

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19

**Phenoconversion of *CYP2C9* in epilepsy limits the predictive value of
CYP2C9 genotype in optimizing valproate therapy**

Katalin Tóth^{1*}, Tamás Búdi^{2*}, Ádám Kiss¹, Manna Temesvári¹, Edit Háfra¹, Andrea Nagy³,
Zsuzsa Szever³, Katalin Monostory¹

¹ Research Centre for Natural Sciences, Hungarian Academy of Sciences
Magyar Tudósok 2, H-1117 Budapest, Hungary
² 2nd Department of Pediatrics, Semmelweis University
Tűzoltó 7-9, H-1094 Budapest, Hungary
³ Heim Pál Children's Hospital
Madarász 22-24, H-1131 Budapest, Hungary

* Katalin Tóth and Tamás Búdi contributed equally to the content of the work.

Corresponding author: Katalin Monostory
Address: Magyar Tudósok 2, H-1117 Budapest, Hungary
Phone: +36 1 382-6747
e-mail: monostory.katalin@ttk.mta.hu

20 **Abstract**

21 **Aims:** Since prominent role in valproate metabolism is assigned to CYP2C9 in pediatric
22 patients, the association between children's CYP2C9-status and serum valproate
23 concentrations or dose-requirements was evaluated.

24 **Methods:** The contribution of *CYP2C9* genotype and CYP2C9 expression in children (N=50,
25 Caucasian) with epilepsy to valproate pharmacokinetics was analyzed.

26 **Results:** Valproate concentrations were significantly lower in normal expressers with
27 *CYP2C9*1/*1* than in low expressers or in patients carrying polymorphic *CYP2C9* alleles.
28 Consistently, the dose-requirement was substantially higher in normal expressers carrying
29 *CYP2C9*1/*1* (33.3 mg/kg vs 13.8-17.8 mg/kg, P<0.0001). Low CYP2C9 expression
30 significantly increased the ratio of poor metabolizers predictable from *CYP2C9* genotype (by
31 46%).

32 **Conclusion:** Due to the substantial down-regulation of CYP2C9 expression in epilepsy,
33 inferring patients' valproate metabolizing phenotype merely from *CYP2C9* genotype results
34 in false prediction.

35

36

37 **Keywords:** personalized medication, epilepsy, pediatric patients, valproate therapy,
38 cytochrome P450, CYPtest, *CYP2C9* genotype, CYP2C9 expression

39

40 **Abbreviations:** CYP cytochrome P450; VPA valproate;

41

42 **Introduction**

43 One percent of Hungarian pediatric population has been reported to suffer from
44 epilepsy [1], but most of them are treated successfully with anticonvulsants. One of the first
45 choices of antiepileptic therapy is valproic acid (VPA), which is generally well-tolerated, and
46 rarely induces serious side effects. Rare complications may occur in patients treated
47 chronically with VPA, including hepatotoxicity, hematologic disorders, hyperammonemic
48 encephalopathy or neurological toxicity [2,3]. The risk of serious adverse effects is increased
49 in children, especially in those younger than 2 years of age. The mechanism of VPA-induced
50 toxicity is not clearly understood, but both the parent compound and some of its unsaturated
51 metabolites have been associated with mitochondrial dysfunction and cytotoxicity [4].

52 VPA, the branched short-chain fatty acid, is extensively metabolized in the liver,
53 resulting in conjugated, unsaturated and hydroxylated metabolites [5,6]. In adults, the
54 majority of VPA dose is eliminated as glucuronide conjugate in the urine. Mitochondrial
55 β -oxidation is the second major route of biotransformation, forming 2-ene-VPA, 2,4-diene-
56 VPA and 3-keto-VPA. The cytochrome P450 (CYP) mediated branch of VPA metabolism is
57 the formation of 4-ene-VPA and hydroxylated metabolites (3-, 4-, and 5-hydroxy-VPA
58 metabolites) [7,8]. Kiang *et al.* have demonstrated that CYP2C9 is the major enzyme in CYP-
59 mediated metabolism of VPA, accounting for about 10-15% of the administered dose,
60 whereas CYP2A6 and CYP2B6 play a minor role in VPA metabolism [9]. Although CYP-
61 mediated pathways contribute to a minor part of VPA metabolism in adults (less than 20% of
62 the administered dose), the CYP-catalyzed oxidation may become the principal route of the
63 metabolism in those special cases when glucuronidation or mitochondrial β -oxidation
64 pathways are compromised or poorly developed, for example, in children. Shifting the
65 metabolic pathways may account for the age-related differences in the incidence of VPA-
66 induced adverse effects. i) Hepatic glucuronidation is known to be developmentally regulated.

67 UDP-glucuronyl transferases involved in VPA glucuronidation [10], are expressed under the
68 adult levels until sometime after 10-15 years of age [11,12]. *In vitro* glucuronide conjugation
69 of VPA has been demonstrated to be catalyzed by UGT1A6, UGT1A9 and UGT2B7 [10]; and
70 Guo et al. have confirmed the role of UGT1A6 *in vivo*; however, UGT2B7 seems to catalyze
71 VPA glucuronidation less efficiently [13]. ii) VPA and some of its metabolites are considered
72 to be the inhibitors of mitochondrial β -oxidation [14]. iii) CYP-dependent metabolism in
73 children exceeds adult activities, and decreases to adult levels by puberty [15]. As a
74 consequence, larger amount of VPA dose is liable to CYP2C9-dependent metabolism in
75 pediatric patients than in adults. Furthermore, the genetic and non-genetic factors, influencing
76 CYP2C9 activity, can increase the predisposition to VPA-induced serious adverse reactions;
77 thus, recognition of risk factors can contribute to the avoidance of adverse events.

