Effect of Ozone and Antioxidants on Wheat and its Pathogen – *Bipolaris sorokininana*

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> (Received 8 January 2016; Accepted 8 June 2016; Communicated by J. Kolmer)

Tropospheric ozone (O₃) adversely affects growth and productivity of crops and also influences crop–pathogen interactions. Adverse effects of O₃ on crops can be mitigated by antioxidants application. In the present study through lab and field experiments impact of O₃ and antioxidants treatment to wheat was assessed on growth of *Bipolaris sorokiniana* (BS-75 strain) pathogen responsible for Spot blotch disease, pathogenesis related (PR) proteins and chitinase content. Results showed that growth of *Bipolaris* was significantly higher in elevated ozone (EO₃) exposed plants as compared to control plants. Antioxidants – ascorbic acid (AA), tagetes extract (T) and quercetin (Q) application on culture media and wheat plants, respectively, retarded the growth of *Bipolaris sorokiniana*. Among the three antioxidants minimum growth of *Bipolaris* was observed in AA-treated plants as compared to control plants. Reduction in chitinase activity and PR proteins content due to EO₃ treatment in wheat plants was 18% and 78%, respectively, as compared to control plants. Increase in chitinase activity and PR proteins content in wheat plants was 45% and 60%, respectively, as compared to control plants.

Keywords: ozone, wheat, pathogen, Bipolaris sorokiniana

Introduction

Tropospheric ozone (O_3) a secondary air pollutant, negatively affects agricultural crops and vegetation (Holland et al. 2006; Fuhrer 2009; IPCC 2013). In India also phytotoxic impact of O_3 on growth and yield of several crops were reported (Mina et al. 2010; Ghude et al. 2014). Besides the direct adverse effects of O_3 on crops, indirectly it alters the interaction between crops and its pest/pathogens (Manning and von Tiedemann 1995; Elad and Pertot 2014). Wheat (*Triticum aestivum* L.) is a staple food crop of India and its spot blotch disease caused by *Bipolaris sorokiniana* responsible for up to 22% and 10–50% yield losses in warm and humid regions of the world and India, respectively (Saari 1998; Chowdhury et al. 2013). Several reviews have extensively covered possible effects of elevated O_3 on future severity of plant diseases (Manning and von Tiedemann 1995; Elad

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and Pertot 2014; Mikkelsen et al. 2015). Alteration in pathogenesis potential of pathogen due to O_3 exposure is of ecological and economical importance. Reports from India on response of O_3 predisposed wheat crop to any pathogens are completely lacking.

Adverse effects of O_3 on crops can be mitigated through application of O_3 protectants containing synthetic or natural antioxidants (Puckette et al. 2008). Among antioxidants, natural antioxidants were preferred more than synthetic antioxidants for mitigating O_3 stress effect because of their environmental safety features. Application of antioxidants on crops to protect them from O_3 adverse impact may also affect their interaction with its pathogen, which was not monitored yet. The objective of this study was to examine effect of O_3 and antioxidants, namely ascorbic acid (AA), quercetin (Q) and tagetes leaves extract (T), on growth of *Bipolaris sorokiniana* (BS-75 strain), severity of blotch disease and defence response of wheat plants through lab and field experiments.

Materials and Methods

Experiment location

Experiments were conducted at the research field and laboratory of Indian Agriculture Research Institute (IARI, 28°35' N, 77°12' E), New Delhi during rabi season of 2011–2012 and 2012–2013.

O₃ and antioxidants pretreatment to wheat

Wheat (*Triticum aestivum* L.) cv. PBW343 was sown in pots on 12 December 2011 and 15 December 2012 by following recommended standard practices. One set of wheat plants were pretreated with different O_3 levels (Table 1) in open top chambers (OTCs; 90×120 cm dimension, made of transparent sheet and installed in open field) for 8 hours per day from seedling to 65-day-old stage. O_3 in OTCs was supplied by O_3 generator (Standard Appliances, Varanasi, India). O_3 concentration in OTCs was monitored with an online ozone analyzer. Plant leaves were monitored regularly for O_3 injury. Another set of wheat plants, since five, leaf stage were treated by 1% AA, 10% T and 500 ppm Q, respectively, at 15 days interval till 65-day-old stage in OTCs supplied with NF air. Control wheat plants were not given any antioxidant treatment.

Treatment	Abbreviated as	O ₃ concentration
Ambient O ₃ *	AO ₃	12–72 ppb
Non-filtered air**	NF	5–10% lower than ambient O_3
Elevated level O ₃	EO3	ambient O ₃ +25-30 ppb
Charcoal filtered air	CF	$65-70\%$ lower than ambient O_3

Table 1. Ozone levels maintained in OTCs to expose wheat plants

*Plants were exposed to ambient/naturally prevailing levels of ozone outside OTC.

