Elsevier Editorial System(tm) for

Transplantation Proceedings

Manuscript Draft

Manuscript Number: TransProc2608R1

Title: Evaluation of Mannose-Binding Lectin is a Useful Approach to Predict the Risk of Infectious Complications Following Autologous Hematopoietic Stem Cell Transplantation

Article Type: Original Works or Clinical Submission

Keywords: mannose-binding lectin, autologous hematopoietic stem cell transplantation, infectious complication

Corresponding Author: Dr. Zita Brigitta Radnay, M.D.

Corresponding Author's Institution: Institute for Internal Medicine, University of Debrecen

First Author: Zita Brigitta Radnay, M.D.

Order of Authors: Zita Brigitta Radnay, M.D.; Miklós Udvardy, Prof., M.D., PhD; Mária Papp, M.D., PhD; Jolán Hársfalvi, PhD; László Rejtő, M.D., PhD; Ildikó Pál, M.D.; Árpád Illés, Prof., M.D., PhD; Attila Kiss, Prof., M.D., PhD

TITLE PAGE

Evaluation of Mannose-Binding Lectin is a Useful Approach to Predict the Risk of Infectious Complications Following Autologous Hematopoietic Stem Cell Transplantation

My manuscript is submitted as an original work.

Authors:

Zita Brigitta Radnay M.D.¹, Miklós Udvardy Prof. M.D.¹, Mária Papp M.D.², Jolán Hársfalvi PhD^{3, 4}, László Rejtő M.D.¹, Ildikó Pál M.D.¹, Árpád Illés Prof. M.D.¹, Attila Kiss Prof. M.D.¹

Affiliations:

¹Department of Hematology, Institute for Internal Medicine, Faculty of Medicine, University of Debrecen, Debrecen, Hungary

²Department of Gastroenterology, Institute for Internal Medicine, Faculty of Medicine,

University of Debrecen, Debrecen, Hungary

³Department of Biophysics and Radiation Biology, Semmelweis University, Budapest, Hungary ⁴Clinical Research Center, Faculty of Medicine, University of Debrecen, Hungary

Email addresses of authors:

radnayzita@gmail.com

udvardy.miklosdr@gmail.com

drpappm@yahoo.com harsfalvi.jolan@med.semmelweis-univ.hu lrejto@med.unideb.hu palildiko89@gmail.com illesarpaddr@gmail.com akiss@med.unideb.hu

Corresponding author:

Zita Brigitta Radnay MD.

Department of Hematology, Institute for Internal Medicine

Faculty of Medicine, University of Debrecen

Nagyerdei krt. 98.

H-4032 Debrecen, Hungary

Telephone number: +36-20-582-9147

Fax number: +36-52-255-984

Email address: radnayzita@gmail.com

Grant information: The authors declare no conflict of interest.

TÁMOP-4.2.2/B-10/1-2010-0024, PhD student fellowship, Hungary

Key words: mannose-binding lectin, autologous hematopoietic stem cell transplantation, infectious complication

Abbreviations: (in alphabetical order)

 Tables: 6
 Figures: 2 (color – Yes / No)

Abstract and keywords

Hematopoietic stem cell transplantation (HSCT) associated immuncompromised state carries high risk of infectious complications. Mannose-binding lectin (MBL) is an acute phase protein involved in innate immune response. Serum MBL level is genetically determined and quite stable. According to literature, significant association was shown between low MBL concentrations and serious infections.

The association between serum MBL level and frequency, severity of infections was studied in 186 patients following autologous HSCT.

Double-monoclonal antibody sandwich ELISA was used to determine MBL antigen level in sera. MBL levels were measured around 100 days following transplantation, in a period without active infection.

21 patients (11%) were MBL deficient. The median time of first infection and number of infections during the first posttransplant year were not significantly different between MBL deficients and non-MBL deficients. Occurrence and number of infections after HSCT correlated with MBL/CRP ratio. Number of severe infections was not higher among MBL deficients. Occurrence of infections after pre-engraftment period in first posttransplant year were significantly different in patient-groups separated by MBL cut-off level.

MBL/CRP ratio might be a useful marker of infectious complications. MBL measurement may be helpful in antibiotic treatment, in case of MBL deficiency earlier and more intensive treatment may be indicated.

mannose-binding lectin, autologous hematopoietic stem cell transplantation, infectious complication

Highlights

- Immuncompromised state carries high risk of infectious complications.
- Time of first infection and number of infections during the first posttransplant year were not significantly different between MBL deficients and non-MBL deficients.
- Occurrence and number of infections after HSCT correlated with MBL/CRP ratio.
- Occurrence of infections after pre-engraftment period in first posttransplant year were significantly different in groups separated by MBL cut-off level.
- MBL/CRP ratio might be a useful marker of infectious complications.

1 Introduction

2

3 The innate immune system means immediate defence against infections and activates an 4 adequate specific immune response [1]. When the adaptive immune response is immature or 5 compromised, the innate immune system constitutes the principle defense against infection 6 [2]. Mannose-binding lectin (MBL) is a C-type serum lectin that plays a central role in the 7 innate immune response. MBL is produced by liver and is an acute phase protein [3,4]. The 8 opsonic activity of MBL was first described in relation to immune deficiency in 1968 [5]. In 9 plasma, MBL is associated with MBL-associated serine proteases (MASPs). MASP-2 is the 10 enzyme of MBL/MASP complex needed for activation of complement factor C4 [6].

11 The subunit of MBL consists of an N-terminal cross-linking region, a collagen-like domain, 12 and a C-terminal carbohydrate-recognition domain (CRD) [7]. The oligomeric configuration 13 permits to have multiple CRDs [8]. MBL binds microbial surface carbohydrates and mediates 14 opsonophagocytosis directly and by activation of the lectin complement pathway [9,10]. 15 Staphylococcus aureus and β -hemolytic group A streptococci bind MBL, but only a part of 16 several species (E. coli, Klebsiella species, Haemophilus influenzae, etc.) showed significant 17 binding [11]. MBL binding is inhibited by encapsulated organisms [10]. MBL allows 18 opsonization of Aspergillus fumigatus, Candida albicans and Criptococcus neoformans, the 19 main microorganisms involved in invasive fungal infections (IFI) [11,12].

MBL is also involved in the recognition of self-targets, such as apoptotic and necrotic cells [13]. The endothelial cells exposed to oxidative stress bind MBL [14]. Neoplastic diseases are often associated with altered glycosylation patterns, so surfaces of malignant cells might be recognised by MBL as non-self [15].

The reason of low MBL level may be the actual MBL concentration or the level of functional activity. If the goal is to estimate the activity of MBL/MASP complex, so MBL pathway

activity, anti-C4 antibody is needed to determine the amount of C4b bound to the surface
[1,16]. The results of this assay correlate well with assay for MBL as antigen, except in case
of MASP-2 deficiency [17,18].

