

OLD AND NEW NEUROENDOCRINE MOLECULES: SOMATOSTATIN, CYSTEAMINE, PANTETHINE AND KYNURENINE

László VÉCSEI^{1,2}, Zoltán HORVÁTH¹, Bernadett TUKA²

¹Department of Neurology, Faculty of Medicine, University of Szeged, Szeged ²MTA-SZTE Neuroscience Research Group, University of Szeged, Szeged

RÉGI ÉS ÚJ NEUROENDOKRIN MOLEKULÁK: SZOMATOSZTATIN, CISZTEAMIN, PANTETHIN ÉS KINURENIN

Vécsei L, MD, DSc; Horváth Z, MD; Tuka B, MSc Ideggyogy Sz 2014;67(3–4):79–84.

The aim of this review is to commemorate Hans Selye, endocrinologist, the most famous researchers of stress and to briefly summarize the major features of somatostatin (SST), cysteamine (CysA) and patethine (PAN) in neuroendocrinological aspect, which are closely related to his scientific work. In addition, some metabolites of kynurenine pathway (KP) were also mentioned in this paper, as new, possible target molecules in neuroendocrinology.

R. Guillemin and A. V. Schally were the main pioneers of the discovery of SST in the 1970's. SST primarily is known as an inhibitor of growth hormone secretion and additionally reduces the gastric acid and pepsin release and also the gastroduodenal mucosal blood flow. These effects are very important in the pathophysiology of peptic ulcer bleeding, which is related to the CysA-evoked perforating duodenal ulcer experimental stress model in rats developed by Selye and Szabo. CysA is a naturally occurring duodenal ulcerogen, which depletes SST in the gastric mucosa and certain brain regions. Furthermore, in addition to depleting SST, CysA also causes adrenocortical necrosis, suggesting an interaction between the central/peripheral nervous system and the neuroendocrine system. The antioxidant PAN, formulated besides the CysA, has similar effects: it attenuates the levels of SST and prolactin in the cerebral cortex and hypothalamus through the accumulation of CysA within cells throughout the body. As new perspectives the KP may be involved in the modulation of neuroendrocrine processes: different agonists and antagonists of glutamate receptors regulate the hypothalamic-pituitary-adrenal axis and kynurenic acid augments the anxiolytic stress responses in neonatal chicks. The pro-inflammatory cytokine-induced and the toxic heavy oil contaminations-evoked alterations in the KP indirectly contribute to the development of neuroendocrine disorders.

In summary, there have been highly important developments in neuroendocrinology since the early findings of Selye. Although there are as yet relatively few data about the potential role of kynurenines in neuroendocrinology, the results already achieved are extremely noteworthy and immensely promising.

Keywords: somatostatin, cysteamine, pantethine, kynurenine

Az összefoglaló célja, hogy megemlékezzünk Selye Jánosról, az endokrinológusról, a stressz leghíresebb kutatójáról, és röviden összefoglaljuk a szomatosztatin (SST), a ciszteamin (CysA) és a pantethin (PAN) legfontosabb jellemzőit neuroendokrinológiai szempontból, amelyek szoros összefüggésben álllnak tudományos munkájával. Emellett megemlítjük a kinurenin (KP) -útvonal néhány metabolitját is mint a neuroendokrinológia néhány lehetséges célmolekuláját. R. Guillemin és A. V. Schally jártak elől az SST felfedezésében az 1970-es években. Az SST-t főként a növekedésihormon-elválasztás inhibitoraként ismerjük, emellett csökkenti a gyomosav és a pepszin felszabadulását és a gastroduodenalis nyálkahártya véráramlását. Ezek a hatások nagyon fontosak a peptikus fekély vérzésének kórélettanában, ami összefügg a CysA által kiváltott perforáló duodenalis fekély kísérletes stresszmodelljével, patkányban, amit Selye és Szabó fejlesztettek ki. A CysA a természetben előforduló duodenalis ulcerogen anyag, ami SST-depléciót okoz a gyomornyálkahártyában és egyes agyi régiókban. Az SST-depléció mellett a CysA adrenocorticalis necrosist is okoz, ami kölcsönhatásra utal a centrális/peri fériás idegrendszer és a neuroendokrin rendszer között. A CysA mellett képződő antioxidáns PAN hatása hasonló: csökkenti az SST és a prolaktin szintjét az agykéregben és a hypothalamusban az által, hogy a CysA testszerte felhalmozódik a sejteken belül. Új szempontként a KP részt vehet a neuroendokrin folyamatok modulációjában: a glutamátreceptorok különböző agonistái és antagonistái szabályozzák a hypothalamus-hypophysis-mellékvese tengelyt, és a kinurénsav fokozza újszülött csibékben az anxiolyticus stresszválaszt. A KP proinflammatorikus citokin által indukált és a toxikus nehézolaj-szennyeződés által kiváltott változásai közvetetten hozzájárulnak a neuroendokrin zavarokhoz. Osszességében nagyon fontos fejlődés ment végbe a neuroendokrinológiában Selye első eredményei óta. Bár még viszonylag kevés adatunk van a kinureninek potenciális szerepéről a neuroendokrinológiában, a már elért eredmények különösen értékesek és nagyon ígéretesek.

