

## New Data to the Knowledge on the Corticulous Mite Fauna in Hungary (Acari: Prostigmata, Astigmata, Oribatida)

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Authors give a report on the results of the regular collections made between 1990 and 1998 on ornamental trees and shrubs, on streets, in parks, in green spaces of housing estates, in private gardens and in arboreta. A total of 19 corticulous mite species were found belonging to 14 families. *Michaelopus corticalis* (Michael, 1885), *Hericia hericia* Robin, 1868, *Eupalopsis maseriensis* (Canestrini et Fanzago, 1876), *Mediolata vandergeesti* (Gomaa et Bolland, 1982), *Cunaxoides kielczewskii* Michocka, 1982, and *Neophyllobius* spec. nov. are reported for the first time for the Hungarian fauna. *Hemisarcoptes budensis* Fain et Ripka, 1998, *M. corticalis* and *Cheletogenes ornatus* (Canestrini et Fanzago, 1876) were the most frequent species in the acarofauna associated with scale insects.

Key words: Corticulous Mites, Acari, Prostigmata, Astigmata, Oribatida

Among arboreal mites corticulous mites are less known contrary to foliage-dwelling mites. In the last three decades, Lebrun (1976), André (1986, 1987), Momen (1987, 1988), Momen and Lundqvist (1996), Suski (1972), Doberski (1986), Bolland (1983, 1986, 1991), Smiley (1992), Kehl and Weigmann (1992), Weigmann and Jung (1992) studied the mites on bark of trees and shrubs. Their results proved the considerable richness of corticulous acarofauna.

A part of cortex-dwelling mites is algae and lichens feeder. An other group is fungivorous and saprophagous. The third group is predaceous upon the smallest arthropods. In Belgium, André (1986) reported 22 species of Tydeidae and 6 species of Stigmaeidae from bark of different tree species. In Bulgaria, Nachev and Trenchev (1987), while in Italy, Vacante (1989) as well as Vacante and Gerson (1987) reported the mites found associated with coccids from fruit trees. Nachev and Trenchev (1987) observed *Thyreophagus entomophagus* (Laboulbene), *Cheletogenes ornatus* (Canestrini et Fanzago), *Cheyletia flabellifera* (Michael), *Hemisarcoptes malus* (Shimer), *Amblyseius cucumeris* (Oudemans) and *Bdella iconica* Berlese under the shields of *Sphaerolecanium prunastri* (Fonscolombe), *Parlatoria oleae* (Colvée), *Quadraspidiotus marani* Zahradnik and *Quadraspidiotus pyri* (Lichtenstein). *Th. entomophagus* was the most frequent species. Vacante (1989) reported *H. malus* from *Lepidosaphes beekii* (Newman), *Aonidiella aurantii* (Maskell), *Parlatoria pergandii* Comstock and *Aspidiotus nerii* Bouché. Several members of Stigmaeidae, Eupalopsellidae, Cunaxidae and Camerobiidae are predators of armored scale insects and can be found on the bark of trees (Cooreman,

1958, Meyer and Ueckermann, 1984, Bolland, 1986, Vacante and Gerson, 1987). Smiley and Knutson (1983), also Gerson (1973, 1985) summarized the knowledge on the role and biology of predatory mites of coccids. Fain et al. (1995) as well as Fain and Ripka (1998) reported new species of the genus *Hemisarcoptes* that predaceous upon diaspidids.