78 There have been several clinical studies, investigating relationship between VPA
79 pharmacokinetics and patients' *CYP* genotypes, although clear evidence for the association
80 between VPA serum concentrations and *CYP2C9* genotype has been rarely provided [13,16].
81 Statistically significant, but relatively small differences in plasma concentrations of VPA have
82 been observed in patients with *CYP2C9**3 allele comparing to those with two wild type
83 alleles [16]. Although polymorphic *CYP* alleles result in non-functional CYP enzymes and
84 permanent poor metabolism, the individuals with functional wild type alleles may become
85 transient poor metabolizers as an effect of internal (e.g. diseases, hormonal status) or
86 environmental factors (e.g. nutrition, medication). This means that *CYP* genotype determines
87 the potential for the expression of functional or non-functional CYP enzyme. For example, a
88 patient with *CYP2C9**2/*2 or *CYP2C9**3/*3 basically displays poor metabolism of CYP2C9
89 substrates, whereas a subject carrying *CYP2C9**1/*1 possesses the potential for having
90 functional CYP2C9 enzyme. However, non-genetic factors, such as co-medications or co-
91 morbidities give rise to altered phenotypes. Thus, *CYP2C9**1/*1 genotype, predicted to be

92 translated to an extensive metabolizer phenotype, may be switched into poor metabolism due
93 to phenoconversion, which eventually influences the patient's response to VPA [17].
94 Furthermore, the genotype-phenotype mismatch results in more poor metabolizers than it
95 would be predicted from *CYP2C9* genotype.

96 A patient's CYP-status can be estimated by the evaluation of *CYP* genotypes and
97 current CYP expression. We have previously described a complex diagnostic system
98 (CYPtestTM) that can determine drug metabolizing capacity by combining *CYP* genotypes and
99 current CYP expression in leukocytes [18]. *CYP2C9* mRNA levels in leukocytes of those
100 subjects who do not carry loss-of-function mutations in *CYP2C9* gene was proven to reflect
101 the hepatic tolbutamide hydroxylation activity selective for *CYP2C9* [18]. A preliminary
102 *CYP2C9* genotyping for *CYP2C9*2* and *CYP2C9*3* can identify the genetically determined
103 poor metabolism of *CYP2C9* enzyme, and then *CYP2C9* expression in leukocytes of patients
104 with wild type alleles (*CYP2C9*1/*1*) can estimate a reduced or even increased *CYP2C9*
105 activity resulted by non-genetic variations. A patient carrying *CYP2C9*1/*1* genotype can be
106 assumed to be an extensive metabolizer and able to biotransform VPA more rapidly than
107 others carrying polymorphic *CYP2C9*2* or *CYP2C9*3* alleles. However, non-genetic factors
108 can modify the expression of the functional wild type alleles resulting in transient poor
109 metabolism similarly to those with non-functional polymorphic *CYP2C9* alleles. In the
110 present study, we investigated *CYP2C9*-status of pediatric patients younger than 15 years of
111 age and its influence on the steady-state serum concentrations of VPA as well as on patients'
112 dose-requirements. We attempted to provide evidence for that *CYP2C9* genotype is not the
113 only determinant factor in *CYP2C9* metabolizer status of a patient, but the expression rate of
114 the wild type gene can highly influence a patient's *CYP2C9* metabolizing capacity and his/her
115 response to a drug.

116

117 **Patients & methods**

118 • **Patients and sampling procedures**

119 Pediatric patients (N=50) suffering from epilepsy diagnosed with partial or generalized
120 seizures were enrolled in the study carried out at Heim Pál Children's Hospital and at the 2nd
121 Department of Pediatrics, Semmelweis University (Budapest, Hungary). We recruited novel
122 epileptic patients, younger than 15 years of age, who were CYP2C9 tested at the beginning of
123 antiepileptic therapy. The patients on non-VPA therapy or on multi-drug therapy were
124 excluded from the study. The patients were also excluded if their VPA therapy was
125 interrupted. The parents or representatives of each pediatric patient gave their informed
126 consent to participate in this study.

127 The patients' demographic data, as well as the details of anticonvulsant therapy were
128 recorded. The patients (boys/girls: 20/30) were at the average age of 6.75 years (range: 0.5 –
129 15 years), and all of them belonged to the Caucasian white population. Blood samples for
130 CYP2C9 testing were taken before the beginning of anticonvulsant therapy. The patients were
131 not given any other medication, but VPA as mono-therapy, and the target dose was adjusted
132 to the patients' body weight according to the clinical protocol [19]. The therapy was initiated
133 at low dosages (10-15 mg/kg), and the target doses were subsequently titrated until optimal
134 clinical response was achieved, generally within 5-10 days. Blood samples for drug assays
135 were taken two and four weeks after the beginning of VPA treatment. The sampling at the
136 second week was applied for checking VPA serum concentration, and the dose was modified
137 if the exposure exceeded the target range of VPA concentration. The serum levels measured at
138 the fourth week were considered to be the stable steady-state concentrations, whereas the
139 doses applied for the stable VPA concentrations were considered to be the maintenance doses.

140 • **CYP2C9 testing**

141 Patients' CYP2C9-status was determined by *CYP2C9* genotyping and by assaying

142 CYP2C9 expression in leukocytes before the beginning of VPA administration. Genomic
143 DNA and leukocytes were isolated from the samples of peripheral blood according to the
144 methods described by Temesvári et al. [18]. *CYP2C9* genotyping was carried out by
145 hydrolysis single nucleotide polymorphism analysis for *CYP2C9*2* and *CYP2C9*3* using
146 TaqMan Probes (BioSearch Technologies, Novato CA). For CYP2C9 expression, total RNA
147 was isolated from leukocytes, RNA was reverse-transcribed into single-stranded cDNA, and
148 then real-time PCR with human cDNA was performed using UPL probe for CYP2C9 (Roche
149 Diagnostics, Mannheim, Germany). The quantity of CYP2C9 mRNA relative to that of the
150 housekeeping gene glyceraldehyde 3-phosphate dehydrogenase was determined. Three
151 categories of CYP2C9 expression were applied to describe low, normal and high expressers.
152 The cutoff values for the CYP2C9 mRNA levels in leukocytes were previously established on
153 the basis of the cutoff values for the hepatic CYP2C9 activity (tolbutamide hydroxylation),
154 allowing a distinction between low, normal (medium) and high expressers (5×10^{-6} and
155 2.5×10^{-5} , respectively) [18].

