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Abstract

Pituitary adenylate cyclase activating polypeptide (PACAP) is an endogenous neuropeptide with a widespread distribution both in the nervous system and peripheral organs. The peptide is also present in the female gonadal system, indicating its role in reproductive functions. While a lot of data are known on PACAP-induced effects in oogenesis and in the regulation of gonadotropin secretion at pituitary level, its placental effects are somewhat neglected in spite of the documented implantation deficit in mice lacking endogenous PACAP. The aim of the present review is to give a brief summary on the occurrence and actions of PACAP and its receptors in the placenta. Radioimmunoassay (RIA) measurements revealed increased serum PACAP levels during the third trimester and several changes in placental PACAP content in obstetrical pathological conditions, further supporting the function of PACAP during pregnancy.

Both the peptide and its receptors have been shown in different parts of the placenta and the umbilical cord. PACAP influences blood vessel and smooth muscle contractility of the uteroplacental unit and is involved in regulation of local hormone secretion. The effects of PACAP on trophoblast cells have been mainly studied in vitro. Effects of PACAP on cell survival, angiogenesis and invasion/proliferation have been described in different trophoblast cell lines. PACAP increases proliferation and decreases invasion in proliferative extravillous trophoblast cells, but not in primary trophoblast cells, where PACAP decreased the secretion of various angiogenic markers. PACAP pretreatment enhances survival of non-tumorous primary trophoblast cells exposed to oxidative stress, but it does not influence the cell death-inducing effects of methotrexate in proliferative extravillous cytotrophoblast cells. Interestingly, PACAP has pro-apoptotic effect in choriocarcinoma cells suggesting that the effect of PACAP depends on the type of trophoblast cells. These data strongly support that PACAP plays a role in normal and pathological pregnancies and our review provides an overview of currently available experimental data worth to be further investigated to elucidate the exact role of this peptide in the placenta.

Keywords (separated Pregnancy - Trophoblast - Proliferation - Migration - Placenta - Human by " - ")

Chapter 23 Occurrence and Functions of PACAP in the Placenta

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Both the peptide and its receptors have been shown in different parts of the 18 placenta and the umbilical cord. PACAP influences blood vessel and smooth muscle 19 contractility of the uteroplacental unit and is involved in regulation of local hor-20 mone secretion. The effects of PACAP on trophoblast cells have been mainly stud-21 ied in vitro. Effects of PACAP on cell survival, angiogenesis and invasion/ 22 proliferation have been described in different trophoblast cell lines. PACAP 23 increases proliferation and decreases invasion in proliferative extravillous 24 trophoblast cells, but not in primary trophoblast cells, where PACAP decreased the 25

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secretion of various angiogenic markers. PACAP pretreatment enhances survival of 26 non-tumorous primary trophoblast cells exposed to oxidative stress, but it does not 27 influence the cell death-inducing effects of methotrexate in proliferative extravil-28 lous cytotrophoblast cells. Interestingly, PACAP has pro-apoptotic effect in chorio-29 carcinoma cells suggesting that the effect of PACAP depends on the type of 30 trophoblast cells. These data strongly support that PACAP plays a role in normal 31 and pathological pregnancies and our review provides an overview of currently 32 available experimental data worth to be further investigated to elucidate the exact 33 role of this peptide in the placenta. 34

Keywords Pregnancy • Trophoblast • Proliferation • Migration • Placenta • Human 35

Introduction 36

Pituitary adenylate cyclase activating polypeptide (PACAP) was first described as a 37 hypothalamic neuropeptide acting on the pituitary [1]. Numerous subsequent stud-38 ies have described its regulatory effects in the hypothalamo-hypophyseal-endocrine 39 gland axis at all levels. Shortly after its discovery it became evident that PACAP 40 occurs at high levels in several peripheral organs, especially in the gonads. Arimura 41 and coworkers showed that after the hypothalamus, highest PACAP levels are found 42 in the testis [2]. This drew the attention to the peptide as a regulator of male fertility 43 and reproduction. Indeed, PACAP was found to influence spermatogenesis at vari-44 ous levels [3-6]. 45 PACAP is also involved in female reproductive functions. Although our knowl-46 edge on PACAP in reproductive functions is still limited, currently available data 47 clearly indicate that the neuropeptide plays an important regulatory role in female 48 reproductive physiology and pathology (rev. [7]). Briefly, PACAP, at the hypotha-49 lamic level, influences receptive behavior in female rodents, in association with 50 gonadotropin releasing hormone and steroids [8], and plays an important modula-51 tory role in pituitary hormone production. The role of PACAP in the hypothalamo-52