The present knowledge of corticolous mites in Hungary remains scanty. Balogh and Mahunka (1980) reported oribatids, Komlovszky (1980, 1984) recorded cunaxid and cheyletid species. Lucza et al. (1996) found 4 cheyletid species on tree bark infested with armored scale insects. Ripka et al. (1997) collected the following 6 tarsonemid species: *Tarsonemus nodosus* Schaarschmidt, 1959, *Tarsonemus lobosus* Suski, 1965, *Tarsonemus karli* Sharonov et Mitrofanov, 1986, *Tarsonemus waitei* Banks, 1912, *Dendroptus willmanni* (Schaarschmidt, 1959) and an other *Dendroptus* sp. together with coccids. *Aceria heteronyx* (Nalepa, 1891) causes small stem galls on twigs and branches of *Acer platanoides*. An other eriophyoid mite, *Acalitus phloeocoptes* (Nalepa, 1890) causes stem galls on *Prunus domestica* and *Prunus spinosa* (Ripka and de Lillo, 1997). Ripka and Kaźmierski (1998a) recorded the following 8 tydeid species in association with scale insects: *Lorryia reticulata* (Oudemans, 1928), *Lorryia obliqua* (Kuznetsov, 1973), *Lorryia* cf. *woolleyi*, *Triophtydeus immanis* Kuznetsov, 1973, *Triophtydeus triophthalmus* (Oudemans, 1929), *Tydeus caudatus* (Duges, 1834), *Tydeus californicus* (Banks, 1904) and *Homeopronematus staerki* (Schruff, 1972). Out of them *T. immanis* was the most frequent mite. The following two tydeids: *Lorryia reticulata* and *L. obliqua* were found solely on bark. Ripka and Kaźmierski (1998b) found *Mediolata mariaefrancae* André, 1977 on the bark of tree infested with scale insects.

## Materials and Methods

Between 1990 and 1998, a mite survey was made on ornamental trees and shrubs of streets, squares, parks, botanical gardens, private gardens, and green areas of housing estates in all districts of Budapest (in some other localities in Hungary and Croatia, also herbaceous plants were occasionally examined). Plant samples from 301 woody plant species (minimum 5 shoots, branches or bark/plant or 25 leaves/plant) were taken in plastic bags from January to late November. During plant examination with binocular microscope (upper and lower surfaces of the leaves, petioles, buds, bark, flowers, galls, etc.), all the mites found on the plant samples were put into AGA solution or directly into lactic acid. After the clearing of mites in the lactic acid, they were placed into Keifer's medium 2 following the Keifer's mounting method for eriophyoid mites (Keifer, 1952). The cleaned and dyed mites were then placed into Keifer's medium 3. The Hoyer's medium with sorbitol (Keifer, 1975), and Heinze's polyvinyl alcohol (PVA) medium (Schmutterer, 1959) were used alternatively for mounting the specimens. The slide preparations were dried in thermostat at 32 °C and then sealed with nail varnish. Specimens were examined with a phase contrast microscope. For mite determination the keys of Fain (1982), Bolland (1991), Smiley (1992) as well as several original species descriptions and other available reports were used.

## Results

Corticolous mites were collected on 48 woody plant species out of 301 studied belonging to 19 families. These mites were found in 90 plant samples. A total of 19 mite species were identified belonging to 14 families (see Table 1 for the exact host/locality data). The following species were new for the Hungarian fauna: *Cunaxoides kielczewskii* Michocka, *Eupalopsis maseriensis* (Canestrini et Fanzago), *Hericia hericia* Robin, *Mediolata vandergeesti* (Gomaa et Bolland), *Michaelopus corticalis* (Michael). *M. corticalis*, *Hemisarcoptes budensis* Fain et Ripka and *Cheletogenes ornatus* were the most frequent species in the mite fauna associated with scale insects. Corticolous mites occurred on 16 species of family Rosaceae, on 8 species of family Oleaceae, and on 4 species of family Salicaceae. *Ch. ornatus* was collected on coccid infested cortex, exceptionally on leaves. *H. budensis*, *C. kielczewskii*, *E. maseriensis*, *M. mariaefrancae*, *M. vandergeesti*, *Neophyllobius* sp. near *sturmerwoodi* and *Neophyllobius* spec. nov. were found on branches infested with the following scale insects: *Pseudaulacaspis pentagona* (Targioni-Tozzetti), *Unaspis euonymi* (Comstock), *Epidiaspis leperii* (Signoret), *Chionaspis salicis* (Linnaeus), *Aulacaspis rosae* (Bouché), *Diaspidiotus gigas* (Thiem et Gerneck), *Diaspidiotus ostreaeformis* (Curtis), *Diaspidiotus perniciosus* (Comstock), and *Lepidosaphes ulmi* (Linnaeus) (for more data on Coccoidea see Ripka et al., 1996). The cortex-dwelling camerobiid, cheyletid, cunaxid, eupalopsellid and stigmatid mites usually occurred in low numbers, but the coccid infestations were severe. Phoretic hypopodes (entomophilous deuteronymphs) of *H. budensis* were collected on *Chilocorus renipustulatus* (Scriba) and *Chilocorus bipustulatus* (L.) (Coleoptera: Coccinellidae).