**Plants were exposed to ambient air without filtration inside OTC.

Bipolaris sorokiniana inoculum

Isolate BS-75 of *Bipolaris sorokiniana* (Sacc.) Shoem., procured from the Department of Plant Pathology of the IARI, New Delhi (India) was used in the study. Mycelium was grown on potato dextrose agar (PDA, Sigma) for 3 weeks at 25 °C in the dark. Conidia and hyphae were scraped off from medium with spatula and mixed with sterile distilled water. Conidial suspensions filtered through cheesecloth to eliminate mycelial fragments. Inoculum densities were adjusted to $2-2.5 \times 10^3$ conidia/ ml.

Bipolaris growth on antioxidants supplemented culture media

A 3 mm diameter disc of 15-day-old culture of *Bipolaris sorokiniana* (strain BS-75) was cultured at 25 °C in replicates on Petri dishes of 90 mm diameter containing solidified 15 ml potato dextrose agar (PDA) medium supplemented with antioxidants (Table 2). The plates were observed for colony diameter and sporulation till 7 days incubation period.

Bipolaris growth on O3 and antioxidants pretreated wheat

After 65 days O_3 treatment, on 66th day wheat plants were taken out of the OTCs for *Bipolaris* inoculation. Six healthy tillers per pot were inoculated in evening before sunset by spraying 15 ml conidial suspension per tiller. Immediately after inoculation plants in each pot were covered with a transparent, needle pricked polythene bag (0.06 mm thickness) for 48 hours. The film blocked wind and maintained moisture inside while transmitting long-wave radiation, consequently dew formation on the wheat leaves surface was enhanced. Plants returned to OTC after inoculated plants for 7 days were number of lesions, lesion size, latent period (number of days between inoculation and appearance of first mature conidia), sporulation and spores per 25 mm².

Antioxidant	Concentration	Abbreviated as
Ascorbic acid	0.5%	AA1
Ascorbic acid	1%	AA2
Ascorbic acid	1.5%	AA3
Quericitin	250 ppm	Q1
Quericitin	500 ppm	Q2
Quericitin	750 ppm	Q3
Tagetes leaves extract	5%	T2
Tagetes leaves extract	10%	T2
Tagetes leaves extract	15%	Т3

Table 2. Concentrations of antioxidants supplement in PDA agar

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PR protein content and chitinase activity in O_3 and antioxidants pretreated and Bipolaris infected wheat

The colorimetric assay of chitinase activity was carried out according to Morgan and Elson (1934). One enzyme unit (EU) activity is the amount of enzyme which produces 100 µg NAG/ml chitin solution in 1 h at 37 °C at pH 5.2. The total protein extraction was first done from wheat leaves using total protein extraction kit supplied by G-Biosciences (Cat # 786-225). From the extracted total protein, PR proteins were quantitatively estimated by using the PR ELISA Kit (Catalog No. E0431 pl) supplied by USCN LIFE. The PR proteins content and chitinase activity were monitored at the end of O_3 and antioxidants treatment (i.e. 65 DAS), after 7 days of incubation period in flag leaves of plants with and without *Bipolaris* pathogen.

Statistical analysis

Data obtained from the experiments conducted in both years were pooled and subjected to homogeneity test. The pooled data subjected to single factor (O₃ and antioxidant) analysis of variance (ANOVA) and least-significant differences were calculated at $P \le 0.05$.

Results

Environmental conditions

Ambient O₃ levels at the experimental site ranged between 12–72 ppb during wheat crop growing period. Average temperature and average relative humidity at the experimental site during the crop growth period were 20 ± 2.5 °C during day, 15 ± 2.0 °C at night and $50 \pm 60\%$, respectively. Ozone injury symptoms on wheat plants were not visible in any treatment, except EO₃. EO₃ exposed plants exhibited 8–10% O₃ injury lesions on old leaves.

	С	AA1	AA2	AA3	Q1	Q2	Q3	T1	T2	Т3
Colony diameter (mm)										
After 2 days	0.8ª	0.5 ^b	0.2 ^b	0 ^b	0.7 ^b	0.6 ^b	0.3 ^b	0.2 ^b	0.3 ^b	0.5 ^b
After 5 days	6.1ª	1.0 ^b	0.6 ^b	0 ^b	5.7ª	4.3 ^b	4.6 ^b	2.9 ^b	1.7 ^b	1.6 ^b
After 7 days	8.0ª	1.5 ^b	0.1 ^b	0 ^b	7.8ª	6.9ª	6.0 ^b	3.9 ^b	2.5 ^b	1.9 ^b
Sporulation 7 days after	++	_	-	-	++	+	+	+	-	_

Table 3. Effect of antioxidants on Bioplaris growth

Mean values marked with the different letters are statistically significantly different among treatments ++ number of conidia between 1.5×10^8 - 2.5×10^8 /ml; + number of conidia between 0.5×10^8 - 1.0×10^8 /ml.