Serum MBL concentrations vary from 5 to 5000 ng/ml, because of genetic mutations within the gene and its promoters [19,20]. More than 10% of the general population may be classified as MBL deficient [1]. The majority of MBL-deficients are healthy without higher susceptibility for infections [21]. MBL deficiency may increase risk of infection when additional impairments of the immune system are present [22].

There is a strong correlation between MBL concentration and genotype [23,24]. Individuals with the same genotypes may differ by 10-fold in MBL levels [25]. The capacity to increase MBL concentration during febrile neutropenia is associated with MBL2 genotype [26]. There is a small increase during acute phase responses [4]. This increase is slow (1-2 weeks after the inducing event) and modest (up to three-fold increase) [1].

The variant monomers have less complement fixation capability and higher turnover [27]. The
impairment of polymerization causes low serum levels of high molecular weight MBL and
impaired MBL function [28].

42 Gram-positive cocci are responsible for the majority of post-bone-marrow transplant 43 bloodstream infections. The most common Gram-positive species are coagulase-negative 44 Staphylococcus, Streptococcus viridans, MRSA, enterococci and Staphylococcus epidermidis 45 [29,30]. Fluoroquinolones prophylaxis reduced the rate of Gram negative infections but it has a lower efficacy against Gram positive microorganisms [31]. The frequency of resistant Gram 46 47 negative bacteraemia increases [32]. This may be associated with wider use of intravascular 48 devices and fluoroquinolones prophylaxis [33]. Occurrence of PCP decreased due to the use 49 of trimethoprim-sulphamethoxazole prophylaxis [34].

50 Viral infections present more frequently between day 31 and 100 post-transplant, the most 51 important are CMV pneumonia and gastrointestinal involvement [35,36,37]. The most 52 common early viral infection, HSV causes gingivostomatitis [38].

The number of fungal infections increases post-HSCT and invasive infections can be a significant cause of morbidity and mortality. The two most common and clinically relevant pathogens are Candida and Aspergillus [39,40]. Fluconazole prophylaxis reduced the incidence of fungal infections [41,42]. IFI is one of the most life-threatening complications following treatment of hematologic malignancies, especially after allogeneic HSCT [43].

The consequence of impaired MBL function would be an enlarged susceptibility to infections 58 59 [24,44,45]. Low MBL concentration may be a risk factor for infection in patients receiving myelosuppressive chemotherapy [46,47,48]. Microbiologically proved systemic or 60 61 disseminated infections are more common among patients with malignancy who have MBL 62 deficiency and who received high-dose chemotherapy and autologous HSCT [49]. The 63 duration and deepness of neutropenia influences the frequency and severity of infection [50]. 64 MBL deficients experience longer episodes of febrile neutropenia [46]. Effector functions of 65 MBL are severely compromised during neutropenia, because neutrophils are required for enhanced phagocytosis after MBL-induced complement activation [51]. 66

The normal MBL haplotype is associated with increasing MBL concentrations, whereas most patients with exon 1 mutations are not able to synthesize functional MBL and don't have elevated serum MBL levels during acute phase response [26,46,52].

According to some studies, that measured the incidence of fever as an end point, did not demonstrate an association with MBL deficiency. Febrile episodes and their duration did not vary on the basis of MBL levels [53,54,55]. Kilpatrick et al [55] found no relationship between MBL levels and chemotherapy-related infection. Rocha et al [56] could not detect an association of mutations in MBL2 gene with the incidence of first infection.

MBL reactive carbohydrate epitopes occur on the surface of several cancer cell lines [15],
there might be a general over-representation of MBL deficiency in patients with malignant
hematological diseases [47].

Oral mucositis is a common toxic side effect among patients receiving high-dose chemotherapy with autologous HSCT. Mucositis complicates treatment outcome by increasing the risk of infection, necessitating enteric or parenteral nutrition and prolonging hospitalization [57].

82

83 Patients and methods

84

The association between serum MBL level and frequency, severity and occurrence of infections has been studied in 186 patients following autologous HSCT. CRP was measured several times according to clinical decision, and the maximal CRP level during the first 14 days after HSCT was taken in account. Correlation between infections and MBL/CRP ratio were determined.

90 Subgroups, i.e. multiple myeloma (MM), non-Hodgkin (NHL) and Hodgkin lymphoma (HL) 91 were formed and infectious complications have been compared. Among the examined 92 patients, number of persons with NHL was 63 (female/male: 25/38, age: 52±11), 27 patients' 93 diagnosis was HL (female/male: 12/15, age: 34±9), and 94 patients had MM (female/male: 94 55/39, age: 56±8). Two patients with other diagnosis were also involved in the trial. The 95 control group consisted of 296 age- and gender-matched healthy individuals (female/male: 96 156/140, age: 50±16 yrs) selected from consecutive blood donors. Control ones did not have 97 any hematological or liver diseases. The control healthy group was the same as previously 98 published in a large study from our Institute [58]. MBL serum levels and occurrence of MBL 99 deficiency in case of healthy ones and patients with hematological diseases were compared.

Reaching the absolute neutrophil count (ANC) more than 1 G/L was taken in account as neutrophil engraftment and platelet count more than 20 G/L as platelet cell-line engraftment. We examined the distribution of microbiological results according to MBL level. It may be hypothesized that the progression, relapse following transplantation is related to MBL level and susceptibility to infections, among other parameters.

105 The range of MBL level in healthy population varies between 5 and 5000 ng/ml, <100 ng/ml 106 is defined as MBL deficiency. MBL antigen levels were measured around 100 days after 107 transplantation, in a period without active infection. MBL level is genetically determined and 108 quite stable. There is a small increase during acute phase responses [4]. In a few cases MBL 109 concentration were also measured before and around 100 days after HSCT and were almost 110 equal. Informed consent was signed by the examined patients. After blood samples were 111 taken, native tubes were centrifuged for 15 minutes at 3000 RPM, then sera samples were 112 stored at -70 °C in small aliquots until measuring.

We used a double monoclonal antibody sandwich ELISA system adopted from Minchinton et
al to determine MBL levels [23,58]. MBL assay was performed at the Clinical Research
Centre of Debrecen University, without prior knowledge of the patients' clinical information.

116 Continuous variables were summarized as means and standard deviation or as medians and 117 interquartile range and were compared with Mann-Whitney U-test or Student T-test. 118 Kolmogorov-Smirnov and Chi-square tests were used to find out the distribution of variations. 119 Kruskal-Wallis ANOVA by Ranks was used to compare data from more than two groups.

121 analysis was performed to determine the cut-off level of MBL. P<0,05 was considered to be

Correlation of variables were analysed with Spearman Rank order correlation test. ROC curve

122 significant. Graphpad Prism 5 and MedCalc were used for statistical analysis.