156 • **Serum VPA assay**

157 The blood samples were taken before the patients were administered the morning dose.
158 The steady-state serum concentration of VPA was determined by the fluorescence
159 polarization immunoassay method (AxSYM Valproic Acid Assay, Abbott Laboratories, IL).
160 The VPA concentrations ranged between 40 and 100 $\mu\text{g/ml}$ were considered to be the
161 therapeutic levels [19].

162 • **Statistical analysis**

163 The serum concentration values of VPA were normalized by the dose and the body
164 weight, and expressed as $(\mu\text{g/ml}) \times (\text{mg dose/kg body weight})^{-1}$. The data of normalized VPA
165 concentrations and dose-requirements for the optimal therapeutic level in the groups with
166 various CYP2C9-statuses were expressed as the median (and range). It should be noted that

167 median values did not differ much (generally by 1-2% and always under 5%) from the mean
168 values. Between-group differences were calculated by the use of Kruskal-Wallis test followed
169 by Dunn's multiple comparisons test. A P value of <0.05 was considered statistically
170 significant.
171

172 **Results**

173 CYP2C9-status of pediatric patients

174 Of 50 pediatric patients aged between 0.5 and 15 years, all expressed at least one
175 functional *CYP2C9* allele, and 70% of patients carried *CYP2C9**1/*1 genotype (Table 1). The
176 patients with two loss-of-function alleles were not enrolled in the study, since they were on
177 non-VPA therapy. Fifteen patients (30%) carried one of the polymorphic variant alleles
178 (*CYP2C9**2 or *CYP2C9**3). The frequencies of *CYP2C9**2 and *CYP2C9**3 alleles in patients
179 (9% and 6%, respectively) were similar to those in Caucasian (white) populations (11% and
180 7%, respectively) [20,21]. CYP2C9 expression assays revealed that almost half of the patients
181 (46%, N=23) were normal CYP2C9 expressers, and substantial portion of the patients (54%,
182 N=27) were low expressers (Table 1). None of the children displayed high CYP2C9
183 expression. On the basis of CYP2C9-status (*CYP2C9* genotypes and CYP2C9 expression),
184 the patients were grouped into two main categories - homozygous wild (*CYP2C9**1/*1) and
185 heterozygous *CYP2C9**1/*mut* genotypes (*CYP2C9**1/*2 or *CYP2C9**1/*3), - and subdivided
186 into two subgroups: normal (medium) and low CYP2C9 expressers (Table 1). Although
187 patients with two wild type alleles are generally considered to be extensive metabolizers,
188 merely 12 children of 35 patients with *CYP2C9**1/*1 genotype were found to be normal
189 CYP2C9 expressers, whereas the other 23 patients were low expressers, predicting poor
190 CYP2C9 metabolism. Furthermore, the group of patients with heterozygous *CYP2C9**1/*mut*
191 genotypes comprised both low and normal CYP2C9 expressers (4 and 11 patients,
192 respectively). It is not surprising, since the mutant alleles are transcribed into CYP2C9
193 mRNA; however, their expression rates are modified by non-genetic factors, such as nutrition,
194 food additives, or hormonal status, similarly to the wild type allele. Co-medication as a non-
195 genetic factor can be excluded, since the children on multi-drug therapy were not enrolled in
196 the present study.

197 Patients' VPA exposure and dose-requirement

198 The statistical analysis displayed significant association between the patients'
199 CYP2C9-status and the steady-state serum levels of VPA normalized by the dose and the
200 body weight. The normalized serum VPA concentrations were significantly lower in the
201 normal expresser patients with *CYP2C9*1/*1* genotype ($2.12 (\mu\text{g/ml}) \times (\text{mg dose/kg bw})^{-1}$)
202 than in low expressers ($5.13 (\mu\text{g/ml}) \times (\text{mg dose/kg bw})^{-1}$) or in patients carrying any
203 polymorphic *CYP2C9* alleles (*CYP2C9*2* or *CYP2C9*3*) ($4.33 (\mu\text{g/ml}) \times (\text{mg dose/kg bw})^{-1}$
204 for normal CYP2C9 expressers and $5.54 (\mu\text{g/ml}) \times (\text{mg dose/kg bw})^{-1}$ for low expressers)
205 (Figure 1). The low expressers and the patients with polymorphic *CYP2C9* alleles showed
206 about 2- to 3-fold higher normalized serum VPA levels as compared to normal expresser
207 patients carrying *CYP2C9*1/*1* genotype. The difference in normalized serum concentrations
208 was not statistically significant between the patients with heterozygous genotypes
209 (*CYP2C9*1/*2* or *CYP2C9*1/*3*) and those low expressers with two functional alleles
210 (*CYP2C9*1/*1*). Moreover, no significant difference in normalized serum levels was
211 observed between normal and low expressers with heterozygous *CYP2C9* genotypes.

212 According to the clinical practice, VPA serum concentrations ranged between 40 and
213 100 $\mu\text{g/ml}$ are considered to be therapeutically optimal in the management of epilepsy [19].
214 The low expresser patients or subjects with heterozygous genotypes required significantly
215 lower dose of VPA for the optimal serum level than normal expressers carrying
216 *CYP2C9*1/*1* genotype (Figure 2). The dose-requirement of VPA for the target serum level
217 was similar for the low expressers and for the patients carrying polymorphic *CYP2C9* alleles
218 (17.8 mg/kg for low expressers carrying *CYP2C9*1/*1*; 16.7 mg/kg for normal expressers
219 with heterozygous genotype; 13.8 mg/kg for low expressers with heterozygous genotype).
220 The conventional clinical practice is to target the VPA dose of 30 to 40 mg/kg in children.
221 The conventional dosing approach was appropriate for normal CYP2C9 expresser patients

222 with *CYP2C9*1/*1* genotype, comprising 24% of the children in the study. The *CYP2C9*
223 genotype-controlled VPA dosing would have targeted reduced VPA dose for 30% of the
224 patients, for those carrying heterozygous *CYP2C9*1/mut* genotypes. However, low expressers
225 with *CYP2C9*1/*1* genotype also required reduced VPA dose for the optimal serum
226 concentration. *CYP2C9* phenoconversion substantially increased the number of children (to
227 76%) on reduced VPA dose.