*M. corticalis* was found under the scales of dead *P. pentagona*, *E. leperii*, *U. euonymi*, *Ch. salicis*, *Diaspidiotus marani* Zahradnik, *D. gigas*, *D. perniciosus*, *L. ulmi* and colonies of *Pseudochermes fraxini* (Kaltenbach). *H. hericia* was collected under the peeling (loose) bark of London plane trunk. Since the paper of Ripka et al. (1997) an other tarsonemid mite was found associated with coccid. *Tarsonemus bachmaieri* Cooreman, 1958 was found under the scales of *Carulaspis juniperi* (Bouché). Oribatids were found only on bark of twigs in parks, cemetery and arboreta. They could not be found on street trees. The oribatid mites do not tolerate the aerial pollution in urban habitat. The vast majority of corticolous mite species is slow-moving, exception *T. immanis*.

**Table 1**

Corticolous mite species collected from ornamental trees and shrubs  
(\* = new species for the Hungarian fauna)

Mite species	Plant species	Sampling place	Sampling date
<b>Acari: Prostigmata</b>			
fam. <b>Cheyletidae</b>			
<i>Cheletacarus raptor</i> Volgin, 1961			
	<i>Populus x canescens</i>	Óbuda	25.08. 1998
	<i>Populus simonii</i>	Vérmező	25.01. 1991
	<i>Rhamnus catharticus</i>	Józsefváros	04.01. 1995
<i>Cheletogenes ornatus</i> (Canestrini et Fanzago, 1876)			
	<i>Celastrus orbiculatus</i>	Gellérthegy	09.09. 1997
	<i>Crataegus laevigata</i>	Gellérthegy	05.02. 1991
		Pestszentlőrinc	02.09. 1992
	<i>Ilex aquifolium</i>	Városliget	04.02. 1995
	<i>Jasminum beesianum</i>	Gellérthegy	27.01. 1995
	<i>Populus x canescens</i>	Óbuda	25.08. 1998
	<i>Populus simonii</i>	Sasad	13.09. 1991
	<i>Populus tremula</i>	Újpalota	30.08. 1994
	<i>Prunus avium</i>	Kispest	02.04. 1992
	<i>Thuja occidentalis</i>	Budaörs (Pest c.)	07.03. 1995
<i>Prosocheyla traubi</i> (Baker, 1949)			
	<i>Populus simonii</i>	Sasad	13.09. 1991
fam. <b>Eupalopsellidae</b>			
* <i>Eupalopsis maseriensis</i> (Canestrini et Fanzago, 1876)			
	<i>Crataegus laevigata</i>	Gellérthegy	24.01. 1995
	<i>Crataegus monogyna</i>	Gellérthegy	26.01. 1995
	<i>Malus pumila</i> var. <i>niedzwetzkyana</i>	Gellérthegy	26.01. 1995
fam. <b>Stigmaeidae</b>			
<i>Mediolata mariae-francae</i> André, 1977			
	<i>Euonymus europaeus</i>	Gellérthegy	15.06. 1997
	<i>Jasminum beesianum</i>	Gellérthegy	27.01. 1995
	<i>Rhamnus catharticus</i>	Józsefváros	04.01. 1995
	<i>Tilia x euchlora</i>	Józsefváros	29.06. 1994
* <i>Mediolata vandergeesti</i> (Gomaa et Bolland, 1982)			
	<i>Crataegus monogyna</i>	Gellérthegy	26.01. 1995
	<i>Ribes uva-crispa</i>	Széchenyi-hegy	04.02. 1991
fam. <b>Camerobiidae</b>			
* <i>Neophyllobius</i> sp. near <i>sturmerwoodi</i> Bolland, 1991			
	<i>Juglans nigra</i>	Kőbánya	29.03. 1995
* <i>Neophyllobius</i> sp. nov.			
	<i>Euonymus europaeus</i>	Gellérthegy	15.06. 1997
	<i>Euonymus europaeus</i>	Gellérthegy	09.09. 1997

Table 1 (cont.)