123

120

124 Results

125

126 Among the examined 186 patients with malignant hematological diseases, 21 patients were 127 proved to be MBL deficient. 51 infectious episodes (elevated CRP level, fever, other clinical 128 symptoms of infection) were found among MBL deficients, and 372 events were in MBL 129 competent group during the first 360 days after HSCT. The median time of onset of first 130 infection post-HSCT was day +7 [3;8] in MBL deficient and day +6 [4;8] among non-MBL 131 deficient patients (Table 1). The distribution of MBL level and also MBL/CRP ratio were log-132 normal among the patients, while distribution of CRP was normal with Kolmogorov-Smirnov 133 and Chi-square tests (Figure 1). With Spearman Rank order correlation test, there were strong 134 correlation between logarithmically transformed (log) MBL/CRP ratio and the time of onset 135 of first infection (p=0,04, and after take in account the occurrence of infection as a censoring 136 variation, p=0.0001) (Figure 2), and between log CRP and the time of first infection following 137 transplantation (p<0,05). The time of first infection correlated neither with MBL level nor 138 with log MBL (p=0,35). Correlation between log MBL and log CRP was almost significant 139 (p=0,052), correlation between log MBL and log MBL/CRP ratio was significant (p=0,001) 140 certainly.

141 Occurrence of infections were similar among MBL deficient and MBL competent ones (2,429 142 [1,478;3,379] vs 2,248 [1,993;2,516] infectious episodes/patient). Number of infections after 143 HSCT correlated with CRP and MBL/CRP ratio but not with MBL level (Spearman Rank 144 order correlation test, r=0,37, -0,17 and 0,07; p=0,02 and 0,34, respectively). Mann-Whitney 145 U-test showed not significant relationship in case of MBL level and occurrence of first 146 infection following transplantation (p=0,37), and MBL level and first infection in 14 days and 147 100 days after HSCT. Connections of occurrence of infection in 14 and 100 days and before 148 reaching ANC more than 1,5 G/L and log MBL were not significant with unpaired T-tests. But relation of occurrence of first infection in 14 and 100 days and before neutrophilengraftment with log CRP and log MBL/CRP ratio were significant.

151 Cut-off level of MBL according to occurrence of severe infections in posttransplant period, 152 determined by ROC curve analysis was 823 ng/ml. Variables of the two patient-groups 153 separated by MBL cut-off level were compared with Spearman Rank order correlation test. 154 Number of infectious episodes (p=0.0611) and time of onset of first infection after HSCT 155 (p=0.0905) were almost significantly different. Occurrence of infections after HSCT 156 (p=0.0480) and occurrence of infections after the pre-engraftment period in first 157 posttransplant year (during the period from day +14 until day 360) (p=0.0389) were 158 significantly different in patient-groups separated by MBL cut-off level.

159 Interestingly, MBL serum level was found to be significantly higher in the examined patients 160 with hematological diseases compared to healthy control population (MBL median, 1479 [380,8;2849] vs 1067 [253,5;2121], unpaired t-test, p= 0,005, significantly different). The 161 162 occurrence of absolute MBL deficiency was not significantly different between hematology 163 patients and healthy controls (11.4% vs 13.9%). The proportion of MBL deficients was the 164 highest among HL patients (Table 2). MBL concentration of the control population and the 165 examined patients according to diagnosis (NHL, HL, MM) were compared. Median MBL 166 level was the highest among patients with NHL. The onset of first infection was the earliest 167 among patients with HL (Table 3). The distribution of infectious episodes according to 168 diagnosis is showed in Table 4.

169 The most common infections after transplantation are respiratory tract infections and170 infections with high CRP, fever and severe mucositis.

171 Time of neutrophil engraftment is related to MBL level significantly in MM group (Spearman

172 Rank order correlation, p=0,024). Strong association was shown between platelet engraftment

time and MBL/CRP ratio among HL patients (p=0,003). Stem cell count and time to
engraftment correlated well (p<0,001).

175 Distribution of Gram positive and negative bacteria species in culture from the patients' 176 central venous catheter and blood is shown in Table 5 and 6. Positive results of central venous 177 catheter culture (n=25) depend on log MBL and MBL/CRP ratio, but the relationship was not 178 significant (t-test, p=0,23 and 0,15).

We examined whether the progression, relapse following transplantation is related to the patients' MBL levels or not. Association between occurrence of relapse and log MBL or log MBL/CRP were not significant (t-test, p=0,9 and 0,76). Among the examined patients, 23 patients have relapsed during the first year following HSCT and other 45 patients later. Time to relapse was not related to MBL and MBL/CRP ratio.

184

185 Discussion

186

Initiation of complement system may occur via classical, alternative and lectin pathway [59]. 187 188 MBL recognizes carbohydrate patterns [60]. Bacterial infections and autoimmune diseases are 189 frequently associated with complement deficiencies [61]. MBL is a C-type serum lectin [62], 190 the carbohydrate-binding sites allow interaction with the saccharide repeats on microbial 191 surfaces but rarely associated with mammalian high-mannose structures [7]. MBL deficiency 192 is a result of impaired assembly or stability of multimers [63]. MBL functions as a TLR co-193 receptor that enables the molecule to coordinate and synchronize the innate immune system 194 [64].

The serum levels of functional MBL correlate with MBL2 coding genotypes [58]. MBL concentration is explained by polymorphisms in the promoter region and in exon 1 of the gene [65,66].

According to literature, MBL deficiency is associated with increased susceptibility to infectious diseases, mainly when adaptive immunity is compromised (in early childhood [45,48], or following chemotherapy [46,47,67]). A significant association was shown between low MBL concentrations and serious infections related to chemotherapy [47]. MBL deficients have a greater number of severe infections and experience their first severe infection earlier, compared to non-deficients [54]. The association between low MBL and infections was independent of whether patients received prophylactic antibiotics or GM-CSF or not [68].

205 The range of MBL level is between 5 and 5000 ng/ml, <100 ng/ml is defined as MBL 206 deficiency. Serum MBL concentration is quite stable, shows small increase during acute 207 phase responses [4]. Among the examined 186 patients 21 ones were MBL deficient. The 208 time of onset of first infection post-HSCT was similar among MBL deficients and non-209 deficients. There were strong correlation between log MBL/CRP ratio and time of first infection following HSCT, but the onset of first infection was not correlated significantly with 210 211 log MBL. Occurrence of infections were similar among MBL deficient and MBL competent 212 ones. The number of infections after HSCT correlated with MBL/CRP ratio but not with MBL 213 level. Connections of occurrence of first infection in 14 and 100 days and before neutrophil 214 engraftment and log MBL were not significant, but with log CRP and log MBL/CRP ratio 215 were significant. We could not find strong association between MBL level and incidence, 216 frequency and time of infections. An explanation can be that effector functions of MBL are 217 severely compromised during neutropenia, because neutrophils are required for enhanced 218 phagocytosis after MBL-induced complement activation [51]. Cut-off level of MBL 219 according to occurrence of severe infections in posttransplant period, determined by ROC 220 curve analysis was 823 ng/ml. Number of infections and time of first infection after HSCT 221 were almost significantly different in groups separated by MBL cut-off level. Occurrence of infections following HSCT and after the pre-engraftment period in first posttransplant yearwere significantly different in patient-groups separated by MBL cut-off level.