pituitary-gonadal axis has been reviewed several times previously [9-13] and is 53 reviewed in the present book in two chapters (11, and 12.). PACAP is present in the 54

ovary, in the ovarian follicular fluid and plays an important role in oocyte matura-55 tion [7, 14, 15]. In humans, the level of immunoreactive PACAP in the follicular 56

fluid of hyperstimulated women is correlated with the number of retrieved oocytes 57

[16]. PACAP also plays a role in the muscle contraction of the vaginal wall as well 58 as that of the uterus and uterine tube [17–19]. Decreased immunoreactivity was 59

shown in vaginal wall diseases and the plasticity of the PACAPergic system was 60

demonstrated after vaginal reconstructive surgery [20, 21]. The PACAPergic inner-61

vation of the female genital tract was also described and has been associated with 62

nerves originating from the paracervical ganglia [22, 23]. 63

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During pregnancy, the serum level of PACAP increases in the third trimester in 64 healthy pregnant women and it markedly decreases during delivery, reaching pre-65 birth levels 3 days after delivery [24]. Winters and colleagues indicated a possible 66 difference between cesarian and vaginal births regarding PACAP levels in the fetal 67 cord blood [25]. In pregnant rats, Papka et al. [26] detected PACAP immunoreactiv-68 ity in cervical nerves, lumbosacral dorsal root ganglia and in the spinal cord. 69 Immunoreactivity showed changes during pregnancy, indicating that sensory nerve-70 derived PACAP is involved in the innervation of the cervix and may play a role in 71 cervical ripening [26]. The role of PACAP in reproduction and offspring care does 72 not seem to finish with birth: PACAP plays a complex regulatory role in breastfeed-73 ing. The action of PACAP on prolactin synthesis and release is complex and influ-74 enced by several factors [27, 28]. The high concentrations of PACAP in the milk 75 suggest that the peptide plays a role in the development of the newborn [29-31]. 76 Equally possible function of PACAP in the milk is a local regulatory action of milk 77 production and/or mammary gland development [32, 33]. (This is reviewed else-78 where in this book by Tamas and co-workers 49.). In addition, PACAP was impli-79 cated to play a role in maternal behavior as showed by the decreased maternal 80 crouching behavior of PACAP knockout mice [34]. 81

It seems that placental functions of the neuropeptide are somewhat neglected in spite of the findings of Isaac and Sherwood [35], who described that the reproductive rate of PACAP knockout mice is lower due to implantation insufficiency, clearly indicating a placental role of endogenous PACAP. The present review briefly summarizes the occurrence and actions of PACAP and its receptors in the placenta. 86

Occurrence of PACAP and PACAP Receptors in the Placenta 87

Occurrence of PACAP and its receptors are summarized in a schematic drawing 88 (Fig. 23.1). The gene encoding VPAC receptors was found to be weakly expressed 89 in human placenta at relative prevalent levels comparable to that in the testis, kidney 90 and thymus [36]. Another study also confirmed these findings [37]. 91

Subsequent studies gave further insight into the occurrence and distribution of 92 PACAP and its receptors in the placenta. Radioimmunoassay and immunocytochem-93 istry first confirmed the expression of PACAP27 and PACAP38 in human placentas 94 [38]. PACAP levels in the placenta were compared to those in the isthmic region of 95 the uterus and in the umbilical cord. Both forms of PACAP could be detected in the 96 examined specimens of the uteroplacental unit. PACAP38 concentration was higher 97 than PACAP27 levels in all examined regions. Uterus and placenta showed similar 98 levels of immunoreactivity, while intensity in the umbilical cord was much weaker 99 [38]. These authors found no immunoreactive nerve fibers in the placenta or umbili-100 cal cord, immunoreactive nerve fibers were only present in the uterus, with isthmic 101 region and nonpregnant myometrium showing stronger immunoreactivity than preg-102 nant uterus. PACAP immunoreactivity by radioimmunoassay was confirmed later by 103 Brubel et al. [39]: both forms of the peptide could be detected with PACAP38 104