Mite species	Plant species	Sampling place	Sampling date
<b>fam. Cunaxidae</b>			
* <i>Cunaxoides kielczewskii</i> Michocka, 1982			
	<i>Picea pungens</i>	Orosháza (Békés c.)	07.07. 1998
	<i>Rhus typhina</i>	Kőbánya	04.01. 1995
	<i>Taxus baccata</i>	Orbán-hegy	25.08. 1994
<b>fam. Tarsonemidae</b>			
<i>Tarsonemus bachmaieri</i> Cooreman, 1958	<i>Thuja</i> sp.	Józsefváros	28.10. 1991
<i>Tarsonemus nodosus</i> Schaarschmidt, 1959	<i>Celastrus orbiculatus</i>	Gellérthegey	09.04. 1997
	<i>Euonymus europaeus</i>	Rákoskeresztúr	17.08. 1997
		Rákoskeresztúr	17.10. 1997
<i>Tarsonemus waitei</i> Banks, 1912	<i>Euonymus europaeus</i>	Rákoskeresztúr	17.08. 1997
		Rákoskeresztúr	17.10. 1997
<b>Acari: Astigmata</b>			
<b>fam. Hemisarcoptidae</b>			
<i>Hemisarcoptes budensis</i> Fain et Ripka, 1998			
	<i>Celastrus orbiculatus</i>	Gellérthegey	09.09. 1997
	<i>Crataegus laevigata</i>	Gellérthegey	24.01. 1995
	<i>Euonymus europaeus</i>	Gellérthegey	27.01. 1995
		Gellérthegey	15.06. 1997
	<i>Fraxinus angustifolia</i>	Víziváros	08.02. 1991
		Óbuda	11.09. 1991
	<i>Fraxinus pennsylvanica</i>	Víziváros	08.02. 1991
	<i>Malus baccata</i>	Gellérthegey	24.01. 1995
	<i>Phellodendron amurense</i>	Margitsziget	21.02. 1991
	<i>Populus x canescens</i>	Óbuda	03.04. 1991
		Óbuda	25.08.1998
	<i>Populus simonii</i>	Lágymányos	05.02. 1991
		Sasad	13.09. 1991
	<i>Prunus domestica</i>	Újpalota	12.02. 1991
	<i>Pyrus betulifolia</i>	Gellérthegey	18.01. 1995
	<i>Pyrus pyraister</i>	Józsefváros	04.01. 1995
	<i>Rhus typhina</i>	Kőbánya	04.01. 1995
	<i>Sophora japonica</i>	Víziváros	11.02. 1991
<b>fam. Acaridae</b>			
* <i>Michaelopus corticalis</i> (Michael, 1885)			
	<i>Acer platanoides</i>	Orosháza (Békés c.)	07.07. 1998
	<i>Acer pseudoplatanus</i>	Csongrád (Csongrád c.)	26.05. 1998
	<i>Celastrus orbiculatus</i>	Gellérthegey	07.08. 1998
	<i>Celastrus scandens</i>	Józsefváros	04.01. 1995
	<i>Cornus alba</i>	Órmező	28.03. 1995

Table 1 (cont.)