MBL serum level was significantly higher in the examined patients compared to healthy control population. The proportion of MBL deficients was the highest and onset of first infection was the earliest among HL patients.

Hematopoietic recovery and engraftment is related to patient-, disease-, and treatment-related variables [69]. Pre-engraftment phase is characterized by neutropenia, breaks in mucocutaneous barrier and vascular accesses required for patient care, and post-engraftment phase with impaired cell-mediated immunity [70].

Stem cell count and time to engraftment correlated well in the patient-group. Time to neutrophil engraftment is related to MBL level significantly in MM group. Strong association was shown between platelet engraftment time and MBL/CRP ratio in HL patients.

234 Infections might lead to delay or reduction in chemotherapy and might compromise the 235 effectiveness of therapy [47]. Infections occur frequently and can be serious following high-236 dose chemotherapy and HSCT. Infections might also compromise the engraftment of stem 237 cells. MBL measurement may be helpful in antibiotic treatment, in case of MBL deficiency 238 earlier and more intensive treatment may be indicated. The most common infections after 239 transplantation are respiratory tract infections and infections with high CRP, fever and severe 240 mucositis. The most of sepsis episodes are associated with infection of the CVC-entry-site 241 [71]. Mostly Gram positive bacteria species were isolated in culture from the examined 242 patients' central venous catheter and blood. Positive results of central venous catheter culture 243 depend on log MBL and MBL/CRP ratio, but not significantly. Infections are cured with 244 appropriate antimicrobial therapy and in some cases with central venous catheter removal 245 [33]. Among the examined patients, relapse and log MBL or log MBL/CRP were not 246 associated significantly.

Extrahepatic transcription of MBL2 gene has been reported in small intestine [72,73]. Transcription of MBL2 is upregulated in inflamed intestinal tissue samples. MBL2 gene is expressed in immune cells infiltrating the inflamed gut [74]. MBL-deficients would be less able to prevent passage of bacteria from the gut to the circulation as compared to MBL competents58 [58]. Oral mucositis grade did not differ significantly between MBL deficient and MBL competent patients in our trial.

MBL2 genotypes were not determined, as individuals with the same genotypes may differ by 10-fold in MBL levels [25]. Measurement of MBL serum levels by ELISA allows reliable quantification of the functional activity of MBL pathway in vivo [75]. Procalcitonin levels were not determined, CRP level is used regularly to monitoring infectious complications in our institution.

258 The relationship between increased susceptibility to infections and low MBL levels seen in 259 some studies, seems less pronounced in patients with suppression of phagocytic activity due 260 to intensive chemotherapy [1]. We could not find strong association between MBL level and 261 incidence, frequency and time of infections. Log MBL/CRP ratio correlated well with time of 262 first infection following HSCT. Lower MBL concentration may predispose to severe 263 infections in immunocompromised state. Occurrence of infections after the pre-engraftment 264 period in first posttransplant year were significantly different in patient-groups separated by 265 MBL cut-off level.

266

267 Acknowledgements

268

I would like to thank for the supportation and help of my supervisor, Attila Kiss Prof. MD. I performed the clinical examination, data analysis of patients information at Department of Hematology, Institute for Medicine, Clinical Center, University of Debrecen and Stem Cell

272 Transplantation Unit, University of Debrecen. I would like to thank for supportation and 273 advices of Miklós Udvardy Prof. MD, head of the Stem Cell Transplantation Unit, and Árpád 274 Illés Prof. MD, head of the Department of Hematology, Institute for Medicine. MBL assays 275 were performed at the Clinical Research Centre of Debrecen University, according to ELISA 276 methods adopted from Minchinton et al and locally settings performed by Maria Papp MD, 277 Jolán Hársfalvi PhD and their workgroup previously. Zsolt Karányi helped in statistical 278 analysis. I was a PhD student for three years, and at my first year I got supportation by 279 fellowship TÁMOP-4.2.2/B-10/1-2010-0024, the next two years were state-aided. Initial 280 results of this work were presented on a poster in 2011 at EBMT Congress, Paris, France 281 (Radnay Z, Kiss A, Papp M, Rejtő L, Hársfalvi J, Udvardy M. Mannose-binding lectin ELISA 282 is a new approach to predict the chance of infectious complications during autologous 283 haematopoietic stem cell transplantation. Bone Marrow Transplant 46 (Suppl. 1), S213-S214, 284 2011.).

285

287

288 The authors declare no conflict of interest.

289

- 291
- 292 [1] Thiel S, Frederiksen PD, Jensenius JC. Clinical manifestations of mannan-binding lectin

293 deficiency. Molecular Immunology, 2006; 43: 86-96.

- 294 [2] Eisen DP, Minchinton RM. Impact of Mannose-Binding Lectin on Susceptibility to
- 295 Infectious Diseases. Clinical Infectious Diseases, 2003; 37: 1496-1505.

²⁸⁶ Conflict of interest

²⁹⁰ References

- [3] Super M, Thiel S, Lu J, Levinsky RJ, Turner MW. Association of low levels of mannan-
- binding protein with a common defect of opsonisation. Lancet, 1989; 2: 1236-9.
- [4] Thiel S, Holmskov U, Hviid L, Laursen SB, Jensenius JC. The concentration of the C-type
- 299 lectin, mannan-binding protein, in human plasma increases during an acute phase response.
- 300 Clin Exp Immunol, 1992; 90: 31-35.
- 301 [5] Miller ME, Seals J, Kaye R, Levitsky LC. A familial plasma-associated defect of 302 phagocytosis. Lancet, 1968; 2: 60-63.
- 303 [6] Thiel S, Vorup-Jensen T, Stover CM, Schwaeble W, Laursen SB, Poulsen K et al. A
 304 second serine protease associated with mannan-binding lectin that activates complement.
- 305 Nature, 1997; 386: 506-510.
- 306 [7] Turner MW. Mannose-binding lectin: the pluripotent molecule of the innate immune
 307 system. Immunol Today, 1996; 17: 532-40.
- 308 [8] Bouwman LH, Roep BO, Roos A. Mannose-Binding Lectin: Clinical Implications for
 309 Infection, Transplantation, and Autoimmunity. Human Immunology, 2006; 67: 247-256.
- 310 [9] Neth O, Jack DL, Johnson M, Klein NJ, Turner MW. Enhancement of complement
- 311 activation and opsonophagocytosis by complexes of mannose-binding lectin with mannose-
- 312 binding lectin-associated serine protease after binding to Staphylococcus aureus. J Immunol,
- 313 2002; 169: 4430-4436.
- 314 [10] van Emmerik LC, Kuijper EJ, Fijen CA, Dankert J, Thiel S. Binding of mannan-binding
- 315 protein to various bacterial pathogens of meningitis. Clin Exp Immunol, 1994; 97: 411-416.
- 316 [11] Neth O, Jack DL, Dodds AW, Holzel H, Klein NJ, Turner MW. Mannose-binding lectin
- 317 binds to a range of clinically relevant microorganisms and promotes complement deposition.
- 318 Infect Immun, 2000; 68: 688-693.
- 319 [12] Jack DL, Klein NJ, Turner MW. Mannose-binding lectin targeting the microbial world
- for complement attack and opsonophagocytosis. Immunol Rev, 2001; 180: 86-99.