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Fig. 23.1 Distribution of PACAP and PACAP receptors in the human placenta. (A). Representative photomicrograph of the human placenta indicating the parts depicted in the (B) schematic illustration of the human placenta. *Star:* PACAP38 immunohistochemistry, *filled circle:* PACAP mRNA, *filled triangle:* PAC1 mRNA, *filled square:* PACAP38 radioimmunoassay, *open square:* PACAP27 radioimmunoassay. Exact localization of VPAC receptor is not known

showing stronger immunoreactivity, in agreement with the above earlier observa-105 tions. Furthermore, it was also found that different parts of the human placenta con-106 107 tained similar levels of PACAPs, such as central maternal, peripheral maternal, central fetal, and peripheral fetal parts. The umbilical cord showed very weak immunoreac-108 tivity. In addition, Brubel et al. [39] also compared the level of immunoreactivity in 109 the 1st trimester samples and full-term placentas. Markedly stronger immunoreactiv-110 ity for PACAP38 was found in full-term placentas on both the maternal and fetal 111 sides. In contrast, PACAP27-immunoreactivity only increased on the maternal side, 112 while it did not change on the fetal side towards the end of pregnancy. 113

Scaldaferri and collagues [40] studied PACAP and PAC1 receptor expression by 114 means of Northern blot analysis, polymerase chain reaction (PCR) and immunohis-115 tochemistry. The authors detected the presence of PACAP and PAC1 receptor in 116 both rat and full-term human placentas. In human placentas, strong immunohisto-117 chemical staining was observed in stromal cells surrounding blood vessels and 118 weaker signal was detected in vessel walls in stem villi. In terminal villi, stromal 119 cells expressed PACAP38 immunoreactivity. In stem villi, the stromal immunoreac-120 tivity was restricted to the periphery, while this spatial distribution pattern was lack-121 ing in terminal villi, where immunostaining was dispersed throughout the stroma. In 122 rat placentas, several immunostained cells were observed in the labyrinth and in the 123 villous-like structures of the intraplacental yolk sac, structures derived from yolk 124 sac extensions into placental disc during late pregnancy [40]. 125

Isoforms of the PAC1 receptor were also studied by RT-PCR. Different isoform 126 expression was revealed in rat and human placentas. Rat placenta was shown to 127 express 3 isoforms: the short, hip or hop variant and the hip-hop variant. In contrast, 128 human placenta only expressed the SV2 form, homologous to the rat hop form. 129 Radioligand receptor binding assay revealed that the relative potencies of PACAP-130 related peptides were PACAP27, PACAP38 with comparable strong binding (almost 131 equipotent, PACAP27 slightly stronger) and VIP with weaker binding (10 times less 132 potent). Growth hormone releasing hormone and unrelated peptides, such as beta-133 endorphin and corticotropin-releasing hormone, did not bind to the receptor [40]. 134

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PACAP and PAC1 receptor mRNAs were investigated in rat placenta also by 135 Koh and coworkers [41]. Rat placenta consists of decidua basalis, junctional and 136 labyrinth zones. Expression of PACAP and PAC1 receptor mRNA was detected by 137 in situ hybridization in decidual cells, and in chorionic vessels and stromal cells of 138 the labyrinth zone [41]. In decidual cells, signals were strongest on day 13.5 of 139 gestation, then decreased with more advanced stages. No signal could be detected 140 in the junctional zone. In contrast, signals were gradually increasing with advancing 141 pregnancy in the labyrinth zone, in the branching villi, stem villi and also in chori-142 onic vessels. 143