Kulcsszavak: szomatosztatin, ciszteamin, pantethin, kinurenin

Correspondent: László VÉCSEI MD, DSc, Department of Neurology, Faculty of Medicine, University of Szeged; H-6725 Szeged, Semmelweis u. 6. Phone: +3662/545351, e-mail: vecsei.laszlo@med.u-szeged.hu

Érkezett: 2013. november 20. Elfogadva: 2014. február 10.

www.elitmed.hu

The hormone *somatostatin (SST)* was isolated from the hypothalamus in the 1970's¹⁻³. It was originally regarded only as an inhibitor of growth hormone (GH) release (and was previously called somatotropin release inhibiting factor), but it is now known that SST has a number of biological effects. Burgus et al. observed that the addition of crude extracts of the ovine hypothalamus to anterior pituitary cells maintained in vitro inhibited the secretion of GH and they isolated a compound that accounted for all the GH-release inhibiting activity of the crude extract¹. After sequencing and synthesis the peptide displayed biological activity both in vitro and in vivo. SST, a small cyclic peptide, exists in two biologically active forms: SST-14 and SST-28, which are produced by the alternative posttranslational cleavage of the single prehormone. SST release can be stimulated by a variety of hormones (growth hormone- and corticotropin-releasing hormone, neurotensin), neuropeptides, neurotransmitters, cytokines, growth factors and nutrients in several tissues⁴. On the other hand, the neurotransmitter GABA and opiates generally inhibit SST secretion⁵. Inflammatory cytokines, such as interleukin-1, tumor necrosis factor alpha and interleukin-6, are potent stimulators of SST release⁶, while transforming growth factor beta and leptin⁷ inhibit the secretion of the peptide. The development of synthetic analogs has led to the effective treatment of clinical disorders including acromegaly, hormone-secreting tumors of the gastrointestinal tract and portal hypertensive bleeding⁴. As a general inhibitor of gastrointestinal endocrine secretion, SST inhibits both gastric acid and pepsin release⁸. Moreover, it combines these effects with a reduction in gastroduodenal mucosal blood flow, which appears to be important in the pathophysiology of peptic ulcer bleeding. These results were demonstrated by several experimental stress models; one of the earliest of these was the cysteamineevoked perforating duodenal ulcer model in rats developed in 1973 by Selye and Szabo⁹.

Cysteamine (CysA) is a small aminothiol generated by hydrolysis of the lipid-lowering drug pantethine (PAN), together with two pantothenic acids (vitamin B5). It is assumed that CysA is involved in the production of cholesterol and triglycerides by means of its binding to inactivate sulfur-containing amino acids in liver enzymes. Moreover, CysA is a naturally occurring duodenal ulcerogen and has the ability to cause adrenocortical necrosis too¹⁰. CysA and its derivatives deplete SST in the gastric mucosa, causing significant increases in gastric acidity and pepsin activity, alterations that contribute to the development of duodenal ulcer⁹. The

ulcerogenic activity of these derivatives is significantly correlated with their SST-depleting activity¹¹. In accordance with these findings, the administration of SST prevents the development of CysAinduced duodenal ulcer. However, the effects of CysA in causing duodenal ulcer and adrenal lesions suggest an interaction between the central/peripheral nervous system and the neuroendocrine system. In 1982, Palkovits et al. demonstrated that a single subcutaneous (sc.) injection of CysA (300 mg/kg) resulted in a quite selective SST depletion in the brain. An approximately 70-80% decrease in SST levels was observed in those areas where SST-producing neurons (periventricular nucleus) and SSTergic nerve terminals (median eminence) are located, whereas the CysA did not produce changes in the levels of other neuropeptides¹².

