

CURRENT RESEARCH ON PHYTOPATHOGENIC FUNGI: AN OVERVIEW

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The majority of plant pathogens comprising about 60% of the total are belonging to a group of eukaryotic microorganisms, commonly known as FUNGI although this name obviously covers a number of organisms that are not fungi in a strict sense [1]. Yet, all these plant pathogens, having different systematic positions within the livings, are of significance as far as agricultural crop production is concerned.

Mycologists in the past were primarily interested in identifying fungal diseases, describing the fungi, studying their life cycle in relation to environment and looking for effective control measures. A big step forward was when investigations expanded toward the physiology of plant diseases and disease resistance. Meanwhile fungal genetics has developed providing a better understanding of plant-fungus relations. Recently, with the introduction of molecular techniques, new approaches of research, such as molecular taxonomy and molecular genetics have been established and the molecular methods were applied in other related fields of studies.

For the illustration of changes in research interest worldwide, the main research topics and all the contributions (poster presentations) accepted at and published by the 7th International Congress of Plant Pathology held in Edinburgh, Scotland between 9-16 August 1998 have been scanned. Based on these, some of the most promising research trends with a few examples will be accounted here and then a short overview of what has recently been done by Hungarian mycologists on the subject will be given.

Research tendencies in the World

Detection and identification. No doubt, modern diagnostic technology started in 1976 with the first application of ELISA to plant virus detection. Subsequently, assays have been targeted to the detection of plant pathogenic fungi. ELISA kits are now commercially available and ELISA and other related antibody-based technologies are widely used. As an example, Jennings et al. [2] have raised Mabs (monoclonal antibodies) to either the *Fusarium* genus or individual species, such as *F. avenaceum*, *F. culmorum* and *F. poae*. Further, the use of protein and enzyme profiles have also been established as

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tools of differentiating between fungal species/strains. Belisario et al. [3] found *Monilinia fructigena*, *M. fructicola* and *M. laxa*, the brown rot pathogen of fruits distinguishable based on protein patterns.

The practical use of nucleic acid-based detection technologies in plant pathology was limited until the introduction of the polymerase chain reaction (PCR) assay in the late 1980s. Afterwards, however, PCR or reverse transcription (RT) PCR have been developed for a wide range of plant pathogens including phytopathogenic fungi, and the combination of PCR and DNA hybridization allowed the detection of multiple pathogens in a single test. The only limiting factor of widespread application of these techniques is their expensiveness. One of the most promising uses of DNA technologies in the plant pathological practise is the detection of latent (symptomless) infection by fungal pathogens in plants. For example, Whisson et al. [4] were able to produce taxon-specific DNA probes derived from DNA extracted from both *Phomopsis viticola* conidia and infected grapevine tissues. A PCR-based assay then allowed the detection of the fungus from symptomless plants.

Apart from molecular methods, there are improved microscopical techniques, like image analyses or comparative electron microscopy (SEM and TEM) of spores, suitable for discriminating between uncertain species [5, 6].

Genetic and molecular aspects of pathogenicity. Within the last few years, the application of molecular genetic methods has enabled the identification of a variety of genes involved in pathogenesis, such as determinants of early infection stages, colonization and of fungal products killing plant cells. One of the most exciting questions is what are the signals influencing spore germination and host penetration, in other words, what are the factors responsible for mutual recognition and subsequent events leading to disease. With the rice blast fungus, *Magnaporthe grisea*, for example, thigmotrophic sensing appeared to involve a fungal cell-surface protein called Mpg-1 that belongs to the known hydrophobins [7]. Another point of interest relates to the presence of specific proteins of fungal origin, called elicitors, the recognition of which by the host plant is a prerequisite of events leading to either compatibility or incompatibility. By means of molecular methods it was possible to clone elicitor genes from the fungus *Phytophthora sojae* [8].

Necrotrophic fungi require a rapid death of plant cells to get into contact with host nutrients. Therefore, a wide range of secondary metabolites, like phytotoxins and lytic enzymes produced by them prior to, during and after penetration are considered to play a role in pathogenesis. In fact, by isolating and cloning the genes governing the production of pectinases and lipases, Ten Have et al. [9] elucidated their significance.