Koh et al. [42] investigated the expression of PACAP and PAC1 receptor mRNA 144 from human legal abortions of 6-7 weeks, from induced abortions of 14-24 weeks 145 (second trimester) and term placentas by cesarian section or normal vaginal deliv-146 ery. In situ hybridization revealed expression of PACAP and PAC1 receptor mRNAs 147 in stem villi and terminal villi. In 7- and 14-week-old samples, PACAP mRNA was 148 detected in stroma cells surrounding blood vessels within stem villi, with moderate 149 expression level, while stronger expression of PACAP mRNA was found at later 150 stages [42]. PACAP mRNA was only weakly expressed in cytotrophoblast and syn-151 cytiotrophoblast cells. PAC1 receptor expression was detected in the same areas: 152 stronger expression was described in stroma cells of the villi, while weaker expres-153 sion in the trophoblast cells. Similar pattern of VIP immunoreactivity was detected 154 by Marzioni and coworkers [43]. Immunostaining was present in both trophoblast 155 layers and in the endothelium of the fetal vessels. 156

The gradual increase of the mRNA expression for both PACAP and its specific 157 receptor implies a role of PACAP in placental growth. The findings with RIA also 158 confirmed these increasing levels in late placentas compared to early placentas [4]. 159

Expression of PACAP in Pathological Pregnancies

Butadiene diepoxide is a reactive metabolite of 1,3-butadiene that is an important 161 industrial chemical and causes a dose-dependent inhibition of deciduoma development in rats [44, 45]. Placental expression of PACAP mRNA significantly decreased 163 in rats pretreated with 1,3-butadiene, a chemical toxin for reproduction in rats [44, 164 46]. The decrease was more drastic on gestation day 12 (63%) than on day 9 (48%). 165

Our recent preliminary investigations have focused on the levels of PACAP38 166 and PACAP27 in cases of different pathological situations. These measurements 167 were done in order to show possible changes of PACAP expression caused by 168 maternal smoking during pregnancy or fetal distress or hypoxia leading to presence 169 of meconium in the amniotic fluid. Human placentas were collected from full-term 170 placentas. Samples were taken from the chorionic villi (fetal side), the decidua 171 (maternal side) and the umbilical cord. Four different groups were examined: (1) 172 normal pregnancy and birth; (2) amniotic fluid with meconium-premature birth 173 (36–38 weeks); (3) premature birth (31–32 weeks) with smoking during pregnancy; 174 (4) post term birth with smoking during pregnancy (n=3 in all groups). 175

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The procedure used was in accordance with protocols approved by the ethical committee (no. 2784,3117, University of Pecs; 8-28/92 009-10 I 8EKU, ETT TUKEB, Ministry of Health, Hungary). Tissue samples were weighed and homogenized in ice-cold distilled water. The homogenate was centrifuged (12,000 rpm, 4 °C, 30 min), and the supernatant was further processed for RIA analysis of PACAP38 and PACAP27 contents, as previously described [47, 48].

Briefly, the conditions were as follows: antisera: PACAP38 "88111-3" (working 182 dilution, 1:10,000) and PACAP27 "88123" (dilution: 1:45,000); tracer: mono-125I-183 labelled ovine PACAP24-38 and mono-125I-labelled ovine PACAP27 prepared in 184 our laboratory (5000 cpm/tube); standard: ovine PACAP38 and PACAP27 ranging 185 from 0 to 1000 fmol/ml; buffer: assay prepared in 1 ml of 0.05 mol/l (pH 7.4) phos-186 phate buffer containing 0.1 mol/l sodium chloride, 0.25% (wt/vol) bovine serum 187 albumin, and 0.05 % (wt/vol) sodium azide; incubation time: 48–72 h of incubation 188 at 4 °C; separation solution; charcoal/dextran/milk powder (10:1:0.2 g in 100 ml of 189 distilled water). Results are given as fmol/mg PACAP38-like immunoreactivity and 190 PACAP27-like immunoreactivity in the tissue samples. Differences between 191 PACAP contents were assessed by ANOVA test. 192

