4 natural eggs" - "1 orange and 4 natural eggs"	1.7651	0.7592	2.325	0.17
"1 green and 4 natural eggs" - "1 orange and 4 blue eggs"	1.6332	0.7854	2.079	0.28
"1 green and 4 natural eggs" - "1 orange and 4 green eggs"	1.5054	0.7817	1.926	0.366
"1 green and 4 blue eggs" - "1 green and 4 orange eggs"	1.2803	0.8193	1.563	0.603
"1 green and 4 blue eggs" - "1 orange and 4 natural eggs"	1.6155	0.7611	2.123	0.258
"1 green and 4 blue eggs" - "1 orange and 4 blue eggs"	1.4836	0.7861	1.887	0.389
"1 green and 4 blue eggs" - "1 orange and 4 green eggs"	1.3558	0.7961	1.703	0.508
"1 green and 4 orange eggs" - "1 orange and 4 natural eggs"	0.3352	0.4608	0.727	0.976

"1 green and 4 orange eggs" - "1 orange and 4 blue eggs"	0.2033	0.5035	0.404	0.998
"1 green and 4 orange eggs" - "1 orange and 4 green eggs"	0.0755	0.4902	0.154	1
"1 orange and 4 natural eggs" - "1 orange and 4 blue eggs"	0.1319	0.4273	0.309	1
"1 orange and 4 natural eggs" - "1 orange and 4 green eggs"	0.2597	0.4317	0.602	0.99
"1 orange and 4 blue eggs" - "1 orange and 4 green eggs"	-0.1278	0.4762	-0.268	1
Majority, response				
"1 green and 4 orange eggs" - "1 orange and 4 green eggs"	1.35	0.76	1.77	0.076
Majority, no. of eggs ejected				
"1 green and 4 orange eggs" - "1 orange and 4 green eggs"	1.008	0.32	3.08	0.002