228 Multiple comparison analysis showed that *CYP2C9*-status (*CYP2C9* genotype and
229 *CYP2C9* expression) influenced the serum concentrations of VPA as well as the dose-
230 requirements for the optimal serum concentration in pediatric patients. However, low
231 *CYP2C9* expression in patients with homozygous wild genotype seemed to display similar
232 effects on VPA exposure and dose-requirement to those carrying polymorphic *CYP2C9*
233 alleles (*CYP2C9*2* or *CYP2C9*3*). Consistently, the serum VPA concentration and dose-
234 requirement of the children carrying two wild type *CYP2C9* alleles (*CYP2C9*1/*1*) were
235 found to be influenced by the *CYP2C9* expression, whereas loss-of-function mutations in
236 *CYP2C9* gene resulted in poor metabolism of VPA independently on the degree of *CYP2C9*
237 expression.

238

239 **Discussion**

240 Drug metabolizing capacity highly influences the patient's response to a drug and the
241 risk of side effects. Genetic and non-genetic factors in drug metabolism give rise to
242 substantial interindividual variability in clinical response of drugs, assigning the patient
243 populations into three groups: poor, intermediate and extensive metabolizers [22]. By
244 recognizing individual differences, personalized medication can help to avoid the therapeutic
245 failure or potential adverse reactions [23]. Pharmacogenetic assays can determine poor drug
246 metabolism by genotyping, identifying non-functional drug metabolizing enzymes [22], but

247 do not provide reliable information about the drug metabolizing capacity of patients who do
248 not have loss-of-function mutations. Non-genetic factors, such as age, diseases, nutrition, or
249 co-medication, can transiently modulate patient's drug metabolizing capacity. Developmental
250 regulation of drug metabolizing enzymes is known to contribute to age-related differences in
251 drug efficacy or toxicity between children and adults [24]. CYP-dependent metabolism is
252 generally low at birth (about 50-70% of adult levels); however, CYP enzyme activities exceed
253 the adult values by the age of 2 years and decrease by puberty [15]. In contrast, the drug-
254 conjugating activities of several UDP-glucuronyl transferases are low or negligible around
255 birth, slightly increasing, but not reaching the adult levels until puberty [11,12]. Concerning
256 VPA, the major metabolic pathway in adults, glucuronidation can shift toward CYP-
257 dependent oxidation in pediatric patients because of reduced glucuronidation ability. On the
258 other hand, chronic administration of VPA leads to the inhibition of β -oxidation pathway of
259 VPA metabolism, assigning a prominent role in the metabolism to CYP enzymes [14,25].

260 CYP2C9, the main catalyst of CYP-dependent metabolism of VPA, is highly
261 polymorphic with *CYP2C9*2* and *CYP2C9*3* being identified as the most frequent variants in
262 Caucasian population [20,21]. These loss-of-function mutations have been reported to be less
263 active in *in vitro* metabolism of VPA than the wild type allele [26]. The influence of
264 *CYP2C9*3* allele on VPA plasma levels was displayed in Chinese patients [16]; however, the
265 moderate increase in normalized VPA concentrations in the patients carrying *CYP2C9*1/*3*
266 may be attributed to the facts that the authors took neither the CYP2C9 expression nor the
267 age-related differences in VPA metabolism into account. Predicting drug metabolizing
268 phenotype from genotype seems to be highly complex even in the case of non-inducible
269 enzymes, such as CYP2D6 [27]; thus, inferring a patient's VPA metabolizing phenotype
270 merely from *CYP2C9* genotype can easily lead to false interpretations. We have previously
271 reported a more than 60-fold difference in CYP2C9 mRNA levels in human liver tissues

272 which means that transient poor metabolizers (low CYP2C9 expressers) exist in the group of
273 patients carrying *CYP2C9*1/*1* genotype [18]. Thus, not only genetic, but non-genetic
274 variations of CYP2C9 are of particular importance in the evaluation of patients' CYP2C9-
275 status. The pediatric patients in the present study was divided into two *CYP2C9* genotype
276 groups (*CYP2C9*1/*1* and *CYP2C9*1/mut*), although both groups comprised low and normal
277 CYP2C9 expresser children. Patients carrying *CYP2C9*1/*1* genotype are generally assumed
278 to be extensive metabolizers; however, *CYP2C9* genotype can be converted to a phenotype
279 different from that would be predicted from the genotype. Hence, the normal expresser
280 children carrying *CYP2C9*1/*1* were basically expected to display extensive metabolizer
281 phenotype, whereas low expressers with *CYP2C9*1/*1* genotype were assumed to behave as
282 poor metabolizers. It should be noted that the mutant *CYP2C9* alleles are translated into non-
283 functional CYP2C9 protein, resulting in poor metabolism, even if they are expressed at
284 normal levels.

285 The present study, involving pediatric patients younger than 15 years of age, has
286 clearly demonstrated that normalized serum concentrations of VPA were associated with
287 patients' CYP2C9-status determined by *CYP2C9* genotyping and CYP2C9 expression
288 analysis. The children with heterozygous *CYP2C9* genotype (*CYP2C9*1/*2* or
289 *CYP2C9*1/*3*) were found to be poor VPA metabolizers, presenting high serum VPA
290 concentrations and requiring low VPA dose. Although the patients carrying two wild type
291 alleles (*CYP2C9*1/*1*) could be supposed to have functional CYP2C9 enzyme, their VPA
292 metabolizing capacity was influenced by CYP2C9 expression. The low expresser patients
293 carrying *CYP2C9*1/*1* showed as high serum VPA concentrations and required as low dose
294 for the optimal VPA levels as those poor VPA metabolizers with heterozygous *CYP2C9*
295 genotype, whereas the normal expressers with two wild type alleles appeared to be more
296 active in VPA metabolism, presenting significantly lower VPA serum levels.

