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### THE HOST PREFERENCE OF PPV ISOLATES IN A HUNGARIAN ORCHARD

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#### SUMMARY

Plum pox virus (PPV) is the causative agent of Sharka disease, the most devastating viral disease on Prunus species. The first report was published by Atanasoff in 1932 from Bulgaria. The economically important host plants are the plum, peach and apricot species but show some disturbing studies occurrences on cherry and sour cherry as well. The variability of the virus shows that nine molecularly different PPV strains have been discovered. In Hungary the PPV-M, PPV-D and the PPV-Rec strains are in majority. Surveys from different countries suggest that the virus strains have some host preference or more frequent presence on different hosts.

0 36 12 2 m, PPV RT-PCR PPV-D. M. Rec : PPV. : PPV = Plum pox virus, M Marcus. D Dideron. Rec = Recombinant, RT = , PCR =

In this research 36 leaf samples were taken from naturally infected, 15-year-old plantation, 12 samples from each significant species (plum, peach and apricot) to investigate this attribute of the virus. In the orchard this three species located next to each other separated by a 2m wide road, resulted a serious chance for infection by any present virus strain.

The virus infection was verified from the leaf samples and the PPV strains were identified by conventional RT-PCR method. According to the results PPV-D, M, and Rec isolates, furthermore mixed infections were detected.

**Key words**: PPV, strains, host preference, plum, peach, apricot

**Abbreviations:** PPV = Plum pox virus, M = Marcus, D = Dideron, Rec = Recombinant, RT = reverse transcription, PCR = polymerase chain reaction

# Plum pox virus (+ssRNA) Potyviridae.

1917-1918

1932

Szirmai (1948), Husz Klement (1950) Németh (1963). PPV

(Pribék et al., 2001), (Salamon and Palkovics, 2002),

#### INTRODUCTION

Plum pox virus (+ssRNA) is a member of the Potyviridae family. The first symptoms of the presence were observed in 1917-1918 on plum in Bulgaria, and the first report was taken in 1932 by Atanasoff.

In Hungary the first report from the economically important stone fruits was taken by Szirmai from apricot (1948), by Husz and Klement from plum (1950) and by Németh from peach (1963). PPV infects not only stone fruits but almond (Pribék et al., 2001) and blackthorn (Salamon and Palkovics, 2002) as well, which is a natural wild host species endanger orchards as a reservoir.

. .) **Prunus** Mvzus Aphis spiraecola, persicae Aphis fabae **Aphis** hederae (Labonne et al., 1995). PPV PPV-M. PPV-D PPV-Rec (Ádám et al., 2015). (Myrta and Boscia, 2001), Dallot et al. PPV-D (1998).PPV-M (Myrta et al., 1998), PPV-D. PPV-Rec Prunus)

The importance of the reservoir plants are underrated, the aphid vectors spread the virus in a non-persistent manner.

The aphids play the most important role in the short distance spread of the virus and the form epidemics. The most effective vectors are the Myzus persicae and the *Aphis spiraecola*, during the dissemination the role of the aphids from non Prunus plants could be equally significant such as the Aphis fabae or the Aphis hederae (Labonne et al., 1995).

The three common strains in Hungary the PPV-M, PPV-D and the PPV-Rec (Ádám et al., 2015). The Dideron isolate origins from France, and it was collected from an apricot tree. The PPV-D strain mainly occurs on plum and apricot, and it is rare on peach (Myrta and Boscia, 2001), however Dallot et al. describe some PPV-D isolates that can effectively infect peach trees (1998).

The Marcus isolate collected from peach, origins from Greece (Myrta et al., 1998). The PPV-M strain is widely spread all over Europe and cause serious problems in peach orchards, but it can also infect plum and apricot. Aphids can propagate this strain more effectively than PPV-D.

Most of the PPV-Rec isolates are from plum host, but the apricot and peach (and different *Prunus* hosts) can be infected by artificial inoculation.

PPV-Rec PPV-D (Glasa et al., 2004). It seems that the host preference of the PPV-Rec isolates is similar to PPV-D (Glasa et al., 2004).

