THE IN VITRO EFFECT OF GARDEN THYME (THYMUS VULGARIS L.) EXTRACT ON BRACHYSPIRA HYODYSENTERIAE

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Filtrates of a water extract of commercially available garden thyme (Thymus vulgaris L.) were used for studying its possible bactericidal effect on Brachyspira hyodysenteriae, the causative agent of swine dysentery, by agar-diffusion technique. Five of the six studied Brachyspira strains have proven to be sensitive and one moderately sensitive in the in vitro tests. It was concluded that water extract of garden thyme possesses inhibitory effects against B. hyodysenteriae. In vivo experiments are needed to check the validity of this conclusion.

Keywords: garden thyme (Thymus vulgaris L.), Brachyspira hyodysenteriae, bactericidal effect

Introduction

For decades, increasing interest has been seen for using natural materials, mainly of herb origin, in human and veterinary medicine and in the food, preserving, and cosmetic industries. Numerous reports have been published that describe the composition and effects of these substances [1, 2]. Aromatic herbs and their extracts have been used for ages in traditional systems of medicine [3], and their bactericidal effects have also been known for a long time [4].

Thymus vulgaris L. (garden thyme) is an aromatic perennial herbaceous plant, which belongs to the genus Lamiales and family Lamiaceae. The plant is native to Mediterranean countries, where it blossoms in the period of May–July. In Central European countries, the plant can be cultivated.

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The herb contains 1.0%–2.5% volatile oils. The main and most active ingredients of the oil are thymol and its isomer carvacrol, which occur in the oil in a minimum of 40%, with thymol being dominant [5].

Beside volatile oils, thyme also contains several other bioactive substances, such as carotenoids, polyphenols, flavonoids, caffeic and rosmarinic acids, carbohydrates, and triterpenes, which possess free-radical entrapping and anti-oxidative capacity [5–8]. Recently, their anti-inflammatory effect has also been proven [9, 10].

The methanolic or aqueous extracts and the essential oil of thyme proved to be antibacterial and antifungal when tested on Gram-positive and Gram-negative bacteria, fungi, and yeasts, e.g., Candida albicans [5]. The effect is seen also when different essential oils were combined [11].

According to our knowledge, no data are available on the effect of thyme against Brachyspira strains. This prompted us to investigate the in vitro effect of the water extract of thyme on the causative agent of swine dysentery (Brachyspira hyodysenteriae).

Materials and Methods

Water extract

A cold (25 °C) water extract of the herb was produced. Commercially available varieties of T. vulgaris L. were ground to powder of which 3 g was added to 100 ml of distilled water then shaken for 1 h in a bolter. The mixture was ultra sieved through ultra-filter membrane columns (KOCH/Romicon WF2, Hollow Fiber) with a pore size of 500 kDa (filtrate A) and 100 kDa (filtrate B). The filtrates were then used for preparing dilutions up to 1:20,000 concentrations.

Microorganism

The investigation has started with a dose-ranging study involving a strain of B. hyodysenteriae (B/01) isolated from a clinically ill fattening pig. Further to this, five other strains originating from the collection of the Department of Microbiology and Infectious Diseases of the Faculty of Veterinary Science, SZIE University have also been included in the investigation.

Isolation and culture media

All six strains were isolated from pigs that showed clinical signs of swine dysentery in different territorial regions of Hungary. Affected pigs were
transferred to, then stunned and bled in slaughterhouses. Isolated parts of affected colon sections were transferred to a pathological laboratory. Small amount of colon mucosal scrapes were dispersed over the surface of freshly prepared tryptone–soy culturing media (Sigma-Aldrich, Saint Louis, USA) that contained 10% defibrinated bovine blood and 400 μg/ml spectinomycin (Sigma-Aldrich, Saint Louis, USA). After inoculation, the plates were incubated for 96 h at 42 °C with strict anaerobic conditions. Anaerobic conditions were safeguarded with gas-generating bags (Oxoid Gas Generating Kit, Anaerobic system BR0038B; Oxoid Ltd., UK) and anaerobic jars (Oxoid Ltd., UK). Determination and identification of the primary and secondary biochemical characteristics of the strains were carried out according to standard methods [12]. The isolated strains were stored deep frozen (−80 °C) in tryptic soy broth that contained 25% sterile glycerine (Scharlab, S.L., Barcelona, Spain).

**Agar-diffusion technique**

Blood agar (modified tryptone–soy agar + 10% defibrinated bovine blood) was used for producing protocultures from thawed mass of bacteria. From each of the protocultures, ten agar cubes of 5 × 5 mm (±5% deviation) were cut out from the areas showing strong hemolysis. The agar-cube inoculums were spread by sterile glass rod over freshly prepared agar plates of 90 mm diameter. The plates were closed and dried for 5 min.

According to pre-determined order, a hole of 5 mm diameter was cut centrally into the inoculated plates. About 100 μl of either filtrate of *T. vulgaris* L. extract was measured into the holes. The plates were placed into anaerostat (Oxoid Ltd., UK) within 15 min after dripping the known dilution of herb extract into the holes and incubated for 4–5 days at 37 °C. Inoculations were done in triplicate regarding each dilution per strain.