297 Phenoconversion of patients' genotype are generally explained by the fact that external or
298 internal factors, notably co-medications, nutrition, diseases, inflammation or hormonal status,
299 modify the expression or the function of drug metabolizing enzyme. The co-administration of
300 VPA and antiepileptic drugs known to be CYP2C9 inducers (e.g. phenytoin, phenobarbital, or
301 carbamazepine) results in increased CYP2C9 expression and enhanced VPA metabolizing
302 capacity of patients on multi-drug therapy. Amini-Shirazi et al. have reported that the
303 concomitant treatment of patients with VPA and CYP2C9 inducers increased the formation
304 rate of 4-ene-VPA metabolite comparing to the patients on VPA monotherapy [28].
305 Nevertheless, the patients with distinct CYP2C9 expression occurred in both *CYP2C9*1/*1*
306 and *CYP2C9*1/mut* genotype groups of the patients involved in our study that could not be a
307 consequence of co-medications, because the patients on multi-drug therapy were excluded
308 from the study. The ratio of low expresser patients was unusually high, more than half of the
309 children involved displayed low CYP2C9 expression, predicting some suppressive factors in
310 the background. The significant release of pro-inflammatory cytokines observed in epileptic
311 patients following seizures seems to be a logical explanation, since the expression of drug
312 metabolizing enzymes is down-regulated as a response to the increasing levels of the acute
313 phase proteins, resulting in substantial impairment of drug metabolism [29-31]. The down-
314 regulation of CYP2C9 by the pro-inflammatory cytokines, such as IL-6 and IL-1 β , is
315 proposed to be mediated by the repression of the nuclear receptors (pregnane X receptor and
316 constitutive androstane receptor) involved in CYP2C9 expression [32,33]. The
317 phenoconversion of other drug metabolizing enzymes, including CYP2C19, CYP2D6,
318 CYP3A4 or NAT2, has also been observed in patients suffering from HIV, cancer or liver
319 disease [34-38]; however, the present work was the first study that provided evidence for the
320 phenoconversion and marked repression of VPA metabolizing CYP2C9 in epileptic children.

321 The novel findings of the present study demonstrated that the normalized VPA serum
322 concentrations in pediatric patients were influenced by the patients' CYP2C9-status
323 determined not only by the genetic variability of *CYP2C9*, but also by CYP2C9 expression.
324 The pediatric patients with various CYP2C9-statuses required different doses of VPA for the
325 optimal serum concentrations. The low CYP2C9 expressers and patients with mutated
326 *CYP2C9* alleles (*CYP2C9*2* or *CYP2C9*3*) required approximately half of the dose for
327 normal (or medium) expressers with *CYP2C9*1/*1* genotype (14-18 mg/kg vs 33 mg/kg). As
328 a consequence, CYP2C9-status can guide the appropriate targeting of VPA dose at the
329 beginning of anticonvulsant therapy. The VPA therapeutic strategy for the normal CYP2C9
330 expressers with *CYP2C9*1/*1* genotype can follow the conventional therapy (target VPA
331 dose of 30-40 mg/kg) [19]. The low expressers and patients with mutated *CYP2C9* alleles
332 (*CYP2C9*2* or *CYP2C9*3*) require substantial modification of VPA dose (14-18 mg/kg) for
333 achieving the desired target serum concentrations. Despite the small size of genotype groups,
334 our results would raise the concerns that the conventional clinical practice may overdose more
335 than 70% of the pediatric patients, and *CYP2C9* genotype-controlled VPA would also
336 increase the misdosing risk in about two third of patients carrying *CYP2C9*1/*1*. It can be
337 concluded, that the phenoconversion of *CYP2C9* limits the predictive value of *CYP2C9*
338 genotyping in optimizing VPA therapy.

339

340 **Conclusion & future perspective**

341 The optimal serum concentration of VPA is strongly influenced by the patients' VPA
342 metabolizing capacity which is also critical to avoid the therapeutic failure or toxicity of VPA.
343 Glucuronide conjugation has been demonstrated to be the major metabolic pathway of VPA in
344 adults; however, the influence of genetic variants of UGT isoenzymes on dose-requirement
345 and treatment outcome remains elusive because of the conflicting results obtained from small

346 cohort studies. According to our knowledge, CYP-mediated oxidation is not the major route
347 of VPA metabolism in adults; however, our present work clearly demonstrated that CYP2C9
348 played a prominent role in children younger than 15 years of age. CYP2C9 pathway may be
349 assumed to be more dominant in neonates and infants because of their strongly deficient
350 glucuronidation ability, and focusing on younger pediatric patients may provide better
351 understanding of the increased risk of VPA-induced toxicity in this vulnerable population.

352 Comparing to the conventional clinical practice, the *CYP2C9* genotype-based
353 medication may bring some benefit to children on VPA therapy; however, metabolic activity
354 of CYP2C9 is often overestimated by the prediction from the patient's *CYP2C9* genotype.
355 The major source of overestimation is CYP2C9 phenoconversion that can be attributed to the
356 *CYP2C9* down-regulation by cytokines in epilepsy. Thus, prospective investigation of
357 pediatric patients' genetic and non-genetic variations in CYP2C9 allows prediction of
358 potential 'poor metabolizers' carrying *CYP2C9* alleles with loss-of-function mutations or
359 displaying low CYP2C9 expression. CYP2C9-status controlled medication may facilitate the
360 improvement of the individual VPA therapy, leading to the dosage optimization for a more
361 effective therapy, and minimizing the risk of severe side effects. Further prospective studies
362 evaluating the clinical outcome are supposed to reveal the benefit of CYP2C9-status
363 controlled VPA therapy over conventional antiepileptic therapy.

364

365

366 **Executive summary**

367 Background

- 368 • The mainstay of antiepileptic therapy is valproic acid (VPA), which is well-tolerated
369 by most of the patients; however, the risk of serious side effects, such as
370 hepatotoxicity or hematologic disorders, is increased in pediatric patients.