Mite species	Plant species	Sampling place	Sampling date
	<i>Cornus stolonifera</i>	Örmező	10.02. 1995
	<i>Crataegus laevigata</i>	Gellérthegey	24.01. 1995
	<i>Crataegus monogyna</i>	Gellérthegey	26.01. 1995
	<i>Crataegus succulenta</i>	Gellérthegey	24.01. 1995
	<i>Daphne mezereum</i>	Gellérthegey	24.01. 1995
	<i>Euonymus europaeus</i>	Gellérthegey	27.01. 1995
		Gellérthegey	09.09. 1997
	<i>Forsythia suspensa</i>	Pasarét	17.09. 1990
		Belváros	10.04. 1993
	<i>Fraxinus excelsior</i>	Pasarét	17.05. 1991
	<i>Ilex aquifolium</i>	Városliget	04.02. 1995
	<i>Jasminum beesianum</i>	Gellérthegey	27.01. 1995
	<i>Ligustrum vulgare</i>	Tabán	08.05. 1991
	<i>Malus baccata</i>	Gellérthegey	24.01. 1995
	<i>Malus halliana</i>	Tabán	05.05. 1992
	<i>Malus x purpurea</i>	Vérmező	05.05. 1992
	<i>Phellodendron amurense</i>	Margitsziget	21.02. 1991
	<i>Populus alba</i>	Budafok	07.02. 1995
		Örmező	12.05. 1995
	<i>Populus x canescens</i>	Óbuda	25.08. 1998
	<i>Populus simonii</i>	Vérmező	25.01. 1991
		Sasad	23.09. 1991
	<i>Prunus cerasifera</i>	Tabán	07.02. 1991
		Gellérthegey	08.05. 1991
	<i>Prunus fruticosus</i>	Zugló	18.03. 1995
	<i>Prunus spinosa</i>	Vérmező	07.03. 1993
	<i>Pyrus betulifolia</i>	Gellérthegey	18.01. 1995
	<i>Pyrus pashia</i>	Gellérthegey	18.01. 1995
	<i>Pyrus pyraeaster</i>	Józsefváros	04.01. 1995
	<i>Rhus typhina</i>	Kőbánya	04.01. 1995
	<i>Syringa vulgaris</i>	Tabán	12.02. 1991
fam. <b>Glycyphagidae</b>			
* <i>Hericia hericia</i> Robin, 1868			
	<i>Platanus x hispanica</i>	Józsefváros	16.05. 1991
<b>Acari: Oribatida</b>			
fam. <b>Micreremidae</b>			
<i>Micreremus brevipes</i> (Michael, 1888)			
	<i>Crataegus monogyna</i>	Gellérthegey	26.01. 1995
	<i>Daphne mezereum</i>	Gellérthegey	24.01. 1995
	<i>Malus spectabilis</i>	Törökvész	03.04. 1992
	<i>Prunus spinosa</i>	Vérmező	07.03. 1993
	<i>Quercus ilex</i>	Lanterna (Croatia)	05.07. 1996
fam. <b>Oribatulidae</b>			
<i>Phauloppia conformis</i> (Berlese, 1895)			
	<i>Juglans nigra</i>	Kőbánya	29.03. 1995

Table 1 (cont.)

Mite species	Plant species	Sampling place	Sampling date
<b>fam. Cymbaeremaeidae</b>			
<i>Scapheremaeus reticulatus</i> (Berlese, 1910)			
	<i>Ilex aquifolium</i>	Városliget	04.02. 1995
	<i>Malus pumila</i> var. <i>niedzwetzkyana</i>	Gellérthegey	26.01. 1995
<b>fam. Ceratozetidae</b>			
<i>Trichoribates trimaculatus</i> (C. L. Koch, 1836)			
	<i>Jasminum beesianum</i>	Gellérthegey	27.01. 1995
<b>fam. Scheloribatidae</b>			
<i>Scheloribates latipes</i> (C. L. Koch, 1844)			
	<i>Jasminum beesianum</i>	Gellérthegey	27.01. 1995

## Discussion

The corticolous fauna of species Acari has not been so far investigated extensively in Hungary.

The studies of Lebrun (1976), André (1986), also Momen and Lundqvist (1996) testify that the expected mite species living in a relatively small area can appear surprisingly rich.

The bark and bark crevices are the most important overwintering sites of foliage-dwelling mites. Some of them viz. tetranychid, tenuipalpid, phytoseiid, stigmatid, tydeid, winterschmidtii species overwinter under the shield of dead coccids (Ripka, 1998a, b, Ripka and Kaźmierski, 1998a, b). There is a closer association between some members of family Tarsonemidae and coccids. *Tarsonemus nodosus*, *T. lobosus*, *T. karli*, *T. waitei* and *Dendroptus willmanni* also an other *Dendroptus* sp. quite often were found together with scale insects (Ripka et al., 1997). Fungivorous tarsonemid mites prefer this microhabitat. They feed on microscopic fungi.

Some tydeid species gave definite preference to cortex.

The predaceous members of corticolous cameroibiid, cheyletid, cunaxid, eupalopsellid, hemisarcoptid, stigmatid and tydeid mite fauna cannot play a great role in regulating numbers of phytophagous insects and mites because of their low densities. But the knowledge on the predatory activities of these mites is sparse and mostly brief.

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