Treatment	Growth parameter ^a	DF	Replication	F ratio	Significance level ^b
O ₃ pretreatment to wheat in OTCs	No. of lesions	36	10	2.6	.003**
	Maximum lesion size (mm)		10	6.5	.01*
	Latent period		10	2.5	.002**
	Spores/25 mm ²		10	1.9	.001***
Antioxidants pretreatment to wheat in OTCs	No. of lesions	36	10	3.5	.007*
	Maximum lesion size (mm)		10	4.1	.005**
	Latent period		10	1.4	.005**
	Spores/25 mm ²		10	9.8	.02*
Antioxidants supplementation in culture media	Colony diameter	36	5	5.1	.002**
	Sporulation		5	1.9	.01*

Table 4. ANOVA of O3 and antioxidants effects on Bipolaris growth

^aThe two uppermost leaves of each plant were evaluated; ^bSignificance levels; *P<0.05; **P<0.01; ***P<0.001.



Figure 1. Bipolaris growth on O₃ pretreated wheat. Values are means \pm SE (n = 10). Values marked with the different letters are statistically significantly different among treatments ($P \le 0.05$)

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Figure 2. Bipolaris growth on antioxidants pretreated wheat. Values are mean \pm SE (n = 10). Values marked with the different letters are statistically significantly different ($P \le 0.05$)

Bipolaris growth on antioxidants supplemented culture media

Antioxidants supplementation significantly affected growth of *Bipolaris sorokiniana* (Tables 3 and 4). Inverse relationship was observed between the antioxidants concentration and the *Bipolaris* growth. Among three antioxidants, AA was the most effective inhibitor for the growth of *Bipolaris sorokiniana* followed by T and Q, respectively, as compared to the control (Table 3).

Bipolaris growth on O₃ and antioxidants pretreated wheat

Effect of O_3 treatments on *Bipolaris* growth was variable. Lesions/leaf and spores/ 25 mm² were maximum and minimum on EO₃ and CF treated plants, respectively. Significant differences in *Bipolaris* lesions size were observed in NF (27%), AO₃ (50%) and EO₃ (90%) treatments as compared to CF (Table 4, Fig. 1). Shortening of latent period with increasing O₃ concentration was observed (Fig. 1). The 50% of EO₃, 25% of AO₃ exposed plants exhibited black spots with conidiophores on leaf surface within 7 days and after 10 days of incubation period, respectively. In CF exposed plants, disease symptoms were visible only on one plant after 20 days of incubation.

Bipolaris growth on wheat plants was adversely affected by antioxidants treatment (Table 4, Fig. 2). Among three antioxidants, *Bipolaris* growth was minimum on AA treated wheat plants followed by T and Q as compared to control plants. On AA, Q and T treated plants, spot blotch lesions appeared after 10, 4 and 6 days of incubation, respectively. Number of lesion in AA, Q and T treated wheat plant were 67%, 9% and 22% less, respectively as compared to control plants. Maximum size of lesion was 67%, 38% and

3% less on AA, Q and T treated wheat plants as compared to control plants. Latent period and Spores/25 mm² was maximum and minimum on AA treated and control wheat plants, respectively.

PR protein content and chitinase activity in O_3 and antioxidants pretreated and Bipolaris infected wheat

Exposure of O_3 significantly affected PR protein content in wheat. 2.74–12.83 ng/g leaf PR protein content was observed in O_3 -treated plants (Fig. 4a). Maximum PR protein content (9.8 ng/g leaf) in CF air plants without pathogen was observed (Fig. 3a). PR protein content (10.1 ng/g leaf) was maximum in AA treated plants and minimum (4 ng/g leaf) in control plants. Among antioxidant treated plants with pathogen and without pathogen, PR protein content was maximum (12.5 ng/g leaf) in AA treated with pathogen plants (Fig. 3b).



🖬 antioxidant (66 das) 🖸 antioxidant -pathogen (73 das) 🔟 antioxidant + pathogen (73 das)

Figure 3. PR protein content in wheat plants with and without pathogen (a) pretreated with O₃ (b) pretreated with antioxidant. Values are mean \pm SE (n = 10). Values marked with the different letters are statistically significantly different ($P \le 0.05$)

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Chitinase activity in EO₃ exposed plants was 27.3% less than CF exposed plants. Chitinase activity was significantly less in O₃ exposed plants with pathogen as compared to O₃ exposed plants without pathogen (Fig. 4a). Enhancement in chitinase activity was observed for wheat plant treated with antioxidants. Maximum (3.1 EU) and minimum (2.2 EU) activity was observed in AA and control plants, respectively. Among antioxidant treated plants with and without pathogen, the chitinase activity was more in antioxidant treated and pathogen infected plants. Plants treated with AA and with disease exhibited maximum (3.2 EU) enzyme activity (Fig. 4b).





