The Beaker Phenomenon and the Genomic **Transformation of Northwest Europe**

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Bell Beaker pottery spread across western and central Europe beginning around 2750 BCE before disappearing between 2200-1800 BCE. The mechanism of its expansion is a topic of long-standing debate, with support for both cultural diffusion and human migration. We present new genome-wide ancient DNA data from 170 Neolithic, Copper Age and Bronze Age Europeans, including 100 Beaker-associated individuals. In contrast to the Corded Ware Complex, which has previously been identified as arriving in central Europe following migration from the east, we observe limited genetic affinity between Iberian and central European Beaker Complex-associated individuals, and thus exclude migration as a significant mechanism of spread between these two regions. However, human migration did have an important role in the further dissemination of the Beaker Complex, which we document most clearly in Britain using data from 80 newly reported individuals dating to 3900-1200 BCE. British Neolithic farmers were genetically similar to contemporary populations in continental Europe and in particular to Neolithic Iberians, suggesting that a portion of the farmer ancestry in Britain came from the Mediterranean rather than the Danubian route of farming expansion. Beginning with the Beaker period, and continuing through the Bronze Age, all British individuals harboured high proportions of Steppe ancestry and were genetically closely related to Beaker-associated individuals from the Lower Rhine area. We use these observations to show that the spread of the Beaker Complex to Britain was mediated by migration from the continent that replaced >90% of Britain's Neolithic gene pool within a few hundred years, continuing the process that brought Steppe ancestry into central and northern Europe 400 years earlier. During the third millennium Before the Common Era (BCE), two new archaeological pottery styles expanded across Europe, replacing many of the more localized styles that preceded them¹. The "Corded Ware Complex" in central, northern and eastern Europe was associated with people who derived most of their ancestry from eastern European Yamnaya steppe pastoralists² ⁴. Bell Beaker pottery is known from around 2750 cal BCE^{5,6} in Atlantic Iberia, although its exact origin is still a matter of debate^{7,8}. By 2500 BCE, it is possible to distinguish in many regions the "Beaker Complex", defined by assemblages of grave goods including stylised bellshaped pots, distinctive copper daggers, arrowheads, stone wristguards and V-perforated buttons⁹. Regardless of the geographic region where it originated (if it did have a single origin), elements of the Beaker Complex rapidly spread throughout western Europe (and northern Africa), reaching southern and Atlantic France, Italy and central Europe 10-12 where they overlapped geographically with the Corded Ware Complex, and from there expanding to Britain and Ireland^{13,14}. A major debate has centred on whether the spread of the Beaker Complex was mediated by the movement of people, culture, or a combination of these 15-18. Genome-wide data have revealed high proportions of Steppe ancestry in Beaker Complex-associated individuals

- from Germany and the Czech Republic²⁻⁴, consistent with their being a mixture of populations
- from the Steppe and the preceding farmers of Europe. However, a deeper understanding of the
- ancestry of people associated with the Beaker Complex requires genomic characterization of
- individuals across the geographic range and temporal duration of this archaeological
- phenomenon.

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Ancient DNA data and authenticity

- To understand the genetic structure of ancient people associated with the Beaker Complex and
- their relationship to preceding, subsequent and contemporary peoples, we enriched ancient
- DNA libraries for sequences overlapping 1,233,013 single nucleotide polymorphisms (SNPs) by
- hybridization DNA capture^{4,19}, and generated new sequence data from 170 ancient Europeans
- dating to ~4700–1200 BCE (Supplementary Table 1; Supplementary Information, section 1).
- We also generated 62 new direct radiocarbon dates (Extended Data Table 1). We filtered out
- libraries with low coverage (<10,000 SNPs) or evidence of contamination (Methods) to obtain a
- 148 final set of 166 individuals: 97 Beaker-associated individuals and 69 from other ancient
- populations (Fig. 1b; Extended Data Table 2), including 61 individuals from Neolithic and
- Bronze Age Britain. We combined our data with previously published ancient DNA data^{2-4,20-37}
- to form a genome-wide dataset of 476 ancient individuals (Supplementary Table 1). The
- 152 combined dataset included Beaker-associated individuals from Iberia (n=20), southern France
- 153 (n=4), northern Italy (n=1), central Europe (n=56), The Netherlands (n=9) and Britain (n=19).
- We further merged these data with 2,572 present-day individuals genotyped on the Affymetrix
- Human Origins array^{22,31} and 300 high coverage genomes sequenced as part of the Simons
- 156 Genome Diversity Project³⁸.