- 371 • In adults, the major metabolic pathways of VPA are glucuronidation and
372 mitochondrial β -oxidation, whereas cytochrome P450 (CYP)-dependent oxidation has
373 minor role in VPA metabolism.
- 374 • In children, CYP2C9-catalyzed oxidation may become the principal route of the
375 metabolism which may lead to age-related differences in the incidence of adverse
376 reactions.
- 377 • Although genetic polymorphism of *CYP2C9* may explain some interindividual
378 differences in pharmacokinetics and dose-requirement of VPA, non-genetic factors
379 give rise to low or even high CYP2C9 expression, modifying the patient's VPA
380 metabolizing capacity.

381 Findings & conclusion

- 382 • *CYP2C9* genotyping of pediatric patients was able to predict VPA poor metabolism in
383 approximately 30% of patients.
- 384 • CYP2C9 expression was down-regulated in more than 50% of children probably due
385 to the cytokine release in epilepsy; thus, inferring the patients' VPA metabolizing
386 phenotype merely from *CYP2C9* genotype resulted in false prediction.
- 387 • Although the VPA therapeutic strategy for the normal CYP2C9 expressers with
388 *CYP2C9**1/*1 genotype can follow the conventional therapy (target VPA dose of 30-
389 40 mg/kg), the low expressers and patients carrying loss-of-function mutation in
390 *CYP2C9* gene require substantial modification of VPA dose (14-18 mg/kg) for
391 achieving the desired target serum concentrations.
- 392 • CYP2C9-status controlled VPA therapy can contribute to the avoidance of misdosing
393 and potential adverse reactions in pediatric patients.

394

395 **References**

- 396 [1] Fogarasi A, Neuwirth M, Gyorsok Z, Czirják S, Vajda J, Bognár L. Epilepsy surgery in
397 childhood: theory and practice. *Orv. Hetil.* 144(48), 2359-2365 (2003).
- 398 [2] Chateauvieux S, Morceau F, Dicato M, Diederich M. Molecular and therapeutic
399 potential and toxicity of valproic acid. *J. Biomed. Biotechnol.* 2010, pii: 479364 (2010).
- 400 [3] Nanau RM, Neuman MG. Adverse drug reactions induced by valproic acid. *Clin.*
401 *Biochem.* 46(15), 1323-1338 (2013).
- 402 [4] Silva MF, Aires CC, Luis PB, et al. Valproic acid metabolism and its effects on
403 mitochondrial fatty acid oxidation: a review. *J. Inherit. Metab. Dis.* 31(2), 205-216
404 (2008).
- 405 [5] Peterson GM, Naunton M. Valproate: a simple chemical with so much to offer. *J. Clin.*
406 *Pharm. Ther.* 30(5), 417-421 (2005).
- 407 [6] Ghodke-Puranik Y, Thorn CF, Lamba JK, et al. Valproic acid pathway:
408 pharmacokinetics and pharmacodynamics. *Pharmacogenet. Genomics.* 23(4), 236-241
409 (2013).
- 410 [7] Rettie AE, Boberg M, Rettenmeier AW, Baillie TA. Cytochrome P-450-catalyzed
411 desaturation of valproic acid in vitro. Species differences, induction effects, and
412 mechanistic studies. *J. Biol. Chem.* 263(27), 13733-13738 (1988).
- 413 [8] Gao S, Miao H, Tao X, et al. LC-MS/MS method for simultaneous determination of
414 valproic acid and major metabolites in human plasma. *J. Chromatogr. B Analyt.*
415 *Technol. Biomed. Life Sci.* 879(21), 1939-1944 (2011).
- 416 [9] Kiang TKL, Ho PC, Anari MR, Tong V, Abbott FS, Chang TKH. Contribution of
417 CYP2C9, CYP2A6, and CYP2B6 to valproic acid metabolism in hepatic microsomes
418 from individuals with the *CYP2C9*1/*1* genotype. *Toxicol. Sci.* 94(2), 261-271 (2006).

- 419 [10] Sakaguchi K, Green M, Stock N, Reger TS, Zunic J, King C. Glucuronidation of
420 carboxylic acid containing compounds by UDP-glucuronosyltransferase isoforms. *Arch.*
421 *Biochem. Biophys.* 424(2), 219-225 (2004).
- 422 [11] McCarver DG, Hines RN. The ontogeny of human drug-metabolizing enzymes: phase II
423 conjugation enzymes and regulatory mechanisms. *J. Pharmacol. Exp. Ther.* 300(2),
424 361-366 (2002).
- 425 [12] Strassburg CP, Vogel A, Kneip S, et al. Developmental aspects of human hepatic drug
426 glucuronidation in young children and adults. *Gut* 50(2), 259-265 (2002).
- 427 [13] Guo Y, Hu C, He X, Qiu F, Zhao L. Effects of *UGT1A6*, *UGT2B7*, and *CYP2C9*
428 genotypes on plasma concentrations of valproic acid in Chinese children with epilepsy.
429 *Drug Metab. Pharmacokinet.* 27(5), 536-542 (2012).
- 430 [14] Ponchaut S, van Hoof F, Veitch K. In vitro effects of valproate and valproate
431 metabolites on mitochondrial oxidations. Relevance of CoA sequestration to the
432 observed inhibitions. *Biochem. Pharmacol.* 43(11), 644-647 (1992).
- 433 [15] Stewart CF, Hampton EM. Effect of maturation on drug disposition in pediatric
434 patients. *Clin. Pharm.* 6(7), 548-564 (1987).
- 435 [16] Tan L, Yu JT, Sun YP, Ou JR, Song JH, Yu Y. The influence of cytochrome oxidase
436 *CYP2A6*, *CYP2B6*, and *CYP2C9* polymorphisms on the plasma concentrations of
437 valproic acid in epileptic patients. *Clin. Neurol. Neurosurg.* 112(4), 320-323 (2010).
- 438 [17] Shah RR, Smith RL. Addressing phenoconversion: the Achilles' heel of personalized
439 medicine. *Br. J. Clin. Pharmacol.* doi: 10.1111/bcp.12441 (2014)
- 440 [18] Temesvári M, Kóbori L, Paulik J, Sárváry E, Belic A, Monostory K. Estimation of
441 drug-metabolizing capacity by cytochrome P450 genotyping and expression. *J.*
442 *Pharmacol. Exp. Ther.* 341(1), 294-305 (2012).