Figure 4. Chitinase activity in wheat plants with and without pathogen (a) pretreated with O₃ (b) pretreated with antioxidant. Values are mean \pm SE (n = 10). Values marked with the different letters are statistically significantly different ($P \le 0.05$)

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Discussion

Exposure to O_3 stresses and pathogen infection induce a rapid accumulation of reactive oxygen species (ROS) in plant tissues. In non-stressed plant tissues enzymatic and nonenzymatic antioxidants are able to neutralize the harmful effects of ROS (Barna et al. 2012). If the abiotic or biotic stress is persistent and too severe, the antioxidants cannot neutralize the overproduction of ROS which leads to cell death and necrosis development. ROS-mediated death of plant tissues can increase or decrease susceptibility to pathogens (Shetty et al. 2008). Response of O₃ pretreated crops to pathogen is dependent on type of pathogen (necrotrophic, hemibiotrophic or biotrophic), O_3 concentration and the crops growth stage (Mikkelsen et al. 2015). In the present study enhancement in susceptibility of O₃ predisposed wheat to B. sorokiniana, a hemibiotrophic pathogen was observed. This observation confirms the Tiedemann (1992) reports that susceptibility of wheat to Septoria nodorum and B. sorokiniana after O₃ exposure increased. Mikkelsen et al. (2015) also reported increased and decreased infection by the hemibiotrophic spot blotch fungus and biotrophic powdery mildew fungus, respectively, under elevated O_3 , compared to ambient conditions, implying that O3_ induced changes in disease severity could be linked to the trophic lifestyle of the pathogens.

In plants, PR proteins prevent various pathogen invasion. Among these PR proteins chitinase and β -1,3-glucanase are two important hydrolytic enzymes play main role of defense reaction against fungal pathogen by degrading cell wall chitin (Ebrahim et al. 2011). In our study O₃ pretreated wheat plants exhibited reduction in PR protein content and chitinase activity as compared to control. This is contrary to previous studies where increase of PR proteins like chitinase and β -1-3-glucanase is reported in different crops on O₃ stress exposure (Thalmair et al. 1996). This can be related to variation in O₃ sensitivity of different cultivars. For example, near ambient ozone exposure induced β -1,3-glucanase (PR-2) activity 22-fold in the O₃ tolerant tobacco line Bel B while in O₃ sensitive line Bel W3 only a fourfold-increase in β -1,3-glucanase activity (Schraudner et al. 1994).

In present study it was observed that different levels of antioxidants exhibiting significant inhibitory effect against *Bipolaris* indicate that antioxidants may be acting as antimicrobial compounds. These observations were in agreement with Cowan (1999) explained that antioxidants as plant product inhibit microbes growth by enzyme inhibition mechanism.

 O_3 stress mitigating capacity of the three antioxidants (AA, Q, T) used in present study exogenously has not been reported in literature yet. The results shows that exogenous antioxidants application to wheat plants, inhibited growth of *Bipolaris* on them and enhanced synthesis of PR protein and chitinase. Our findings are in line with Didyk and Blum (2011), reported exogenous application of antioxidants limits foliar necrosis, biomass reduction and alteration of metabolism caused by O_3 stress. Querictin and tagetes leaf extract reduced foliar injury by 10–13% and 14%, respectively, in *Trifolium. subterraneum* exposed to ambient O_3 level for 28 days (Didyk and Blum 2011). Ascorbic acid application reduced the visible leaf injury by 50–89% and 100% in 45–55 ppb and 0–250 ppb O_3 exposed *Nicotiana tabaccum* (Chen and Gallie 2005). In literature it has been reported that application of exogenous ascorbic acid enhances plant tolerance to pathogens because of activation of general defense gene signaling network (Conklin and Barth 2004). However, Rice-Evans et al. (1997) reported that under *in vitro* conditions querce-tin (flavonoids) was more effective antioxidant than ascorbic acid.

Results of the study indicate that O_3 concentration and wheat susceptibility to *Bipola*ris sorokiniana are positively correlated. Antioxidants besides their O_3 stress mitigating effects also retards growth of *Bipolaris* through enhancing synthesis of defense compounds. More studies need to carry out to quantify the contribution of exogenously applied antioxidants in antimicrobial activity and strengthening of defense mechanism of plants compounds and activity. Our study suggests to prevent losses in wheat productivity due to synergistic effect of O_3 and hemibiotrophic pathogens proper package of agriculture practices for using antioxidants need to be developed.

Acknowledgement

Financial support from the Indian Council for Agricultural Research to Indian Agriculture Research Institute is gratefully acknowledged.

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