Y-chromosome analysis

- We determined Y-chromosome haplogroups for the 54 male Beaker-associated individuals
- 159 (Supplementary Table 3). Individuals from the Iberian Peninsula carried Y haplogroups known
- to be common across Europe during the earlier Neolithic period^{2,4,20,26,32,39}, such as I2a (n=3) and
- 161 G2 (n=1) (Supplementary Table 3). In contrast, Beaker-associated individuals outside Iberia
- 162 (n=44) largely carried R1b lineages (84%), associated with the arrival of Steppe migrants in
- 163 central Europe during the Late Neolithic/Early Bronze Age^{2,3}. For individuals in whom we
- 164 could determine the R1b subtype (n=22), we found that all but one had the derived allele for the
- R1b-S116/P312 polymorphism, which defines the dominant subtype in western Europe today⁴⁰.
- Finding this early predominance of the R1b-S116/P312 polymorphism in ancient individuals
- from central and northwestern Europe suggests that people associated with the Beaker Complex
- 168 may have had an important role in the dissemination of this lineage throughout most of its
- present-day distribution.

Genomic insights into the spread of the Beaker Complex

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Principal component analysis (PCA) revealed striking heterogeneity among individuals assigned to the Beaker Complex (Fig. 1c, Extended Data Fig. 1a). Genetic differentiation in our dataset was mainly driven by variable amounts of Steppe-related ancestry, with Beaker Complex individuals falling along the axis of variation defined by Yamnaya steppe pastoralists and Middle Neolithic/Copper Age European populations. We obtained qualitatively consistent inferences using ADMIXTURE model-based clustering⁴¹ (Extended Data Fig. 1b). We grouped Beaker Complex individuals based on geographic proximity and genetic similarity (Supplementary Information, section 4), and used $qpAdm^2$ to model their ancestry as a mixture of western European hunter-gatherers (WHG), northwestern Anatolian farmers, and Yamnaya steppe pastoralists (the first two of which contributed to earlier European farmers; Supplementary Information, section 6). We find that the great majority of Beaker Complex individuals outside of Iberia derive a large portion of their ancestry from Steppe populations (Fig. 2a), whereas in Iberia, such ancestry is absent in all sampled individuals, with the exception of two (I0461 and I0462) from the Arroyal I site in northern Spain. We detect striking differences in ancestry not only at a pan-European scale, but also within regions and even within sites. Unlike other individuals from the Upper Alsace region of France (n=2), an individual from Hégenheim resembles previous Neolithic populations and can be modelled as a mixture of Anatolian Neolithic and western hunter-gatherers without any Steppe-related ancestry. Given that the radiocarbon date of the Hégenheim individual is older (2832-2476 cal BCE (quoting 95.4% confidence intervals for this and other dates) (Supplementary Information, section 1) than other samples from the same region (2566-2133 cal BCE), the pattern could reflect temporal differentiation. At Szigetszentmiklós in Hungary, we find Beaker Complexassociated individuals with very different proportions (from 0% to 74%) of Steppe ancestry but overlapping dates. This genetic heterogeneity is consistent with early stages of mixture between previously established European farmers and migrants with Steppe ancestry. An implication is that, even at a local scale, the Beaker Complex was associated with people of diverse ancestries. While the Yamnaya-related ancestry in Beaker Complex associated individuals had an origin in the Steppe^{2,3}, the other ancestry component (from European Neolithic farmers) could potentially be derived from several parts of Europe, as genetically closely related populations were widely distributed across the continent during the Neolithic and Copper Age periods^{2,4,22,25,26,28,32}. To obtain insight into the origin of the Neolithic-related ancestry in Beaker Complex-associated individuals, we began by looking for regional patterns of genetic differentiation within Europe during the Neolithic and Copper Age periods. To study genetic affinity to different Early Neolithic (EN) populations, we computed f_4 -statistics of the form $f_4(Outgroup, Test; Iberia EN,$