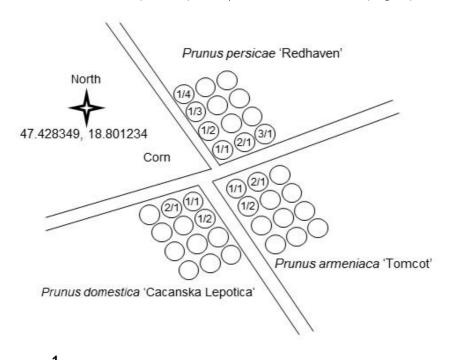
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#### MATERIAL AND METHODS

Twelve leaf samples were collected from *Prunus domestica* 'Cacanska Lepotica' on *Prunus cerasifera* (myrabolan) rootstock, from *Prunus persicae* 'Redhaven' on *Prunus amygdalus* (almond) rootstock and from *Prunus armeniaca* 'Tomcot' on myrabolan rootstock in the summer of 2015. The 36 samples were originated from different specimens but from the same orchard near Budapest.

The plum, peach and apricot trees were separated by a 2 metres wide road (Fig. 1).



. 1).

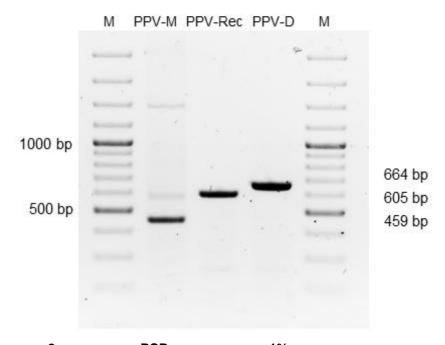
Fig. 1. The experimental field

Spectrum Plant Total RNA Kit (Sigma-Aldrich) GeneJet Plant RNA Purification Mini Kit (Thermo Scientific), (Thermo Scientific) 1500 ng (Maiss et al., 1989), M4T (Chen and Adams, 2001) **PPV PPV** mD5, mM3 mD3 mM5, 1) (Subr et al., 2004) PCR, 3' NIb. 1 % **PCR** . 2).

RNA extraction was by Spectrum performed **Plant** Total RNA Kit (Sigma-Aldrich) and GeneJet Plant RNA Purification Mini Kit (Thermo Scientific) according to the manufacturer's instruction. For cDNA preparation bν RevertAid Reverse Transcriptase (Thermo Scientific) 1500 ng RNA were used. In the RT reaction (Maiss et al., 1989) the reverse primer was the M4T (Chen and Adams, 2001) located at the 3' end, at the polyA tail of the virus. To confirm the presence of PPV, and for the identification of the three most common PPV strains in Hungary the mM5, mD5, mM3 and the mD3 primers (Table 1.) were used (Šubr et al., 2004) in the PCR, targeted recombination breakpoint located in the 3' end of the NIb gene. The results were visualised by gel electrophoresis on 1 % agarose gel. By this method the tree strain have different PCR products in length (Fig. 2.).

Table 1. The primer orientations, the sequences and the targeted genomic regions

( )		
Primer (orientation)	Sequence (5'-3')	Genomic region
M4T (-)	GTTTTCCCAGTCACGACT <sub>(15)</sub>	polyA tail
mM5 (+)	GCTACAAAGAACTGCTGAGAG	3'NIb-5'CP
mM3 (-)	CATTTCCATAAACTCCAAAAGAC	3'NIb-5'CP
mD5 (+)	TATGTCACATAAAGGCGTTCTC	3'NIb-5'CP
mD3 (-)	GACGTCCCTGTCTCTGTTTG	3'NIb-5'CP



. 2. PCR 1% Fig. 2. The PCR product of the three strains on 1 % agarose gel

Plum pox virus

36

PPV-D, PPV-M PPV-Rec RT-PCR.

(20%), PPV-M (28%), PPV-D (16.5%).

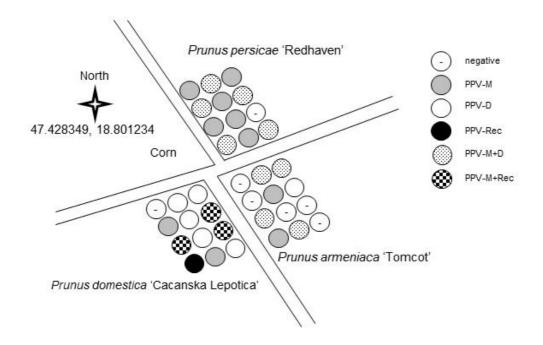
PPV-

Rec (2.5%).