The naturally occurring antioxidant PAN is a stable disulfide precursor of pantetheine. The latter is an intermediate in the production of coenzyme A (CoA) in the organism (Figure 1.). From a biochemical aspect, the enzymatic cleavage of PAN produces CysA (and later taurine) and pantothenic acid. After absorption from the food, 4'-phosphopantetheine is reformed by the action of pantothenate kinase, after which ribose and adenine molecules attach to it in the mitochondrium to create CoA or bind to acyl carrier protein¹³. CoA and acyl carrier protein function as acyl or acetyl carriers. CoA facilitates the transfer of acetyl groups from pyruvate to oxaloacetate, thereby initiating the Szent-Györgyi-Krebs (tricarboxylic acid) cycle. CoA is involved in several ways in the fat metabolism, including the synthesis, transportation and degradation of fatty acids. Several clinical studies

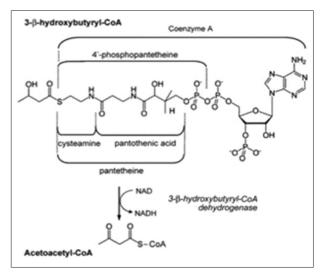


Figure 1. Metabolic pathway of pantethine (ImageCLEF 2011-Medical Datasheet)

have demonstrated its moderate benefits on dyslipidemic subjects¹⁴ and also its inhibition of platelet aggregation, which offers an effective therapeutic option for the treatment of patients with different forms of atherosclerotic vascular disease¹⁵.

From a neuroendocrinological aspect, the administration of PAN attenuates the levels of SSTand prolactin-immunoreactivity in the cerebral cortex and hypothalamus¹⁶ through the accumulation of CysA within cells throughout the body. It has therefore been suggested that this substance may be a useful pharmacological tool for elucidation of the role of SST in the central nervous system. The function of catecholaminergic pathways cannot be excluded completely since PAN or possibly the forming CysA inhibits dopamine beta-hydroxylase¹⁷. In 1989, Sellini et al. observed that a single high dose of PAN increased the levels of adrenocorticotropic hormone and cortisol (still within the normal range), but had no effect on the GH and Pro levels. This might be explained by a PAN-induced stimulus of the pituitary-adrenal axis or an increased synthesis of acetylcholine¹⁸.

There is evidence that CysA and PAN have behavior-modulating functions and other hormonal effects too. CysA can more effectively diminish locomotor, rearing and grooming activities than an equimolar dose of PAN. PAN influences several other behavioral responses in animals¹⁹: it stimulates the food intake in satiated rats, depending upon the stage of the circadian rhythm, but inhibits the food intake in fasted animals¹⁷. This effect is possibly mediated through the disinhibition of central appete-regulating SST-ergic pathways. It influences shuttle box learning²⁰ and causes locomotor inhibition (4 h after sc. treatment) and activation (24 h after repeated sc. injection) in open-field test^{21, 22}. It leads to the attenuation of SST-induced barrel rotation²³. From the aspect of passive avoidance behavior, there is no effect after sc. administration and merely a slight disruption after intracerebroventricular (icv.) treatment²¹. CysA, and to a lesser extent PAN, reduced the concentration of noradrenaline and increased those of dopamine and 3,4-dihydroxyphenylacetic acid in the hypothalamus. Pantothenic acid itself did not influence either the hypothalamic catecholamine concentrations or the behavior of rats²².

Kynurenines, as ligands of glutamate (Glu) receptors may also be important modulators of the neuroendocrine system. In 1976, Coyle and Schwarcz revealed that kainic acid (KA) is associated with lesions of the striatal neurons, as in Huntington's disease (HD) and it depletes SST²⁴. In 1989, Beal et al. demonstrated that the striatal exci-

totoxin lesions caused by the injection of quinolinic acid (QA), resulted in relative sparing of the SST and neuropeptide-Y (NPY) levels in rats²⁵. Cortical injections of certain agonists acting at the Glu receptors depleted the Glu and GABA levels, while the SST- and NPY immunoreactivity were either unchanged or significantly increased. Other Nmethyl-D-aspartate (NMDA) excitotoxins, such as KA and alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) caused significant decreases in the concentration of SST²⁶. Chronic QA-induced lesions resulted in similar alterations: elevated SST and NPY concentrations and reduced GABA, substance P-ir and choline acetyltransferase activity²⁷. The pattern of selective neuronal damage caused in the cerebral cortex by NMDA receptor agonists was similar to that observed in HD. However, there are contradictions, because there were no changes in SST, NPY or SP-ir with aging in the cerebral cortex or hippocampus following QA-induced striatal lesions²⁸, but there are different behavioral effects of KA- or QA-induced striatal lesions29.