- [13] Nauta AJ, Raaschou-Jensen N, Roos A, Daha MR, Madsen HO, Borrias-Essers MC et al.
 Mannose-binding lectin engagement with late apoptotic and necrotic cells. Eur J Immunol,
 2003; 33: 2853.
- [14] Collard CD, Vakeva A, Morrissey MA, Agah A, Rollins SA, Reenstra WR et al.
 Complement activation after oxidative stress. Role of the lectin complement pathway. Am J
 Pathol, 2000; 156: 1549-1556.
- [15] Ma Y, Uemura K, Oka S, Kozutsumi Y, Kawasaki N, Kawasaki T. Antitumor activity of
 mannan-binding protein in vivo as revealed by a virus expression system: mannan-binding
 protein-dependent cell-mediated cytotoxicity. Proc Natl Acad Sci USA, 1999; 96: 371-375.
- [16] Super M, Levinsky RJ, Turner MW. The level of mannan-binding protein regulates the
 binding of complement-derived opsonins to mannan and zymosan at low serum
 concentrations. Clin. Exp. Immunol, 1990; 79: 144-150.
- [17] Thiel S, Moller-Kristensen M, Jensen L, Jensenius JC. Assays for the functional activity
 of the mannan-binding lectin pathway of complement activation. Immunobiology, 2002; 205:
 446-454.
- 336 [18] Stengaard-Pedersen K, Thiel S, Gadjeva M, Moller-Kristensen M, Sorensen R, Jensen
- 337 LT et al. Inherited deficiency of mannan-binding lectin-associated serine protease 2. N. Engl.
- 338 J. Med, 2003; 349: 554-560.
- 339 [19] Madsen HO, Garred P, Thiel S, Kurtzhals JA, Lamm LU, Ryder LP et al. Interplay
- 340 between promoter and structural gene variants control basal serum levels of mannan-binding
- 341 protein. J Immunol, 1995; 155: 3013-20.
- 342 [20] Madsen HO, Garred P, Kurtzhals JA, Lamm LU, Ryder LP, Thiel S et al. A new frequent
- allele is the missing link in the structural polymorphism of the human mannan-bindingprotein. Immunogenetics, 1994; 40: 37-44.

[21] Tacx AN, Groeneveld ABJ, Hart MH, Aarden LA, Hack CE. Mannan binding lectin in
febrile adults, no correlation with microbial infection and complement activation. J. Clin.
Pathol, 2003; 56: 956-959.

[22] Dahl M, Tybjaerg-Hansen A, Schnohr P, Nordestgaard BG. A population-based study of
morbidity and mortality in mannose-binding lectin deficiency. J Exp Med, 2004; 199: 13911399.

[23] Minchinton RM, Dean MM, Clark TR, Heatley S, Mullighan CG. Analysis of the
relationship between mannose-binding lectin (MBL) genotype, MBL levels and function in an
Australian blood donor population. Scand J Immunol, 2002; 56: 630-41.

354 [24] Garred P, Madsen HO, Hofmann B, Svejgaard A. Increased frequency of homozygosity
355 of abnormal mannan-binding-protein alleles in patients with suspected immunodeficiency.
356 Lancet, 1995; 346: 941-3.

357 [25] Steffensen R, Thiel S, Varming K, Jersild C, Jensenius JC. Detection of structural gene
358 mutations and promoter polymorphisms in the mannan-binding lectin (MBL) gene by
359 polymerase chain reaction with sequence-specific primers. J Immunol, Methods, 2000; 349:
360 554-560.

[26] Frakking FNJ, van de Wetering MD, Brouwer N, Dolman KM, Geissler J, Lemkes B et
al. The role of mannan-binding lectin (MBL) in pediatric oncology patients with febrile
neutropenia. European Journal of Cancer, 2006; 42: 909-916.

[27] Petersen SV, Thiel S, Jensenius JC. The mannan-binding lectin pathway of complement
activation: biology and disease association. Mol Immunol, 2001; 38: 133-49.

366 [28] Roos A, Garred P, Wildenberg ME, Lynch NJ, Munoz JR, Zuiverloon TC et al.

367 Antibody-mediated activation of the classical pathway of complement may compensate for

368 mannose-binding lectin deficiency. Eur J Immunol, 2004; 34: 2589.

- [29] Pawson H, Jayaweera A, Wigmore T. Intensive care management of patients following
 haematopoietic stem cell transplantation. Current Anaest & Critical Care, 2008; 19: 80-90.
- [30] Poutsiaka DD, Price LL, Ucuzian A, Chan GW, Miller KB, Snydman DR. Blood stream
- infection after hematopoietic stem cell transplantation is associated with increased mortality.
- 373 Bone Marrow Transplant, 2007; 40: 63-70.
- [31] Cruciani M, Rampazzo R, Malena M, Lazzarini L, Todeschini G, Messori A et al.
 Prophylaxis with fluoroquinolones for bacterial infections in neutropenic patients: a metaanalysis. Clin Infect Dis, 1996; 23(4): 795-805.
- [32] Cherif H, Kronvall G, Björkholm M, Kalin M. Bacteraemia in hospitalised patients with
 malignant blood disorders: a retrospective study of causative agents and their resistance
 profiles during a 14-year period without antibacterial prophylaxis. The Hematology Journal,
 2003; 4: 420-426.
- [33] Bonadio M, Morelli G, Mori S, Riccioni R, Papineschi F, Petrini M. Fluoroquinolone
 resistance in hematopoietic stem cell transplant recipients with infectious complications.
 Biomedicine & Pharmacotherapy, 2005; 59: 511-516.
- [34] Leung AN, Gosselin MV, Napper CH, Braun SG, Hu WW, Wong RM et al. Pulmonary
 infections after bone marrow transplantation: clinical and radiographic findings. Radiology,
 1999; 210: 699-710.
- [35] Wah TM, Moss HA, Robertson RJH, Barnard DL. Pulmonary complications following
 bone marrow transplantation. Br J Radiol, 2003; 76: 373-379.
- [36] Enright H, Haake R, Weisdorf D, Ramsay N, McGlave P, Kersey J et al.
 Cytomegalovirus pneumonia after bone marrow transplantation. Risk factors and response to
 therapy. Transplantation, 1993; 55(6): 1339-45.