PPV-M+D (25%), PPV-M+Rec (9%) ( . 3).

### **RESULTS AND DISCUSSION**

The results of the study indicate that the Plum pox virus was present in the orchard on the investigated plum, peach and apricot trees. In the 36 samples PPV-D, PPV-M and PPV-Rec strains were detected by conventional RT-PCR. Seven samples were negative (20%), ten was individually PPV-M infected (28%), six was PPV-D infected (16.5%) and in one case PPV-Rec infection was observed (2.5%). Nine PPV-M+D mixed infection (25%), and three PPV-M+Rec infection (9%) were identified (Fig. 3) among the trees.



. 3. Fig. 3. The results on the experimental field

. 50% PPV-M, PPV-M 42% PPV-M+D. PPV-D PPV-Rec 42% PPV . 16% PPV-M, 8% PPV-D. 32% M+D. PPV-M D 50-50%

In case of peach samples 50% of the studied trees were infected by only PPV-M, but in eleven samples PPV-M was detected as in 42% PPV-M+D infection occurred. PPV-D isolates were not detected solo in peach trees, and PPV-Rec isolate was not detected at all.

42% of the examined apricot samples were negative to these three PPV strains. 16% of the samples were infected by PPV-M, 8% were infected by PPV-D strain individually. In case of apricot 32% of the samples contain M+D mixed infection, according to our results the PPV-M and D strains occur approximately 50-50% on the

, . 40%

PPV-D, 16% PPV-M 8% PPV-Rec. (24%) PPV-M+Rec.

, PPV-M. 12 , 11

PPV-D

PPV-M+D.

PPV-Rec

SharCo, 223

examined apricot trees. Recombinant isolate was detected neither on apricot nor on peach trees.

From plum samples one negative result was observed, and all of the three strains were identified. 40% of the mula samples were infected by PPV-D, 16% with PPV-M and 8% was PPV-Rec infected. In three cases (24%) PPV-M+Rec infection was verified.

#### **CONCLUSIONS**

Among the examined peach trees the PPV-M strain was in majority as it was expected. From 12 trees 11 were infected by this strain and the one negative sample was originated a tree which was in very bad condition. Despite the PPV-D isolates are not effectively transmittable by aphids and not too common on peach, almost the half of the studied peach samples were infected by this strain in a mixed infection with PPV-M.

Naturally PPV-Rec infected peach trees are very rare according to studies (e.g. in SharCo project from 223 recombinant isolates one originated from peach), and in connection with these results we do not manage to detect recombinant isolates in the samples.

It is hard to make conclusion in regards of apricot trees, because almost the half of the samples were negative. In the PPV-D PPV-M

PPV-D,

PPV-M

M. persicae, 50-50% Marcus Dideron.

PPV-M+Rec.

D+M

M+Rec.

M: 26 22

M; 36 , 22

. 15 PPV-D PPV-Rec.

PPV

PPV-

virus infected trees PPV-D and PPV-M strains occur individually and in mixed infection.

The expectation was PPV-D dominance, but in this part of the orchard PPV-M could spread easily to apricot trees from peach by the most effective vector *M. persicae*, result the approximately 50-50% infection by Marcus and Dideron strains.

In this study recombinant isolate was not identified on apricot.

On the plum trees the three strains common in Hungary were identified as it was expected, and PPV-M+Rec mixed infection was detected on this part of the orchard. It is interesting that among the peach and apricot samples D+M mixed infection was frequent; in the plum samples only M+Rec was detected. Recombinant isolates only occur on plum in this experiment.

According to our results in the experimental orchard the PPV-M strain is dominant; from 36 samples 22 contain this type of isolate in solo or mixed infection. 15 samples were infected by PPV-D strain and in 4 samples PPV-Rec were detected.

This number of samples is too low for serious conclusions, but the host preference of PPV strains mentioned by studies seems to be confirmed.

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