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LBK EN) for Neolithic and Copper Age test populations predating the emergence of the Beaker Complex. As previously described², there is genetic affinity to Iberian Early Neolithic farmers in Iberian Middle Neolithic/Copper Age populations, but not in central and northern European Neolithic populations (Fig. 2b), which could be explained by differential affinities to huntergatherer individuals from different regions⁴² (Extended Data Fig. 2). A new finding that emerges from our analysis is that Neolithic individuals from southern France and Britain also show a greater affinity to Iberian Early Neolithic farmers than to central European Early Neolithic farmers (Fig. 2b), similar to previous results obtained in a Neolithic farmer genome from Ireland²⁸. By modelling Neolithic populations and WHG in an admixture graph framework, we replicate these results and further show that they are not driven by different proportions of hunter-gatherer admixture (Extended Data Fig. 3; Supplementary Information, section 5). Our results suggest that a portion of the ancestry of the Neolithic farmers of Britain was derived from migrants who spread along the Atlantic coast. Megalithic tombs document substantial interaction along the Atlantic façade of Europe, and our results are consistent with such interactions reflecting movements of people. More data from southern Britain (where our sampling is sparse) and nearby regions in continental Europe will be needed to fully understand the complex interactions between Britain and the continent in the Neolithic⁴³.

The distinctive genetic signatures of pre-Beaker Complex populations in Iberia compared to central Europe allow us to test formally for the origin of the Neolithic farmer-related ancestry in Beaker Complex individuals in our dataset (Supplementary Information, section 6). We grouped individuals from Iberia (n=19) and from outside Iberia (n=84) to increase power, and evaluated the fit of different Neolithic/Copper Age groups with *qpAdm* under the model: Yamnaya + Neolithic/Copper Age. For Beaker Complex individuals from Iberia, the best fit was obtained when Middle Neolithic and Copper Age populations from the same region were used as a source for their Neolithic farmer-related ancestry, and we could exclude central and northern European populations (P < 4.69E-03) (Fig. 2c). Conversely, the Neolithic farmer-related ancestry in Beaker Complex individuals outside Iberia was most closely related to central and northern European Neolithic populations with relatively high hunter-gatherer admixture (e.g. *Globular_Amphora_LN*, P = 0.14; TRB_Sweden_MN, P = 0.29), and we could significantly exclude Iberian sources (P < 3.18E-08) (Fig. 2c). These results support largely different origins

Nearly complete turnover of ancestry in Britain

237 British Beaker Complex individuals (n=19) show strong similarities to the central European

for Beaker Complex individuals, with no discernible Iberia-related ancestry outside Iberia.

- Beaker Complex both in genetic profile (Extended Data Fig. 1) and in material culture: the great
- 239 majority of individuals from both regions are associated with "All Over Corded" Beaker