- 392 [37] Castagnola E, Cappelli B, Erba D, Rabagliati A, Lanino E, Dini G. Cytomegalovirus
 393 infection after bone marrow transplantation in children. Human Immunology, 2004; 65: 416394 422.
- 395 [38] Soubani AO, Miller KB, Hassoun PM. Pulmonary complications of bone marrow
 396 transplantation. Chest, 1996; 109: 1066-1077.
- 397 [39] Raman T, Marik PE. Fungal infections in bone marrow transplant recipients. Expert
 398 Opinion Pharmacother, 2006; 7(3): 307-15.
- [40] De La Rosa GR, Champlin RE, Kontoyiannis DP. Risk factors for the development of
 invasive fungal infections in allogeneic blood and marrow transplant recipients. Transplant
 Infect Dis, 2002; 4(1): 3-9.
- 402 [41] Goodman JL, Winston DJ, Greenfield RA, Chandrasekar PH, Fox B, Kaizer H et al.
 403 Controlled trial of fluconazole to prevent fungal infections in patients undergoing bone
 404 marrow transplantation. N Engl J Med, 1992; 326(13): 845-851.
- [42] Slavin MA, Osborne B, Adams R, Levenstein MJ, Schoch HG, Feldman AR et al.
 Efficacy and safety of fluconazole prophylaxis for fungal infections after marrow
 transplantation- a prospective, randomized, double-blind study. J Infect Dis, 1995; 171(6):
 1545-1552.
- 409 [43] Martino R, Subira M. Invasive fungal infections in haematology: new trends. Ann
 410 Hematol, 2002; 1: 233-243.
- [44] Summerfield JA, Ryder S, Sumiya M, Thursz M, Gorchein A, Monteil MA et al.
 Mannose binding protein gene mutations associated with unusual and severe infections in
 adults. Lancet, 1995; 345: 886.
- [45] Koch A, Melbye M, Sorensen P, Homoe P, Madsen HO, Molbak K et al. Acute
 respiratory tract infections and mannose-binding lectin insufficiency during early childhood. J
 Am Med Assoc, 2001; 285: 1316.

- [46] Neth O, Hann I, Turner MW, Klein NJ. Deficincy of mannose-binding lectin and burden
 of infection in children with malignancy: a prospective study. Lancet, 2001; 358: 614-618.
- [47] Peterslund NA, Koch C, Jensenius JC, Thiel S. Association between deficiency of
 mannose-binding lectin and severe infections after chemotherapy. Lancet, 2001; 358: 637638.
- 422 [48] Summerfield JA, Sumiya M, Levin M, Turner MW. Association of mutations in
 423 mannose binding protein gene with childhood infection in consecutive hospital series. BMJ,
 424 1997; 314: 1229-1232.
- [49] Horiuchi T, Gondo H, Miyagawa H, Otsuka J, Inaba S, Nagafuji K et al. Association of
 MBL gene polymorphisms with major bacterial infection in patients treated with high-dose
 chemotherapy and autologous PBSCT. Genes Immun, 2005; 6: 162-166.
- 428 [50] Bodey GP, Buckley M, Sathe YS, Freireich EJ. Quantitative relationships between
 429 circulating leukocytes and infection in patients with acute leukemia. Ann Intern Med, 1966;
 430 64: 328-340.
- [51] Bergmann OJ, Christiansen M, Laursen I, Bang P, Hansen NE, Ellegaard J et al. Low
 levels of mannose-binding lectin do not affect occurrence of severe infections or duration of
 fever in acute myeloid leukaemia during remission induction therapy. Eur J Haematol, 2003;
 70: 91-97.
- 435 [52] Dean M, Minchinton RM, Heatley S, Eisen DP. Mannose binding lectin acute phase
 436 activity in patients with severe infection. J Clin Immunol, 2005; 25: 346-352.
- 437 [53] Mullighan CG, Heatley S, Doherty K, Szabo F, Grigg A, Hughes TP et al. Mannose438 binding lectin gene polymorphisms are associated with major infection following allogeneic
 439 hemopoietic stem cell transplantation. Blood, 2002; 99: 3524-3529.
- 440 [54] Vekemans M, Robinson J, Georgala A, Heymans C, Muanza F, Paesmans M et al. Low
- 441 mannose-binding lectin concentration is associated with severe infections in patients with

442 hematological cancer who are undergoing chemotherapy. Clin Infect Diseases, 2007; 44:443 1593-1601.

Kilpatrick DC, Mclintock LA, Allan EK, Copland M, Fujita T, Jordanides NE et al. No
strong relationship between mannan binding lectin or plasma ficolins and chemotherapyrelated infections. Clin Exp Immunol, 2003; 134: 279-284.

[56] Rocha V, Franco RF, Porcher R, Bittencourt H, Silva VA, Latouche A et al. Host defense
and inflammatory gene polymorphisms are associated with outcomes after HLA-identical
sibling bone marrow transplantation. Blood, 2002; 100: 3908-3918.

[57] Milstein DMJ, te Boome LCJ, Cheung YW, Lindeboom JAH, van den Akker HP,
Biemond BJ et al. Use of sidestream dark-field (SDF) imaging for assessing the effects of
high-dose melphalan and autologous stem cell transplantation on oral mucosal
microcirculation in myeloma patients. Oral Surg Oral Med Oral Pathol Oral Radiol Endod,
2010; 109: 91-97.

[58] Papp M, Altorjay I, Vitalis Z, Tornai I, Palatka K, Kacska S et al. Mannose-binding
lectin deficiency confers risk for bacterial infections in a large Hungarian cohort of patients
with liver cirrhosis. Journal of Hepatology, 2010; 53: 484-491.

[59] Thiel S. Complement activating soluble pattern recognition molecules with collagen-like
regions, mannan-binding lectin, ficolins and associated proteins. Mol Immunol, 2007; 44:
3875-3888.

[60] Beinrohr L, Dobo J, Zavodszky P, Gal P. C1, MBL-MASPs and C1-inhibitor: novel
approaches for targeting complement-mediated inflammation. Trends in Molecular Medicine,
2008; 14: 511-521.

464 [61] Botto M, Kirschfink M, Macor P, Pickering MC, Würzner R, Tedesco F. Complement in

465 human diseases: Lessons from complement deficiencies. Mol Immunol, 2009; 46: 2774-2783.