Since P J G M de Wit and his co-workers [10] were successful in cloning first an avirulence gene from the fungus *Cladosporium fulvum*, additional avirulence genes have been identified in different laboratories, e.g. in Australia [11] and in the UK [12].

Genetic diversity of fungal populations. The mechanisms of population genetics over space and time appear to be rather complex and multifactorial. However, there are increasing experimental evidences that genetic structures of phytopathogenic fungi are basically influenced (determined) by mating systems, immigration (gene flow) and selection [13] as well as by somatic heterokaryosis [14].

From the practical point of view, population changes as variation in virulence, aggressiveness or fungicide-sensitivity are being of particular interest. Accordingly, the number of investigations using either conventional methods (host differentials, mating analysis), molecular techniques (DNA polymorphism) or both have markedly been increased. Recently, Pipe et al. [15] applied, as a new tool, microsatellite markers with *Phytophthora infestans* for the analysis of diversity in population structure.

Response to environment. Ecology of phytopathogenic fungi came to the focus recently in many mycological laboratories. Of particular interest are investigations looking at the survival and growth of fungi in soils of various qualities, as well as the various biotic interactions including those between pathogenic and non-pathogenic species [16]. To study the complexity of such soil systems and to elucidate the processes taking place in the rhizosphere, molecular approaches, e.g. the use of GUS-transformants of *Trichoderma harzianum* have been applied [17]. Since many of the non-plant pathogenic soil microorganisms are considered as potential biocontrol agents, techniques of isolation and characterization and mass-production of the candidates have been the subject of studies worldwide. In addition, as a most promising tool, the so-called molecular breeding of some of the biological antagonists were initiated [18].

Research progress in Hungary

Much of our studies on phytopathogenic fungi, being either basic or applied, have been carried out in research institutes (MTA-NKI, MBK, MTA-MgKI) and agricultural universities (GATE, PATE, DATE, KÉE, SE), respectively. Based on recent publications, a selection of research topics will be shown here to provide with a short and informative overview.

Etiology. Of particular interest are studies related to woody plant diseases (fruit and forest trees, grapevine) where newly appearing pathogens or pathogen complexes are causing severe damage [19, 20], or investigations of the pathogenic mycofloras of natural vs. agricultural ecosystems [21].

Detection/identification. Besides classical methods, molecular techniques in the detection and identification of phytopathogenic fungi have been introduced. For example, the comparison of pectic enzyme zymograms of *Cytospora* species [22], and the esterase isozyme patterns of different *Fusarium* species [23] permitted to distinguish between different taxons, or DNA polymorphism and RAPD analysis verified the taxonomic position of genera and species of the so-called "Helminthosporium" form complex [24].

Population structure and genetics. Much of recent studies have been focussing on inter- and intraspecific variations of some important phytopathogenic fungi and both classical methods and new molecular techniques have been applied. For example, the relative dominance of *Fusarium* species in winter wheat and maize [25, 26], changes in mating type, virulence pattern and fungicide sensitivity of *Phytophthora infestans* [27], evolutionary processes in the virulence character of field populations of *Blumeria graminis*, *Puccinia graminis* and *Plasmopara halstedii* [28–30] have been the subject of investigations in order to combat with these pathogens. Furthermore, PCR-RAPD

techniques allowed to distinguish between *Fusarium* species belonging to the teleomorph *Gibberella fujikuroi* [31], and cloning a repetitive element from *Fusarium poae* made it possible to understand, at least in part, the genetic background of fungus variability [32].

Biological antagonists. The potential of using antagonistic fungi as biocontrol agents in agriculture has prompted Hungarian mycologists to set up and expand research in different laboratories. A wide range of investigations, from the isolation, identification and systematics of such fungi through molecular genetics to the patented bioproduct have been made e.g. with *Trichoderma* spp. [33], *Coniothyrium minitans* and *Ampelomyces* spp [34]. In addition, VCG groups within the Hungarian populations of *Cryphonectria parasitica* have been determined enhancing to characterize population structures and to detect hypovirulence, a phenomenon known to play an important role in the biocontrol process against this pathogen [35]. And another, special aspect of studies on antagonistic fungi resulted in identifying species as potential mycoherbicides [36].

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