As sample sizes were low, no definite statistical comparison was possible, but 193 some conclusions on tendencies can be drawn based on these preliminary results. We 194 found detectable differences in levels of PACAP38 between pathological and physi-195 ological pregnancies suggesting that PACAP expression may be disturbed or upregu-196 lated during pathological events related to pregnancy (Fig. 23.2), PACAP27 levels 197 did not show pronounced alterations in any examined condition. In accordance with 198 previous results, levels of PACAP27 were significantly lower than PACAP38 levels 199 in each sample. Under hypoxic condition, as indicated by the presence of meconium 200 in the amniotic fluid, PACAP levels did not change in most samples. Only a slight 201 decrease was observed in the chorionic villi and in the umbilical cord. Samples from 202 premature births of smoking mothers showed marked increases in all regions exam-203 ined except for the central decidua, where a slight decrease was observed. Decreases 204 were also detected from samples derived from post term births of smoking mothers. 205 Although our results are preliminary from limited number of clinical cases, some 206 tendencies are promising as it seems that PACAP levels change in some pathological 207 conditions. PACAP alterations have been observed in several diseases in both tissue 208 samples and body fluids including plasma, cerebrospinal fluid and follicular fluid [16, 209 49]. Recent studies have shown that PACAP levels were lower in lung cancer, colon 210 and kidney tumor samples compared to healthy tissue, while higher in prostate cancer 211 samples compared with samples from benign prostatic hyperplasia [50, 51]. As 212 PACAP is indicated as a potential biomarker for various conditions by several stud-213 ies, it would be important to conduct a clinical study including enough pathological 214 placenta samples to draw final conclusions (clinical review: see Reglodi et al. in this 215 book, chapter 2.). 216

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Fig. 23.2 Levels of PACAP27 and PACAP38 in placentas from healthy and pathological pregnancies. Results are given as mean fmol/mg tissue PACAP±SEM. Differences between levels of PACAP38 and PACAP27-like immunoreactivities were significant in all cases

Effects of PACAP in Vessels of the Placenta

Effects of PACAP on different components and cells of the placenta are summarized 218 in Table 23.1. Steenstrup et al. [38] investigated the effects of PACAP on vessels and 219 smooth muscle contractility in the uteroplacental unit. They found that preincubation 220 of the vessels with PACAPs and VIP produced a significant and concentration-221 dependent inhibition of the norepinephrine-induced contraction on the intramyome-222 trial and stem villus arteries. The high concentration needed for significant relaxation 223 indicates that the local release of the peptides is necessary to achieve this effect 224 in vivo. These results show that PACAP causes relaxation of the placental vessels. In 225 contrast, no effect was observed on either the amplitude, tone or frequency of strips 226 of spontaneously contracted myometrial smooth muscle obtained from pregnant 227 women [38]. These observations indicate that PACAP may be involved in the regula-228 tion of the uteroplacental blood flow. The time-related localization of endometrial/ 229



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t1.2			PACAP	
t1.3			concentration	
t1.4	Cells/region of placenta	Effect	(M)	Reference
t1.5	Stem villous arteries	Relaxation	10-10-10-6	[38]
t1.6	Intramyometrial arteries	Relaxation	10-10-10-6	[38]
t1.7 t1.8	JEG3 choriocarcinoma cells	cAMP increase, IL-6 secretion, alpha-subunit gene transcription	10-7	[52]
t1.9 t1.10 t1.11	JEG3 choriocarcinoma cells	No influence on survival of cells exposed to oxidative stress	10 ⁻⁷	Horvath et al. unpublished observation
t1.12 t1.13 t1.14	JAR choriocarcinoma cells	Decreased survival in cells exposed to oxidative stress or hypoxia	10 ⁻⁷	[53]
t1.15 t1.16 t1.17	JAR choriocarcinoma cells	No influence on survival of cells exposed to LPS, ethanol, methotrexate	10 ⁻⁹ -10 ⁻⁷	[39, 53]
t1.18 t1.19 t1.20 t1.21 t1.22 t1.23	JAR choriocarcinoma cells	Phosphorylation of ERK1/2 and JNK↑; Akt, GSK-3β, and p38 MAPK↓, Bax expression↓; in cells exposed to oxidative stress, PACAP decreased phosphorylation of all these	10-7	[53]
t1.24 t1.25	JAR choriocarcinoma cells	Agonistic effects of PACAP6-38 on signaling	10-7	[62]
t1.26 t1.27 t1.28	HTR-8/SVneo nontumorous primary trophoblast cells	Pretreatment increased survival in oxidative stress, co-treatment no effect	10 ⁻⁸ -10 ⁻⁷	[61]
t1.29 t1.30	HTR-8/SVneo nontumorous primary trophoblast cells	No effect on invasion	10-7	[61]
t1.31 t1.32 t1.33 t1.34 t1.35 t1.36	HTR-8/SVneo nontumorous primary trophoblast cells	Reduced levels of angiogenic factors active A, ADAMTS-1, angiogenin, angiopoietin-1, endocrine gland-derived vascular endothelial growth factor, and endoglin	10-6	[61]
t1.37 t1.38 t1.39 t1.40	HIPEC65 proliferative extravillous cytotrophoblast cells	Induced proliferation, but no effect on methotrexate-induced cell death decreased invasion	10 ⁻⁷	[61]
t1.41 t1.42 t1.43 t1.44	Decidual and peripheral mononuclear cells from early pregnancies	No effect on secreted angiogenic molecules or inflammatory cytokine production	10 ⁻⁶	[61]