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pottery. The presence of large amounts of Steppe-related ancestry in the British Beaker Complex (Fig. 2a) contrasts sharply with Neolithic individuals from Britain (n=35), who have no evidence of Steppe genetic affinities and cluster instead with Middle Neolithic and Copper-Age populations from mainland Europe (Extended Data Fig. 1). Thus, the arrival of Steppe ancestry in Britain was mediated by a migration that began with the Beaker Complex. A previous study showed that Steppe ancestry arrived in Ireland by the Bronze Age²⁸, and here we show that – at least in Britain – it arrived by the Copper Age / Beaker period. Among the different continental Beaker Complex groups analysed in our dataset, individuals from Oostwoud (Province of Noord-Holland, The Netherlands) are the most closely related to the great majority of the Beaker Complex individuals from southern Britain (n=14). They had almost identical Steppe ancestry proportions (Fig. 2a), the highest shared genetic drift (Extended Data Fig. 4b) and were symmetrically related to other ancient populations using f₄statistics (Extended Data Fig. 4a), showing that they are consistent with being derived from the same ancestral population without additional mixture into either group. We next investigated the magnitude of population replacement in Britain with qpAdm² by modelling Beaker Complex and Bronze Age individuals as a mixture of continental Beaker Complex (using the Oostwoud individuals as a surrogate) and the British Neolithic population (Supplementary Information, section 6). Fig. 3a shows the results of this analysis, ordering individuals by date and showing excess Neolithic ancestry compared to continental Beaker Complex as a baseline. For the earliest individuals (between ~2400-2000 BCE), the Neolithic ancestry excess is highly variable, consistent with migrant communities who were just beginning to mix with the previously established Neolithic population of Britain. During the subsequent Bronze Age we observe less variation among individuals and a modest increase in Neolithic-related ancestry (Fig. 3a), which could represent admixture with persisting populations with high levels of Neolithic-related ancestry (or alternatively incoming continental populations with higher proportions of Neolithic-related ancestry). In either case, our results imply a minimum of 93±2% local population turnover by the Middle Bronze Age (Supplementary Information, section 6). Specifically, for individuals from Britain around 2000 BCE, at least this fraction of their DNA derives from ancestors who at 2500 BCE lived in continental Europe. An independent line of evidence for population turnover comes from Y-chromosome haplogroup composition: while R1b haplogroups were completely absent in the Neolithic samples (n=25), they represent 95% and 75% of the Y-chromosomes in Beaker Complex-Early Bronze Age and Middle Bronze Age males in Britain, respectively (Fig. 3b; Supplementary Table 3). Our genetic time transect in Britain also allowed us to track the frequencies of alleles with known phenotypic effects. Derived alleles at rs12913832 (SLC45A2) and rs16891982 (HERC2/OCA2), which contribute to reduced skin and eye pigmentation in Europeans,

- dramatically increased in frequency during the Beaker and Bronze Age periods (Extended Data
- Fig. 5). Thus, the arrival of migrants associated with the Beaker Complex significantly altered
- the pigmentation phenotypes of British populations. However, the lactase persistence allele at
- SNP rs4988235 remained at very low frequencies in our dataset both in Britain and continental
- 280 Europe, showing that the major increase in its frequency in Britain, as in mainland Europe,
- occurred in the last 3,500 years^{3,4,39,44}.

Discussion

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- 283 The term "Bell Beaker" was introduced by late 19th-century and early 20th-century
- archaeologists to refer to the distinctive pottery style found across western and central Europe at
- 285 the end of the Neolithic, initially hypothesized to have been spread by a genetically
- homogeneous group of people. This idea of a "Beaker Folk" became unpopular after the 1960s
- as scepticism about the role of migration in mediating change in archaeological cultures grew⁴⁵,
- 288 although J.G.D. Clark speculated that the Beaker Complex expansion into Britain was an
- exception 46, a prediction that has now been borne out by ancient genomic data.
- Our results clearly prove that the expansion of the Beaker Complex cannot be described by a
- simple one-to-one mapping of an archaeologically defined material culture to a genetically
- 292 homogeneous population. This stands in contrast to other archaeological complexes analysed to
- 293 date, notably the *Linearbandkeramik* first farmers of central Europe², the Yamnaya of the
- 294 Pontic-Caspian Steppe^{2,3}, and to some extent the Corded Ware Complex of central and eastern
- Europe^{2,3}. Instead, or results support a model in which both cultural transmission and human
- 296 migration played important roles, with the relative balance of these two processes depending on
- 297 the region. In Iberia, the majority of Beaker Complex-associated individuals lacked Steppe
- affinities and were genetically most similar to preceding Iberian populations. In central Europe,
- 299 Steppe ancestry was widespread and we can exclude a substantial contribution from Iberian
- 300 Beaker Complex-associated individuals, contradicting initial suggestions of gene flow between
- these groups based on analysis of mtDNA⁴⁷ and dental morphology⁴⁸. Small-scale contacts
- 302 remain plausible, however, as we observe small proportions of Steppe ancestry in two
- individuals from northern Spain.
- Although cultural transmission seems to have been the main mechanism for the diffusion of the
- 305 Beaker Complex between Iberia and central Europe, other parts of the Beaker Complex
- 306 expansion were driven to a substantial extent by migration, with Beaker-associated burials in
- 307 southern France, northern Italy, and Britain, representing the earliest occurrence of Steppe-
- 308 related ancestry so far known in all three regions. This genomic transformation is clearest in
- 309 Britain due to our dense genetic time transect. The earliest Beaker pots found in Britain show
- influences from both the lower Rhine region and the Atlantic façade of western Europe⁴⁹.