- 466 [62] Kilpatrick DC. Mannan-binding lectin: clinical significance and applications. Biochimica
- 467 et Biophysica Acta, 2002; 1572: 401-413.
- 468 [63] Holmskov U, Thiel S, Jensenius JC. Collectins and ficolins: humoral lectins of the
- 469 innate immune defense. Annu Rev Immunol, 2003; 21: 547-578.
- 470 [64] Ip WK, Takahashi K, Ezekowitz RA, Stuart LM. Mannose-binding lectin and innate
- 471 immunity. Immunol Rev, 2009; 230: 9-21.
- 472 [65] Turner MW, Hamvas RM. Mannose-binding lectin: structure, function, genetics, and
- 473 disease associations. Rev Immunogenet, 2000; 2: 305-322.
- 474 [66] Garred P, Larsen F, Seyfarth J, Fujita R, Madsen HO. Mannose-binding lectin and its
- 475 genetic variants. Genes Immun, 2006; 7: 85-94.
- 476 [67] Eisen DP, Minchinton RM. Impact of mannose-binding lectin on susceptibility to
 477 infectious diseases. Clin Infect Dis, 2003; 37: 1496-1505.
- 478 [68] Vekemans M, Georgala A, Heymans C, Muanza F, Paesmans M, Klastersky J et al.
- 479 Influence of mannan binding lectin serum levels on the risk of infection during chemotherapy-
- 480 induced neutropenia in adult haematological cancer patients. Clin Microbiol Infect, 2005; 11:
- 481 20.
- 482 [69] Kozlowska-Skrzypczak M, Gil L, Komarnicki M. Factors affecting neutrophil recovery
- 483 after autologous bone marrow-derived stem cell transplantation in patients with acute myeloid
- 484 leukemia. Transplantation Proceedings, 2009; 41: 3868-3872.
- 485 [70] Dykewicz CA. Summary of the guidelines for preventing opportunistic infections among
- 486 hematopoietic stem cell transplant recipients. Clin Infect Dis, 2001; 33: 139-144.
- 487 [71] Pearson ML. Guideline for prevention of intravascular device-related infections. Part I.
- 488 Intravascular device-related infections: on overview. Am J Infect Control, 1996; 24: 262-93.

- 489 [72] Wagner S, Lynch NJ, Walter W, Schwaeble WJ, Loos M. Differential expression of the
- 490 murine mannose-binding lectins A and C in lymphoid and nonlymphoid organs and tissues. J
- 491 Immunol, 2003; 170: 1462-1465.
- 492 [73] Seyfarth J, Garred P, Madsen HO. Extrahepatic transcription of the human mannose-
- 493 binding lectin gene (mbl2) and the MBL-associated serine protease 1-3 genes. Mol Immunol
- 494 2006; 43: 962-971.
- 495 [74] Milanese M, Segat L, Marziliano N, Crovella S. The expression of innate immunity496 genes
- 497 in Italian Crohn disease patients. Eur J Histochem, 2007; 51: 199-202.
- 498 [75] Petersen SV, Thiel S, Jensen L, Steffensen R, Jensenius JC. An assay for the mannan-
- 499 binding lectin pathway of complement activation. J Immunol Methods, 2001; 257: 107-116.

	Total	MBL <100 ng/ml	MBL >100 ng/ml
number of patients	186	21	165
patients with infections	168	19	149
infected/total (%)	90.3	90.5	90.3
number of infectious episodes	423	51	372
infectious episodes/ one patient	2.274	2.429	2.248
development of first infection (day, median, range)	6 [4;8]	7 [3;8]	6 [4;8]
mean follow-up (day)	331	343	329

bloodstream-infection	32 (7.6%)	3 (5.9%)	29 (7.8%)
fever, high CRP, severe mucositis	106 (25.1%)	15 (29.4%)	91 (24.5%)
upper respiratory tract infection	47 (11.1%)	6 (11.8%)	41 (11.0%)
lower respiratory tract infection	63 (14.9%)	12 (23.5%)	51 (13.7%)
oral mycosis	16 (3.8%)	1 (2.0%)	15 (4.0%)
herpes zoster	14 (3.3%)	1 (2.0%)	13 (3.5%)
HSV	7 (1.7%)	1 (2.0%)	6 (1.6%)
EBV	1 (0.2%)	0	1 (0.2%)
CMV	12 (2.8%)	1 (2.0%)	11 (3.0%)
GI tract disease	56 (13.2%)	7 (13.7%)	49 (13.2%)
elevated CRP level	42 (9.9%)	2 (3.9%)	40 (10.8%)
urogenital and other infection	27 (6.4%)	2 (3.9%)	25 (6.7%)

Table 1. The distribution of infections by MBL levels

	Control	Patients	NHL	HL	MM
case number	296	184	63	27	94
number of MBL- deficients	41	21	7	5	9
median MBL- level (ng/ml)	1067 [253.5;2121]	1479 [380.8;2849]	1623 [406.2;2847]	1365 [322.3;2850]	1338 [324.6;2902]
MBL deficient/ total (%)	13.9	11.4	11.1	18.5	9.6

Table 2. MBL levels of the examined and healthy population

	Total	NHL	HL	MM
number of patients	184	63	27	94
number of infectious episodes	415	186	67	162
infections/one patient	2.27	2.95	2.48	1.72
development of first infection (day, median, range)	6 [3;8]	4 [2.5;6]	4 [0;7]	8 [6;9]
grade of mucositis (mean)	1.44	1.56	1.5	1.34
MBLlevel(ng/ml)(median, range)	1479 [380.8;2849]	1623 [406.2;2847]	1365 [322.3;2850]	1338 [324.6;2902]
mean follow-up (day)	327	330	324	325

Table 3. Comparison of MBL levels and infections according to diagnosis

	Total	NHL	HL	MM
number of infectious episodes	415 (100%)	186 (100%)	67 (100%)	162 (100%)
bloodstream-infection	30 (7.2%)	10 (5.4%)	7 (10.4%)	13 (8.0%)
fever, high CRP, severe mucositis	95 (22.9%)	47 (25.3%)	16 (23.9%)	32 (19.8%)
upper respiratory tract infection	46 (11.1%)	18 (9.7%)	6 (9.0%)	22 (13.6%)
lower respiratory tract infection	62 (14.9%)	26 (14.0%)	12 (17.9%)	24 (14.8%)
oral mycosis	16 (3.9%)	7 (3.8%)	1 (1.5%)	8 (4.9%)
herpes zoster	13 (3.1%)	5 (2.7%)	3 (4.5%)	5 (3.1%)
HSV, EBV, CMV	20 (4.8%)	10 (5.4%)	1 (1.5%)	9 (5.6%)
GI tract disease	56 (13.5%)	30 (16.1%)	7 (10.4%)	19 (11.7%)
elevated CRP level	51 (12.3%)	21 (11.3%)	10 (14.9%)	20 (12.3%)
urinary tract and other infection	26 (6.3%)	12 (6.5%)	4 (6%)	10 (6.2%)