t1.1 Table 23.1 Summary of the effects of PACAP in the placenta

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uterine PACAP was described by Spencer et al. [45]. PACAP mRNA pattern showed230similarity with that of the progesterone receptor during decidualization, but it was231consistently lower during gestation. Furthermore, both uterine and placental mRNA232expression pattern of decidual prolactin-related protein corresponded to PACAP and233progesterone receptor mRNA levels. They suggest that PACAP could be important234in facilitating endometrial blood flow and increase availability of metabolic sub-235strates to the developing deciduoma or embryo [45].236

Effects on Hormonal Secretion of the Placenta

Using JEG3 choriocarcinoma cells PACAP was found to induce a 12-fold increase 238 in cAMP secretion [52]. This action of PACAP was rapid, cAMP increase started 239 already after 30 min. cAMP in the placenta is known to stimulate alpha-subunit 240 expression. Alpha-subunit of the hypophyseal hormones LH, FSH, and TSH is also 241 present in the placental hormone human chorionic gonadotropin. PACAP38 was 242 found to positively regulate alpha-gene transcription in JEG3 cells, with maximal 243 effect at 100 nM concentration. The time course of this effect showed that PACAP 244 effect started after 8 h. Similar effects were observed with the homolog peptide, 245 VIP, but the effects of VIP an alpha-gene transcription started only after 24 h [52]. 246 These findings show that PACAP may be involved in placental hormone secretion, 247 and the similar effects of PACAP and VIP suggest that these effects are mediated by 248 the shared VPAC receptors. 249

Effects of PACAP on Survival of Trophoblast Cells

Effects of PACAP on trophoblast cell survival were studied in JAR human choriocar-251 cinoma cells [53]. PACAP treatment alone did not influence the survival rate. Cells 252 exposed to oxidative stress induced by H2O2 showed decreased survival rate, which 253 was further decreased by PACAP. A similar effect was observed in cells undergoing 254 chemically induced hypoxia by CoCl₂. No effects on survival were observed in cells 255 exposed to lipopolysaccharide (LPS), methotrexate or ethanol [39, 53]. These find-256 ings were contradicting the general survival-promoting effect of PACAP observed in 257 many different cell lines and tissues both in vitro and in vivo [54]. Examining the 258 signaling pathway revealed that PACAP treatment alone slightly increased phosphor-259 ylation of ERK1/2 (extracellular signal-regulated kinase) and JNK (c-Jun N-terminal 260 kinase) but decreased that of Akt (protein kinase B), MAPK (mitogen-activated pro-261 tein kinase), GSK-3 β (glycogen synthase kinase 3 beta) and the expression of Bax. 262 Oxidative stress alone increased phosphorylation of JNK and slightly decreased that 263 of Akt, ERK, and GSK-3β, while no changes were observed in the expression of 264

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phospho-p38 and Bax. In cells exposed to oxidative stress, PACAP treatment 265 decreased phosphorylation of all of these examined signaling molecules compared to 266 H₂O₂-treated controls. These results indicate that PACAP sensitizes the cells to some 267 stressors, like oxidative stress and in vitro hypoxia, while it does not affect the delete-268 rious effects of other stressors [53]. The reason for this is not known at the moment, 269 and may or may not reflect the physiological role of the peptide, since these experi-270 ments were performed in choriocarcinoma cells, in which the receptor expression 271 and signaling induced by PACAP may be significantly altered. 272