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However, such dual influence is not mirrored in the genetic data, as the British Beaker Complex individuals were genetically most similar to lower Rhine individuals from the Netherlands. The arrival of the Beaker Complex precipitated a profound demographic transformation in Britain, exemplified by the absence of individuals in our dataset without large amounts of Steppe-related ancestry after 2400 BCE. It is possible that the uneven geographic distribution of our samples, coupled with different burial practises between local and incoming populations (cremation versus burial) during the early stages of interaction could result in a sampling bias against local individuals. However, the signal observed during the Beaker period persisted through the later Bronze Age, without any evidence of genetically Neolithic-like individuals among the 27 Bronze Age individuals we newly report, who traced more than 90% of their ancestry to individuals of the central European Beaker Complex. Thus, the genetic evidence points to a substantial amount of migration into Britain from the European mainland beginning around 2400 BCE. These results are notable in light of strontium and oxygen isotope analyses of British skeletons from the Beaker and Bronze Age periods⁵⁰, which have provided no evidence of substantial mobility over individuals' lifetimes from locations with cooler climates or from places with geologies atypical of Britain. However, the isotope data are only sensitive to firstgeneration migrants, and do not rule out movements from regions such as the lower Rhine, which is consistent with the genetic data, or from other geologically similar regions for which DNA sampling is still sparse. Further sampling of regions on the European continent may reveal additional candidate sources. By analysing DNA data from ancient individuals we have been able to provide important constraints on the processes underlying cultural and social changes in Europe during the third millennium BCE. Our results raise new questions and motivate further archaeological research to identify the changes in social organization, technology, subsistence, climate, population sizes⁵¹ or pathogen exposure^{52,53} that could have precipitated the demographic changes uncovered in this study.

Methods

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Ancient DNA analysis

We screened skeletal samples for DNA preservation in dedicated clean rooms. We extracted DNA⁵⁴⁻⁵⁶ and prepared barcoded next generation sequencing libraries, the majority of which were treated with uracil-DNA glycosylase to greatly reduce the damage (except at the terminal nucleotide) that is characteristic of ancient DNA^{57,58} (Supplementary Information, section 2). We initially enriched libraries for sequences overlapping the mitochondrial genome⁵⁹ and ~3000 nuclear SNPs using synthesized baits (CustomArray Inc.) that we PCR amplified. We sequenced the enriched material on an Illumina NextSeq instrument with 2x76 cycles, and 2x7 cycles to read out the two indices⁶⁰. We merged read pairs with the expected barcodes that overlapped by at least 15 base pairs, mapped the merged sequences to hg19 and to the reconstructed mitochondrial DNA consensus sequence⁶¹ using the samse command in bwa (v0.6.1)⁶², and removed duplicated sequences. We evaluated DNA authenticity by estimating the rate of mismatching to the consensus mitochondrial sequence⁶³, and also requiring that the rate of damage at the terminal nucleotide was at least 3% for UDG-treated libraries⁶³ and 10% for non-UDG-treated libraries⁶⁴. For libraries that were promising after screening, we enriched in two consecutive rounds for sequences overlapping 1,233.013 SNPs ('1240k SNP capture')^{2,19} and sequenced 2x76 cycles and 2x7cycles on an Illumina NextSeq500 instrument. We processed the data bioinformatically as for the mitochondrial capture data, this time mapping only to the human reference genome hg19 and merging the data from different libraries of the same individual. We further evaluated authenticity by studying the ratio of X-to-Y chromosome reads and estimating X-chromosome contamination in males based on the rate of heterozygosity⁶⁵. Samples with evidence of contamination were either filtered out or restricted to sequences with terminal cytosine deamination to remove sequences that could have derived from modern contaminants. Finally, we filtered out from our analysis dataset samples with fewer than 10,000 targeted SNPs covered