Preclinical studies have indicated that the kynurenine pathway (KP) may be involved in the modulation of neuroendrocrine processes. Especially the function of kynurenic acid (KYNA) as a new, potential neuroendocrine molecule is emphasized. The main branch (approx. 95%) of the tryptophan (Trp) metabolism is the formation of kynurenines. Trp may be converted to L-kynurenine (KYN) by Trp- or indoleamine 2,3-dioxygenase (IDO) via a transition product. KYN serves as a key molecule between the neurotoxic and neuroprotective directions of the pathway. The neurotoxic QA is produced from KYN via additional toxic metabolites, which generate toxic free radicals, oxidative stress and lipid peroxidation, and hence excitotoxicity. In contrast, the characteristically neuroprotective KYNA is formed directly from KYN catalyzed by kynurenine aminotransferase (KAT) (**Figure 2.**)³⁰. Most of the neuroprotective, antiexcitotoxic effects of KYNA are explained by the inhibition of excitatory amino acid receptors. It has been proposed to act primarily as an antagonist at ionotropic AMPA and KA receptors, and as a noncompetitive antagonist at the strychnine-insensitive glycine-binding site of the NMDA receptors. KYNA can be an antagonist of the alpha7-nicotinic acetylcholine receptors, and a ligand for the orphan G protein-coupled receptors and the recently revealed aryl hydrocarbon receptors31. There is extensive literature on the role of the KP in different neurological diseases. Its protective impacts are emphasized in HD³², Parkinson's disease³³,

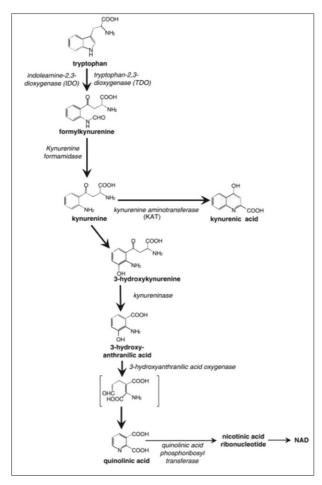


Figure 2. Main pathway of tryptophan-kynurenine metabolism

migraine^{34, 35}, multiple sclerosis^{36, 37}, schizophrenia³⁸, stroke^{39, 40} and epilepsy⁴¹, but its action in endocrine/neuroendocrine mechanisms has not yet been fully investigated.

In 1985, Rogers and Evangelista observed that leucine-stimulated insulin release from rat pancreatic islets can be inhibited by kynurenine metabolites (3-hydroxykynurenine and 3-hydroxyanthranilic acid)⁴². The regulatory effects of QA were investigated in ovariectomized, estradiol-primed rats. It was shown that icv. administered QA evoked an acute, dose-dependent increase of serum luteinizing hormone concentrations, which was blocked by KYNA. Brain morphologic disturbances were not detected in consequence of the treatments⁴³. The agonists and antagonists of Glu receptors regulate the hypothalamic-pituitary-adrenal axis (HPA) by different subtypes of amino acid receptors. Glu, KA and L-aspartate (Asp) significantly diminished the release of corticotropinreleasing hormone, while Asp and NMDA signifi-

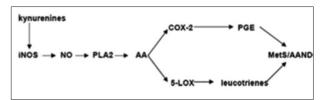


Figure 3. Relationship between kynurenines, metabolic syndromes (MetS) and age-associated neuroendocrine disorders (AAND) (Oxenkrug, G.F. Ann N Y Acad Sci, 2010)