Similar results were described in retinoblastoma cells, where PACAP treatment 273 induced cell death [55] in spite of the well-known protective effects of PACAP in the 274 retina [56, 57]. PACAP also inhibited the growth of the neuronal tumor medulloblastoma 275 in spite of the well-known neuroprotective effects of the peptide [58-60]. An equally 276 possible explanation is that the sensitizing effect of PACAP to stressors such as hypoxia 277 and oxidative stress may be involved in the adaptation promoting effect of PACAP in 278 pregnancy and under pathological conditions. The first explanation is supported by later 279 findings in non-tumorous primary trophoblast cells (HTR-8/Svneo cells), where PACAP 280 pretreatment led to a significant increase in survival measured by MTT test in cells 281 exposed to oxidative stress by H₂O₂, while co-treatment had no effect [61]. However, 282 PACAP treatment did not influence the cell death-inducing effect of methotrexate in 283 HIPEC65 proliferative extravillous cytotrophoblast cell line, but induced proliferation of 284 these cells when treated alone with PACAP [61]. These results show that the effects of 285 PACAP on proliferation and survival of trophoblast cells depend on the type of stressor, 286 the timing of treatment and the type of cell. 287

Another interesting finding regarding survival and signaling effects in this cell line was that the generally accepted PAC1/VPAC2 antagonist PACAP6-38 exerted agonistic effects in JAR cells [62]. This does not seem to be specific for this cell line, since agonistic effects were found for example on rat trachea neuropeptide release, retinoblastoma cell survival and cartilage and bone development [55, 62– 64]. The reason for this might be the expression of a yet unknown splice variant of the receptor or the tumorous nature of the JAR cells.

295 Effects of PACAP on Invasion of Trophoblast Cells

PACAP treatment did not influence the invasion of HTR-8/Svneo human first trimester extravillous primary trophoblast cells, while it decreased the invasion of

HIPEC invasive, proliferative extravillous cytotrophoblast cells [61].

299 Effects of PACAP on Trophoblast Angiogenesis

Effects of PACAP on angiogenic factors were investigated in HTR-8/Svneo cells using an angiogenesis array method. Levels of several angiogenic markers were markedly decreased in the cell culture supernatant after 24 h of PACAP treatment.

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Secreted levels of active A, ADAMTS-1, angiogenin, angiopoietin-1, endocrine 303 gland-derived vascular endothelial growth factor, and endoglin were reduced [61]. 304 In human peripheral blood and decidual mononuclear cells obtained from healthy 305 pregnant women undergoing elective termination of apparently normal pregnancies 306 no effect on levels of secreted angiogenic molecules was found. Similarly, PACAP 307 treatment had no effect on the inflammatory cytokine production of these cells [61]. 308

Observations in PACAP Knockout Mice, Effects on Implantation

It has been reported several times that the reproductive rate of PACAP and PAC1 311 312 receptor knockout mice is lower than that of wild types [34, 65]. The reason for this is not elucidated yet, several mechanisms may be responsible for this effect. For 313 314 example, PACAP is involved in spermatogenesis and sperm motility [4], in steroid 315 hormone synthesis, in ovarian folliculogenesis, and in reproductive behavior [34, 35]. The small litter can also be due to the premature intrauterine death and early 316 317 postnatal death due to defects in breathing and temperature regulation [66, 67]. In addition to this multifactorial mechanism, it seems that placental defects are also 318 319 partially responsible for the lower reproductive rate of PACAP knockout mice. Isaac 320 and Sherwood [35] observed that while the puberty onset, estrous cycle and seminal 321 plugs of PACAP knockout mice were normal, significantly fewer PACAP null 322 females gave birth following mating than wild types. The authors found no defect in ovulation, ovarian histology or fertilization of released eggs, only 13% had 323 324 implanted embryos 6.5 days after mating compared to 81 % in wild types. Levels of 325 prolactin and progesterone were significantly lower in PACAP knockout females. These observations suggest that impaired implantation is involved in the observed 326 327 decreased fertility, the details of which need further clarification [35].

In summary, we give a brief review of data supporting a role of PACAP in normal and pathological pregnancies. The currently available experimental data are worth to be further investigated to elucidate the exact role in the placenta and evaluate the potential biomarker value of PACAP in reproductive pathology. 328 329 330

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