Mitochondrial haplogroup determination

We used the mitochondrial capture bam files to determine the mitochondrial haplogroup of each sample with new data, restricting to reads with MAPQ \geq 30 and base quality \geq 30. First, we constructed a consensus sequence with samtools and beftools⁶⁶, using a majority rule and requiring a minimum coverage of 2. We called haplogroups with HaploGrep2⁶⁷ based on

at least once and samples that were first-degree relatives of others in the dataset (keeping the

sample with the larger number of covered SNPs) (Supplementary Table 1).

- 370 phylotree⁶⁸ (mtDNA tree Build 17 (18 Feb 2016)). Mutational differences compared to the
- 371 rCRS and corresponding haplogroups can be viewed in Supplementary Table 2.

Y-chromosome analysis

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- We determined Y-chromosome haplogroups for both new and published samples
- 375 (Supplementary Information, section 3). We made use of the sequences mapping to 1240k Y-
- 376 chromosome targets, restricting to sequences with mapping quality ≥ 30 and bases with quality
- 377 ≥30. We called haplogroups by determining the most derived mutation for each sample, using
- 378 the nomenclature of the International Society of Genetic Genealogy (http://www.isogg.org)
- version 11.110 (21 April 2016). Haplogroups and their supporting derived mutations can be
- viewed in Supplementary Table 3.

Merging newly generated data with published data

- We assembled two datasets for population genetics analyses:
- 385 HO includes 2,572 present-day individuals from worldwide populations genotyped on the
- Human Origins Array^{22,31,69} and 470 ancient individuals. The ancient set includes 103 Beaker
- Complex individuals (87 newly reported, 5 with shotgun data³ for which we generated 1240k
- 388 capture data and 11 previously published^{3,4}), 68 newly reported individuals from relevant
- ancient populations and 298 previously published^{2–4,20–37} individuals (Supplementary Table 1).
- We kept 591,642 autosomal SNPs after intersecting autosomal SNPs in the 1240k capture with
- the analysis set of 594,924 SNPs from Lazaridis et al.²².
- 393 -HOIII includes the same set of ancient samples and 300 present-day individuals from 142
- populations sequenced to high coverage as part of the Simons Genome Diversity Project³⁸. For
- this dataset, 1,054,671 autosomal SNPs were used, excluding SNPs of the 1240k array located
- on sex chromosomes or with known functional effects.
- 398 For both datasets, ancient individuals were merged by randomly sampling one read at each SNP
- position, discarding the first and the last two nucleotides of each read.

Principal component analysis

- 401 We carried out principal component analysis (PCA) on the HO dataset using the smartpca
- 402 program in EIGENSOFT⁷⁰. We computed principal components on 990 present-day West
- 403 Eurasians and projected ancient individuals using lsqproject: YES and shrinkmode: YES.

ADMIXTURE analysis

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- We performed model-based clustering analysis using ADMIXTURE⁴¹ on the *HO* reference
- dataset, including 2,572 present-day individuals from worldwide populations and the ancient
- 407 individuals. First, we carried out LD-pruning on the dataset using PLINK⁷¹ with the flag --
- 408 indep-pairwise 200 25 0.4, keeping 306,393 SNPs. We ran ADMIXTURE with the cross
- validation (--cv) flag specifying from K=2 to K=20 clusters, with 20 replicates for each value of
- 410 K and keeping for each value of K the replicate with highest log likelihood. In Extended Data
- 411 Fig. 1b we show the cluster assignments at K=8 of newly reported individuals and other
- relevant ancient samples for comparison. This value of K was the lowest for which components
- of Caucasus hunter-gatherers (CHG) and European hunter-gatherers were maximized.