cantly enhanced arginine-vasopressin release, whereas this was decreased by KA and quisqualic acid. KYNA completely abolished the effects of Asp in connection with both neurohumoral activators⁴⁴. A human study has indicated that cytokines such as interferon- and interleukin-10 regulate the expression of IDO in cells of hypothalamic and pituitary origin in connection with sickness behavior of patients⁴⁵. In 2010, Oxenkrug observed similar results: the pro-inflammatory cytokines facilitate the activity of IDO, which shifts the Trp metabolism to the formation of kynurenines. These molecules indirectly contribute to the development of metabolic syndromes and age-associated neuroendocrine disorders via apoptotic, neurotoxic and prooxidative effects (Figure 3.). A genetic predisposition to the presence of certain polymorphisms of pro-inflammatory cytokine genes might lead to the "superinduction" of IDO⁴⁶. The anxiolytic effects of KYNA were recently evaluated in neonatal chicks. A stress model was developed by social isolation, which was augmented by icv. corticotropin-releasing hormone. The stress responses were decreased by the icv. administration of Trp and (more effectively) KYNA. Attenuated distress vocalization, active wakefulness and increased time of sleeping position were observed after the KYNA treatment. Moreover, a depressed plasma corticosterone concentration was measured⁴⁷. When the toxic reverberations of the heavy oil spill near the northwest coast of Spain (2002) were examined seven years after the disaster from the aspects of endocrine and immunological alterations, the biomarker analyses revealed a significantly increased plasma level of cortisol, and decreased level of KYN and CD16+56+lymphocytes in exposed vs. unexposed individuals. More serious changes were observed in the chronically contaminated subjects, which suggested a chronic elevation of HPA activity and the possibility of endocrine diseases⁴⁸.

In summary, it may be stated that there have been highly important developments in neuroendocrinology since the early findings of Selye. Although there are as yet relatively few data about the potential role of kynurenines in neuroen-docrinology, the results already achieved are extremely noteworthy and immensely promising.

ACKNOWLEDGEMENT

This work was supported by the following grants: TÁMOP-4.2.2. A-11/1/KONV-2012-0052, Neuroscience Research Group of the Hungarian Academy of Sciences and University of Szeged.

REFERENCES

- 1. Burgus R, Ling N, Butcher M, Guillemin R. Primary structure of somatostatin, a hypothalamic peptide that inhibits the secretion of pituitary growth hormone. Proc Natl Acad Sci U S A 1973;70(3):684-8.
- 2. *Pelletier G, Labrie F, Arimura A, Schally AV*. Electron microscopic immunohistochemical localization of growth hormone-release inhibiting hormone (somatostatin) in the rat median eminence. Am J Anat 1974;140(3):445-50.
- 3. *Pelletier G, Leclerc R, Dube D, Labrie F, Puviani R, Arimura A, et al.* Localization of growth hormone-release-inhibiting hormone (somatostatin) in the rat brain. Am J Anat 1975;142(3):397-401.
- Herder WW, Lamberts SWJ. Somatostatin in clinical endocrinology. Müller Ee (ed.). Peptides and non peptides of oncologic and neuroendocrine relevance. Italia: Springer-Verlag; 2003; p. 73-82.
- Gamse R, Vaccaro DE, Gamse G, DiPace M, Fox TO, Leeman SE. Release of immunoreactive somatostatin from hypothalamic cells in culture:inhibition by gamma-aminobutyric acid. Proc Natl Acad Sci U S A 1980;77(9): 5552-6.
- Scarborough DE. Somatostatin regulation by cytokines. Metabolism 1990;39(9 Suppl 2):108-11.
- 7. Watanobe H, Habu S. Leptin regulates growth hormone-releasing factor, somatostatin, and alpha-melanocyte-stimulating hormone but not neuropeptide Y release in rat hypothalamus in vivo:relation with growth hormone secretion. J Neurosci 2002;22(14):6265-71.
- Bech K. Autonomic control of secretion of gastric acid and pepsin. J Auton Pharmacol 1989;9(6):419-28.
- Selye H, Szabo S. Experimental model for production of perforating duodenal ulcers by cysteamine in the rat. Nature 1973;244(5416):458-9.
- McComb DJ, Kovacs K, Horner HC, Gallagher GT, Schwedes U, Usadel KH, et al. Cysteamine-induced adrenocortical necrosis in rats. Exp Mol Pathol 1981;35(3):422-34.
- Szabo S, Reichlin S. Somatostatin in rat tissues is depleted by cysteamine administration. Endocrinology 1981;109 (6):2255-7.
- Palkovits M, Brownstein MJ, Eiden LE, Beinfeld MC, Russell J, Arimura A, et al. Selective depletion of somatostatin in rat brain by cysteamine. Brain Res 1982;240 (1):178-80.
- 13. *Ono S, Kameda K, Abiko Y*. Metabolism of panthethine in the rat. J Nutr Sci Vitaminol (Tokyo) 1974;20(3):203-13.
- McRae MP. Treatment of hyperlipoproteinemia with pantethine: A review and analysis of efficacy and tolerability. Nutrition Research 2005;25(4):319-33.
- 15. *Horvath Z, Vecsei L*. Current medical aspects of pantethine. Ideggyogy Sz 2009;62(7-8):220-9.
- 16. *Reichlin S, Bollinger-Gruber JA*. Pantethine, a cysteamine precursor, depletes immunoreactive somatostatin and prolactin in the rat. Endocrinology 1985;117(2):492-5.