Sensitivity of *Brachyspira* strains to the given dilution of the herb extract was estimated on the basis of the average diameters of the growth-inhibition zones that formed around the holes. The inhibition of growth was indicated by the lack of hemolysis, as opposed to a clearance in the blood agar in the areas where growth and hemolysis took place. On this basis, the strains were assorted into three groups as follows:

1. Sensitive (S): Average diameter of the inhibition ring is >25 mm, which refers to the *in vitro* sensitivity of the strain to the given dilution of the herb extract.
2. Moderately sensitive (MS): Average diameter of the inhibition ring is between 15 and 25 mm, which refers to the questionable effect of the extract at the given solution.
(3) Resistant (R): Average diameter of the inhibition ring is <15 mm. This expresses the *in vitro* inefficiency of the herb solution.

**Results**

*B. hyodysenteriae* is a Gram-negative spirochete, which grows in anaerobic conditions and forms strong beta hemolysis beneath and around the colonies.

Both herb extracts inhibited the growth of *B. hyodysenteriae* strains proportionally to the dilution rate of the extracts (Table I). As it is seen from Table I, the 1:20,000 dilution of the sample A was found ineffective while the same dilution of the other extract (B) produced MS reaction.

On the basis of this preliminary data, the sample B has proven superior to the other one; therefore, the sensitivity reaction of *B. hyodysenteriae* strains was tested with the 1:5,000 dilution of this sample. Data are disclosed in Table II.

As shown by the data, five of the six *B. hyodysenteriae* strains (83.3%) have proven sensitive and one strain (167/6) gave MS reaction.

**Discussion**

Microbicide and fungicide effects of essential oils and different extracts of garden thyme have been studied for a long time. Their efficiency against a large variety of pathogens and food-borne germs has been proven by recent investigations [11, 13, 14].

**Table I.** Effect of different dilutions of garden thyme herb extracts on a *B. hyodysenteriae* strain (B/01) in agar-diffusion tests

<table>
<thead>
<tr>
<th>Samples (garden thyme extracts)</th>
<th>Degree of dilution</th>
<th>Average diameter of the inhibition ring (mm)</th>
<th>Evaluation</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>500x</td>
<td>50</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>1,000x</td>
<td>45</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>2,500x</td>
<td>36</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>5,000x</td>
<td>35</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>10,000x</td>
<td>30</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>20,000x</td>
<td>15</td>
<td>R</td>
</tr>
<tr>
<td>B</td>
<td>500x</td>
<td>60</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>1,000x</td>
<td>50</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>2,500x</td>
<td>47</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>5,000x</td>
<td>40</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>10,000x</td>
<td>28</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td>20,000x</td>
<td>25</td>
<td>MS</td>
</tr>
</tbody>
</table>

*Note:* S = sensitive; MS = moderately sensitive; R = resistant.
Several of the studies [1, 15, 16] indicated the sensitivity of Gram-positive bacteria, while others [17–19] have shown expressed sensitivity of Gram-negative bacteria.

In our investigation, a strong dose-dependent effect was observed. This finding is in good agreement with other reports [20–22] involving microbes other than B. hyodysenteriae. It is a limitation of our study that the active agents have not been identified and therefore the study is to be viewed as a pilot trial. Antibiotic resistance of the strains was not tested, nevertheless, the results serve as a promising basis for further studies conducted in vivo and on strains with known antibiotic resistance spectrum and known cell count.

Effective substances of garden thyme also affect the antibiotic-resistant microbes [23–26]; in certain cases, they might replace antibiotics [27–29] and might present sound alternative to growth promoters [4, 30]. These joint effects improve food safety and sustainability of animal agriculture.

### Conclusion

The in vitro investigation reported above seems to support our earlier presumption that water extract of garden thyme possesses antagonistic effects to B. hyodysenteriae. This conclusion, however, should be controlled with further, first of all in vivo experiments in the light that results of the in vitro and in vivo experiments might be different [31].

### Conflict of Interest

The authors declare that they have no conflict of interest.

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**Table II.** Effect of different dilutions of garden thyme herb extracts on different B. hyodysenteriae strains in agar-diffusion tests

<table>
<thead>
<tr>
<th>B. hyodysenteriae strains</th>
<th>Sample B and rate of dilution</th>
<th>Average diameter of the inhibition ring (mm)</th>
<th>Evaluation</th>
</tr>
</thead>
<tbody>
<tr>
<td>167/6</td>
<td>5,000x</td>
<td>25.0</td>
<td>MS</td>
</tr>
<tr>
<td>168/6</td>
<td>5,000x</td>
<td>26.0</td>
<td>S</td>
</tr>
<tr>
<td>169/6</td>
<td>5,000x</td>
<td>31.0</td>
<td>S</td>
</tr>
<tr>
<td>170/6</td>
<td>5,000x</td>
<td>32.0</td>
<td>S</td>
</tr>
<tr>
<td>171/6</td>
<td>5,000x</td>
<td>29.0</td>
<td>S</td>
</tr>
<tr>
<td>B/01</td>
<td>5,000x</td>
<td>38.0</td>
<td>S</td>
</tr>
<tr>
<td>Average</td>
<td></td>
<td>30.2</td>
<td>S</td>
</tr>
<tr>
<td>SD ±</td>
<td></td>
<td>4.7</td>
<td></td>
</tr>
</tbody>
</table>

*Note: S = sensitive; MS = moderately sensitive; R = resistant; SD = standard deviation.*