- 443 [19] Guerrini R. Valproate as a mainstay of therapy for pediatric epilepsy. *Paediatr. Drugs*
444 8(2), 113-129 (2006).
- 445 [20] Zanger UM, Turpeinen M, Klein K, Schwab M. Functional pharmacogenetics/genomics
446 of human cytochromes P450 involved in drug biotransformation. *Anal. Bioanal. Chem.*
447 392(6), 1093-1108 (2008).
- 448 [21] Kurose K, Sugiyama E, Saito Y. Population differences in major functional
449 polymorphisms of pharmacokinetics/pharmacodynamics-related genes in Eastern
450 Asians and Europeans: implications in the clinical trials for novel drug development.
451 *Drug Metab. Pharmacokinet.* 27(1), 9-54 (2012).
- 452 [22] Ingelman-Sundberg M. Pharmacogenetics: an opportunity for a safer and more efficient
453 pharmacotherapy. *J. Intern. Med.* 250(3), 186-200 (2001).
- 454 [23] Wilke RA, Musana AK, Weber WW. Cytochrome P450 gene-based drug prescribing
455 and factors impacting translation into routine clinical practice. *Person. Med.* 2(3), 213-
456 224 (2005).
- 457 [24] Anderson GD. Children versus adults: pharmacokinetic and adverse-effect differences.
458 *Epilepsia* 43(Suppl 3), 53-59 (2002).
- 459 [25] Fromenty B, Pessayre D. Inhibition of mitochondrial beta-oxidation as a mechanism of
460 hepatotoxicity. *Pharmacol. Ther.* 67(1), 101-154 (1995).
- 461 [26] Ho PC, Abbott FS, Zanger UM, Chang TKH. Influence of *CYP2C9* genotypes on the
462 formation of a hepatotoxic metabolite of valproic acid in human liver microsomes.
463 *Pharmacogenomics J.* 3(6), 335-342 (2003).
- 464 [27] Hicks JK, Swen JJ, Gaedigk A. Challenges in CYP2D6 phenotype assignment from
465 genotype data: a critical assessment and call for standardization. *Curr. Drug Metab.*
466 15(2), 218-232 (2014).

- 467 [28] Amini-Shirazi N, Ghahremani MH, Ahmadkhaniha R, et al. Influence of *CYP2C9*
468 polymorphism on metabolism of valproate and its hepatotoxic metabolite in Iranian
469 patients. *Toxicol. Mech. Methods*. 20(8), 452-457 (2010).
- 470 [29] Yu N, Di Q, Hu Y, Zhang YF, et al. A meta-analysis of pro-inflammatory cytokines in
471 the plasma of epileptic patients with recent seizure. *Neurosci. Lett*. 514(1): 110-115
472 (2012).
- 473 [30] Uludag IF, Bilgin S, Zorlu Y, Tuna G, Kirkali G. Interleukin-6, interleukin-1 beta and
474 interleukin-1 receptor antagonist levels in epileptic seizures. *Seizure*. 22(6): 457-461
475 (2013).
- 476 [31] Aitken AE, Morgan ET. Gene-specific effects of inflammatory cytokines on
477 cytochrome P450 2C, 2B6 and 3A4 mRNA levels in human hepatocytes. *Drug Metab.*
478 *Dispos*. 35(9): 1687-1693 (2007).
- 479 [32] Pascussi JM, Gerbal-Chaloin S, Pichard-Garcia L, et al. Interleukin-6 negatively
480 regulates the expression of pregnane X receptor and constitutively activated receptor in
481 primary human hepatocytes. *Biochem. Biophys. Res. Commun*. 274(3): 707-713 (2000).
- 482 [33] Pascussi JM, Dvorák Z, Gerbal-Chaloin S, Assenat E, Maurel P, Vilarem MJ.
483 Pathophysiological factors affecting CAR gene expression. *Drug Metab. Rev*. 35(4):
484 255-268 (2003).
- 485 [34] Jones AE, Brown KC, Werner RE, et al. Variability in drug metabolizing enzyme
486 activity in HIV-infected patients. *Eur. J. Clin. Pharmacol*. 66(5): 475-485 (2010).
- 487 [35] O'Neil WM, Gilfix BM, DiGirolamo A, Tsoukas CM, Wainer IW. N-acetylation among
488 HIV-positive patients and patients with AIDS: when is fast, fast and slow, slow? *Clin.*
489 *Pharmacol. Ther*. 62(3): 261-271 (1997).

- 490 [36] Helsby NA, Lo WY, Sharples K, et al. CYP2C19 pharmacogenetics in advanced cancer:
491 compromised function independent of genotype. *Br. J. Cancer.* 99(8): 1251-1255
492 (2008).
- 493 [37] Burns KE, Goldthorpe MA, Porteus F, Browett P, Helsby NA. CYP2C19 genotype-
494 phenotype discordance in patients with multiple myeloma leads to an acquired loss of
495 drug-metabolising activity. *Cancer Chemother. Pharmacol.* 73(3): 651-655 (2014).
- 496 [38] Rost KL, Brockmöller J, Esdorn F, Roots I. Phenocopies of poor metabolizers of
497 omeprazole caused by liver disease and drug treatment. *J. Hepatol.* 23(3): 268-277
498 (1995).
- 499
- 500

501 **Reference annotations**

502

503 [16] Tan L, Yu JT, Sun YP, Ou JR, Song JH, Yu Y. The influence of cytochrome oxidase
504 CYP2A6, CYP2B6, and CYP2C9 polymorphisms on the plasma concentrations of valproic
505 acid in epileptic patients. *Clin. Neurol. Neurosurg.* 112(4), 320-323 (2010).

506 **Evaluates the role of various CYP alleles (e.g. *CYP2C9*3*) in VPA pharmacokinetics

507 [17] Shah RR, Smith RL. Addressing phenoconversion: the Achilles' heel of personalized
508 medicine. *Br. J. Clin. Pharmacol.* doi: 10.1111/bcp.12441 (2014)

509 **Reviews the main sources of phenoconversion

510 [19] Guerrini R. Valproate as a mainstay of therapy for pediatric epilepsy. *Paediatr. Drugs*
511 8(2), 113-129 (2006).