- 17. Abucham J, Bollinger-Gruber J, Reichlin S. Pantethine, a somatostatin depleting agent, increases food intake in rats. Pharmacol Biochem Behav 1989;33(3):585-9.
- Sellini M, Sartori MP, Baccarini S, Bassi R. Various hormonal parameters (ACTH, cortisol, somatotropic hormone and prolactin) following administration of a single high dose of pantethine in healthy subjects. Boll Soc Ital Biol Sper 1987;63(2):143-5.
- 19. Vecsei L, Widerlov E, Ekman R, Alling C. Dose- and timeresponse effects of pantethine on open-field behavior, and on central neurotransmission in rats. Pharmacol Biochem Behav 1990;35(1):165-70.
- Vecsei L, Widerlov E, Ekman R, Alling C. Cysteamine and pantethine effects on passive avoidance behavior, shuttle box learning, open-field activity, striatal catecholamines and somatostatin. Arch Int Pharmacodyn Ther 1989;299: 14-27.
- Vecsei L, Alling C, Widerlov E. Comparative studies of intracerebroventricularly administered cysteamine and pantethine in different behavioral tests and on brain catecholamines in rats. Arch Int Pharmacodyn Ther 1990;305: 140.51
- 22. Vecsei L, Widerlov E, Alling C. Effects of pantethine, cysteamine and pantothenic acid on open-field behavior and brain catecholamines in rats. Arch Int Pharmacodyn Ther 1989;300:14-21.
- 23. Vecsei L, Alling C, Heilig M, Widerlov E. Effects of cysteamine and pantethine on open-field behavior, hypothalamic catecholamine concentrations, and somatostatin-induced barrel rotation in rats. Pharmacol Biochem Behav 1989;32(3):629-35.
- 24. Coyle JT, Schwarcz R. Lesion of striatal neurones with kainic acid provides a model for Huntington's chorea. Nature 1976;263(5574):244-6.
- 25. Beal MF, Kowall NW, Swartz KJ, Ferrante RJ, Martin JB. Differential sparing of somatostatin-neuropeptide Y and cholinergic neurons following striatal excitotoxin lesions. Synapse 1989;3(1):38-47.
- 26. *Beal MF, Swartz KJ, Finn SF, Mazurek MF, Kowall NW*. Neurochemical characterization of excitotoxin lesions in the cerebral cortex. J Neurosci 1991;11(1):147-58.
- 27. Beal MF, Ferrante RJ, Swartz KJ, Kowall NW. Chronic quinolinic acid lesions in rats closely resemble Huntington's disease. J Neurosci 1991;11(6):1649-59.
- 28. Finn SF, Hyman BT, Storey E, Miller JM, Beal MF. Effects of aging on quinolinic acid lesions in rat striatum. Brain Res 1991;562(2):276-80.
- Vecsei L, Beal MF. Comparative behavioral and neurochemical studies with striatal kainic acid- or quinolinic acid-lesioned rats. Pharmacol Biochem Behav 1991;39(2): 473-8.
- Vecsei L, Szalardy L, Fulop F, Toldi J. Kynurenines in the CNS:recent advances and new questions. Nat Rev Drug Discov 2013;12(1):64-82.