Table 4. The distribution of infections by diagnosis

Table

culture of central vein catheter	Total	MBL <100	MBL >100
number of patients	100	17	83
positive result of culture	25 (100%)	7 (100%)	18 (100%)
Staphylococcus epidermidis	10 (40%)	3 (42.9%)	7 (38.9%)
Staphylococcus coagulase negative	3 (12%)	1 (14.3%)	2 (11.1%)
Staphylococcus aureus	1 (4%)	0	1 (5.6%)
Enterococcus faecalis	5 (20%)	1 (14.3%)	4 (22.2%)
Streptococcus alpha- hemolising	1 (4%)	1 (14.3%)	0
Klebsiella pneumoniae	1 (4%)	1 (14.3%)	0
Pseudomonas aeruginosa	1 (4%)	0	1 (5.6%)
Acinetobacter baumannii	2 (8%)	0	2 (11.1%)
Bacillus	1 (4%)	0	1 (5.6%)

Table 5. Results of culture from central venous catheter

Blood culture	Total	MBL<100	MBL>100
number of patients	186	21	165
	55 (100%)	5 (100%)	50 (100%)
positive result of culture	(43 patient)	(4 patient)	(39 patient)
Staphylococcus epidermidis	17 (30.9%)	1 (20%)	16 (32%)
Staphylococcus hominis	5 (9.1%)	2 (40%)	3 (6%)
Staphylococcus hemolyticus	6 (10.9%)	1 (20%)	5 (10%)
Staphylococcus coagulase negative	9 (16.4%)	0	9 (18%)
Staphylococcus aureus	2 (3.6%)	0	2 (4%)
Enterococcus faecalis	4 (7.3%)	1 (20%)	3 (6%)
Streptococcus	3 (5.5%)	0	3 (6%)
Propionibacterium acnes	5 (9.1%)	0	5 (10%)
Pseudomonas aeruginosa	3 (5.5%)	0	3 (6%)
other Gram negative	1 (1.8%)	0	1 (2%)

Table 6. Results of blood culture according to MBL level



Figure 1. The distribution of MBL level in the examined patient group with hematological malignancies



Figure 2. Correlation between log MBL/CRP and log time of first infection

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

Dear Barry D. Kahan, PhD, MD, Editor-in-Chief, Transplantation Proceedings

Thank you for the review of my "Original Works or Clinical Submission" manuscript numbered TransProc2608 entitled "A New Approach to Predict the Chance of Infectious Complications Following Autologous Hematopoietic Stem Cell Transplantation: Mannose-Binding Lectin ELISA" for consideration for publication in Transplantation Proceedings.

Reviewer's comments were:

The authors report a prospective study examining mannose-binding lectin (MBL) levels and risk of autologous hematopoietic stem cell transplantation (HSCT). The results are interesting and provide more evidence about MBL levels as predictors of infection after HSCT.

The title of this manuscript is misleading for the novelty of the study, and should be changed. Mannose-Binding Lectin ELISA, which has been used in other previous studies, is not a new approach at all. The kit is commercial available too.

The changed title of the manuscript would be:

Evaluation of Mannose-Binding Lectin is a Useful Approach to Predict the Risk of Infectious Complications Following Autologous Hematopoietic Stem Cell Transplantation

Authors state the willingness and ability to pay all page charges, this document is uploaded again because of the changed Title of the manuscript.

I checked again my manuscript complies with the guidelines to authors. Title Page contain all author email addresses and the designated corresponding author. This Title Page is uploaded in the attached files area, along with this letter. Abstract, Text and References were double spaced, these are not changed. Content and form of the Text of manuscript is not changed.

Thank you very much for the extensive review and the intend to publish this manuscript as an Original article in the issue containing "Original Works or Clinical Submission" manuscripts in a future publication.

I am very grateful for your kind interest in this manuscript.

Sincerely,

Zita Brigitta Radnay MD.

Department of Hematology, Institute for Internal Medicine

Faculty of Medicine, University of Debrecen

Nagyerdei krt. 98.

H-4032 Debrecen, Hungary

Telephone number: +36-20-582-9147, Email address: radnayzita@gmail.com

TRANSPLANTATION PROCEEDINGS BARRY D. KAHAN, PhD, MD, Editor-in-Chief

<u>Editorial Office:</u> 11707 Trudeau Drive Houston, TX 77065 Telephone: 713-984-0533

Barry D. Kahan, PhD, MD - Editor-in-Chief Email: bkahan@transplantation-proceedings.org

THIS SIGNED FORM IS REQUIRED AND MUST BE UPLOADED WITH YOUR MANUSCRIPT UPON SUBMISSION THROUGH EES. WE WILL NOT PROCEED WITH YOUR MANUSCRIPT REVIEW UNLESS THIS FORM IS INCLUDED.

MANUSCRIPT RECEIPT - FINANCIAL AGREEMENT

Title Page With ALL Author Email Addresses: 2

Submitted Text Pages: 21

Abstract Included Yes

Submitted Tables: 6 Submitted Figures: 2

2

Total Pages Submitted (excluding Title Page and Abstract): 29

Manuscript Title:

Evaluation of Mannose-Binding Lectin is a Useful Approach to Predict the Risk of Infectious Complications Following Autologous Hematopoietic Stem Cell Transplantation

By submission of this manuscript to *Transplantation Proceedings*, I acknowledge I have read the <u>Guidelines to Authors of Manuscripts Submitted As an Original Work</u> and agree with the contents, and that I have attached a <u>completed and signed</u> Authorship And Conflict Of Interest Statement (ACIS) <u>on behalf of each author</u> listed on this manuscript.

I acknowledge that if accepted, I am responsible for all manuscript page charges, which will be billed to me by Elsevier, the publisher of *Transplantation Proceedings*, at the rate of <u>US\$99.95 per submitted manuscript page</u>, understanding that each Table and Figure will count as one manuscript page each along with the text. I understand that page charges are based on the typed, submitted page, not on the printed page, and that THREE complimentary pages are automatically provided by *Transplantation Proceedings* for manuscripts accepted as an original work to be published in one of our dedicated issues. Authors will be contacted with a tracking number, the number of pages confirmed, and will be informed of the number of pages for which they are responsible. Further, I understand that use of color reproduction of graphics will result in an additional charge. The Abstract and Title page are complimentary by *Transplantation Proceedings*.

Additionally, I agree that this manuscript has not been submitted or published in any other journal, including *Transplantation Proceedings*, and no parts of the manuscript are duplicated. I understand that if the manuscript is accepted for publication, copyright of the manuscript is transferred to Elsevier.

Tita hadne

Signature of Corresponding Author

Signature of Financially Responsible Party

21TA RADNAY

1

ARPADILLES MD, DSCI

Printed Name

Printed Name