512 *Reviews the principles and clinical practice of VPA therapy

513 [24] Anderson GD. Children versus adults: pharmacokinetic and adverse-effect differences.
514 *Epilepsia* 43(Suppl 3), 53-59 (2002)

515 *Reviews the differences in pharmacokinetics and adverse effects of antiepileptic drugs
516 between children and adults

517 [26] Ho PC, Abbott FS, Zanger UM, Chang TKH. Influence of CYP2C9 genotypes on the
518 formation of a hepatotoxic metabolite of valproic acid in human liver microsomes.
519 *Pharmacogenomics J.* 3(6), 335-342 (2003).

520 ** Evaluates the effect of CYP2C9 genetic polymorphism on the metabolism of VPA

521 [28] Amini-Shirazi N, Ghahremani MH, Ahmadkhaniha R, et al. Influence of CYP2C9
522 polymorphism on metabolism of valproate and its hepatotoxic metabolite in Iranian patients.
523 *Toxicol. Mech. Methods.* 20(8), 452-457 (2010).

524 **Evaluates the impact of concomitant treatment with CYP2C9 inducers on VPA
525 metabolism

526 **Acknowledgement**

527 The authors are indebted to Tímea Zentai for her skillful assistance in this study.

528

529 **Conflict of interest & Financial disclosure**

530 The study was supported by the grant from the Hungarian Research Fund (OTKA K104459)

531 and by the grants from the National Development Agency and the European Union (Grants

532 GOP-1.3.1-11/B-2011-0042 and GOP-1.1.1-11-2012-0027). The authors have no other

533 relevant affiliations or financial involvement with any organization or entity with a financial

534 interest in or financial conflict with the subject matter or materials discussed in the manuscript

535 apart from those disclosed. The authors declare that there are no personal conflicts of interest.

536

537 **Ethical conduct of research**

538 CYPtesting of the patients was approved by the Hungarian Committee of Science and

539 Research Ethics. The study was performed under the regulation of Act CLIV of 1997 on

540 Health and of the decree 23/2002 of the Minister of Health of Hungary, and in accordance

541 with the declaration of Helsinki. The representatives of each patient gave their informed

542 consent to participate in this study.

543

544

545 Table 1. Demographic data of patients with various CYP2C9-statuses
 546

CYP2C9-status		Number of patients	Age (year)*	Body weight (kg)*	Boys/girls
<i>CYP2C9*1/*1</i>	Normal expressers	12	4 (0.5 – 15)	25 (6 – 60)	4/8
	Low expressers	23	7 (1.5 – 15)	27 (14 – 65)	10/13
<i>CYP2C9*1/mut</i>	Normal expressers	11	4 (3 – 14)	19 (14 – 52)	5/6
	Low expressers	4	7.5 (4 -15)	22.5 (15 – 60)	1/3
Total		50	6.75 (0.5 – 15)	21.5 (6 – 65)	20/30

547 *: median (range); *CYP2C9*1/mut*: *CYP2C9*1/*2* or *CYP2C9*1/*3*
 548

549

550 **Figure legends**

551

552 Figure 1. Serum concentrations of valproic acid in patients with various CYP2C9-statuses.

553 The serum concentrations were measured four weeks after the beginning of valproic
554 acid therapy.

555 *CYP2C9*1/mut*: heterozygous *CYP2C9* genotype (*CYP2C9*1/*2* or *CYP2C9*1/*3*);
556 normal: normal (medium) CYP2C9 expressers; low: low CYP2C9 expressers; bw: body
557 weight; *: significant difference ($P < 0.05$); solid line: median of the groups

558

559 Figure 2. Valproic acid dose required for the therapeutic serum concentrations in patients with
560 various CYP2C9-statuses.

561 *CYP2C9*1/mut*: heterozygous *CYP2C9* genotype (*CYP2C9*1/*2* or *CYP2C9*1/*3*);
562 normal: normal (medium) CYP2C9 expresser; low: low CYP2C9 expresser; *:
563 significant difference ($P < 0.05$); ns: not significant

564

565
566
567
568
569
570
571
572
573
574
575
576
577
578
579
580
581
582
583

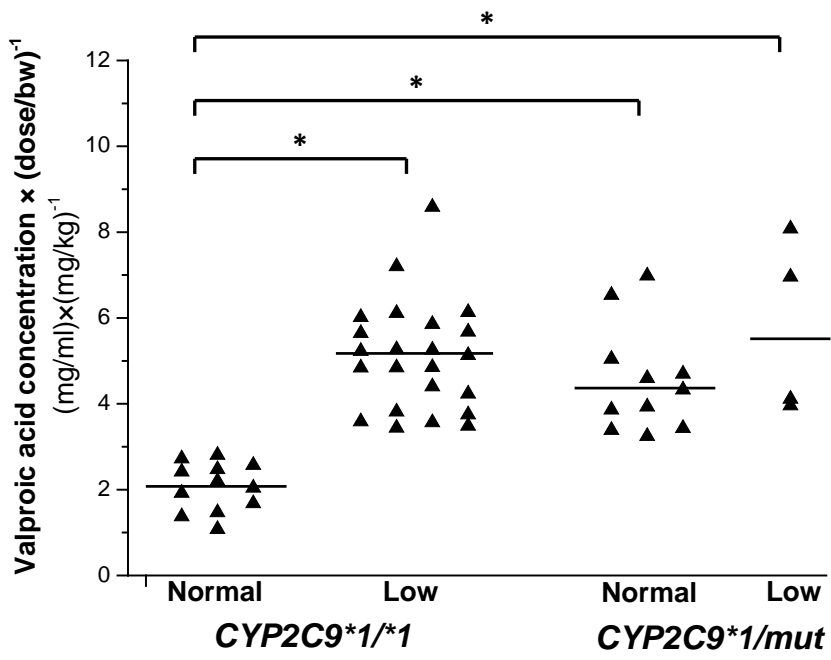


Figure 1

584

585

586

587

588

589

590

591

592

593

594

595

596

597

598

599

600

601 Figure 2

602