- 31. Stone TW, Stoy N, Darlington LG. An expanding range of targets for kynurenine metabolites of tryptophan. Trends Pharmacol Sci 2013;34(2):136-43.
- 32. Zadori D, Nyiri G, Szonyi A, Szatmari I, Fulop F, Toldi J, et al. Neuroprotective effects of a novel kynurenic acid analogue in a transgenic mouse model of Huntington's disease. J Neural Transm 2011;118(6):865-75.
- Zinger A, Barcia C, Herrero MT, Guillemin GJ. The involvement of neuroinflammation and kynurenine pathway in Parkinson's disease. Parkinsons Dis 2011;2011: 716859.
- 34. Knyihar-Csillik E, Toldi J, Mihaly A, Krisztin-Peva B, Chadaide Z, Nemeth H, et al. Kynurenine in combination with probenecid mitigates the stimulation-induced increase of c-fos immunoreactivity of the rat caudal trigeminal nucleus in an experimental migraine model. J Neural Transm 2007;114(4):417-21.
- 35. *Guo S, Vecsei L, Ashina M.* The L-kynurenine signalling pathway in trigeminal pain processing:a potential therapeutic target in migraine? Cephalalgia 2011;31(9):1029-38.
- 36. Guillemin GJ, Kerr SJ, Pemberton LA, Smith DG, Smythe GA, Armati PJ, et al. IFN-beta1b induces kynurenine pathway metabolism in human macrophages:potential implications for multiple sclerosis treatment. J Interferon Cytokine Res 2001;21(12):1097-101.
- Hartai Z, Klivenyi P, Janaky T, Penke B, Dux L, Vecsei L. Kynurenine metabolism in multiple sclerosis. Acta Neurol Scand 2005;112(2):93-6.
- Sathyasaikumar KV, Stachowski EK, Wonodi I, Roberts RC, Rassoulpour A, McMahon RP, et al. Impaired kynurenine pathway metabolism in the prefrontal cortex of individuals with schizophrenia. Schizophr Bull 2011;37(6): 1147-56.
- 39. Brouns R, Verkerk R, Aerts T, De Surgeloose D, Wauters A, Scharpe S, et al. The role of tryptophan catabolism along the kynurenine pathway in acute ischemic stroke. Neurochem Res 2010;35(9):1315-22.
- 40. Gigler G, Szenasi G, Simo A, Levay G, Harsing LG, Jr., Sas K, et al. Neuroprotective effect of L-kynurenine sulfate

- administered before focal cerebral ischemia in mice and global cerebral ischemia in gerbils. Eur J Pharmacol 2007;564(1-3):116-22.
- 41. Demeter I, Nagy K, Gellert L, Vecsei L, Fulop F, Toldi J. A novel kynurenic acid analog (SZR104) inhibits pentylenetetrazole-induced epileptiform seizures. An electrophysiological study :special issue related to kynurenine. J Neural Transm 2012;119(2):151-4.
- 42. Rogers KS, Evangelista SJ. 3-Hydroxykynurenine, 3-hydroxyanthranilic acid, and o-aminophenol inhibit leucine-stimulated insulin release from rat pancreatic islets. Proc Soc Exp Biol Med 1985;178(2):275-8.
- 43. Johnson MD, Whetsell WO, Jr., Crowley WR. Quinolinic acid stimulates luteinizing hormone secretion in female rats:evidence for involvement of N-methyl-D-aspartate-preferring receptors. Exp Brain Res 1985;59(1):57-61.
- 44. *Patchev VK, Karalis K, Chrousos GP*. Effects of excitatory amino acid transmitters on hypothalamic corticotropin-releasing hormone (CRH) and arginine-vasopressin (AVP) release in vitro:implications in pituitary-adrenal regulation. Brain Res 1994;633(1-2):312-6.
- 45. Tu H, Rady PL, Juelich T, Smith EM, Tyring SK, Hughes TK. Cytokine regulation of tryptophan metabolism in the hypothalamic-pituitary-adrenal (HPA) axis:implications for protective and toxic consequences in neuroendocrine regulation. Cell Mol Neurobiol 2005;25(3-4):673-80.
- 46. *Oxenkrug GF*. Metabolic syndrome, age-associated neuroendocrine disorders, and dysregulation of tryptophan-kynurenine metabolism. Ann N Y Acad Sci 2010;1199:1-14.
- 47. Yoshida J, Tomonaga S, Ogino Y, Nagasawa M, Kurata K, Furuse M. Intracerebroventricular injection of kynurenic acid attenuates corticotrophin-releasing hormone-augmented stress responses in neonatal chicks. Neuroscience 2012; 220:142-8.
- 48. Laffon B, Aguilera F, Rios-Vazquez J, Garcia-Leston J, Fuchs D, Valdiglesias V, et al. Endocrine and immunological parameters in individuals involved in Prestige spill cleanup tasks seven years after the exposure. Environ Int 2013;59:103-11.