414 f-statistics

- We computed f-statistics on the *HOIII* dataset using ADMIXTOOLS⁶⁹ with default parameters
- 416 (Supplementary Information, section 4). We used qpDstat with f4mode: Yes for f_4 -statistics and
- 417 qp3Pop for outgroup f3-statistics. We computed standard errors using a weighted block
- 418 jackknife⁷² over 5 Mb blocks.

Inference of mixture proportions

- We estimated ancestry proportions on the HOIII dataset using qpAdm² and a basic set of 9
- 421 Outgroups: Mota, Ust Ishim, MA1, Villabruna, Mbuti, Papuan, Onge, Han, Karitiana. For
- some analyses (Supplementary Information, section 6) we added additional outgroups to this
- 423 basic set.

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424 Allele frequency estimation from read counts

- We used allele counts at each SNP to perform maximum likelihood estimation of allele
- frequencies in ancient populations as in ref.⁴. In Extended Data Fig. 5, we show derived allele
- frequency estimates at three SNPs of functional importance for different ancient populations.

Data availability

- 429 All 1240k and mitochondrial capture sequencing data is available from the European Nucleotide
- 430 Archive, accession number XXXXXXXX [to be made available on publication].
- Pseudo haploid genotype data is available from the Reich Lab website at [to be made available
- 432 on publication].

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Author Contributions

- 458 S.B., M.E.A, N.R., A.S.-N., A.M., N.B., M.F., E.H., M.M., J.O., K.S., R.P., J.K., W.H., I.B. and
- D.R. performed or supervised wet laboratory work. G.T.C. undertook the radiocarbon dating of
- a large fraction of the British samples. I.A., K.K., A.B., K.W.A., A.A.F., E.B., M.B.-B., D.B.,
- 461 C.B., C.Bo., L.B., T.A., L.Bü., S.C., L.C.N., O.E.C., G.C., B.C., A.D., K.E.D., N.D., M.E.,

- 462 C.E., M.K., J.F.F., H.F., C.F., M.G., R.G.P., M.H.-U., E.Had., G.H., N.J., T.K., K.M., S.P.,
- 463 P.L., O.L., A.L., J.L.M., T.M., J.I.M, K.Mc., M.B.G., A.Mo., G.K., V.K., A.C., R.Pa., A.E.,
- 464 K.Kö., T.H., J.L.C., C.L., M.P.P., P.W., T.D.P., P.P., P.-J.R., P.R., R.R., M.A.R.G., A.S., J.S.,
- 465 A.M.S., V.S., L.V., J.Z., D.C., T.Hi., V.H., A.Sh., K.-G.S., P.W.S., R.P., J.K., W.H., I.B., C.L.-
- 466 F. and D.R. assembled archaeological material. I.O., S.M., T.B., A.M., E.A., M.L., I.L., N.P.,
- 467 Y.D., Z.F., D.F., P.d.K., M.G.T. and D.R. analysed or supervised analysis of data. I.O., C.L.-F.
- and D. R. wrote the manuscript with input from all co-authors.

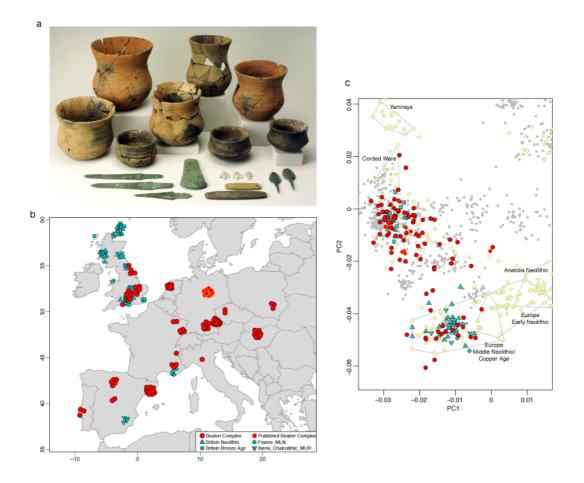


Figure 1. Genetic structure of individuals included in this study. a, Beaker Complex grave goods from La Sima III barrow⁷³. Photo: Alejandro Plaza, Museo Numantino. **b**, Geographic distribution of samples with new genome-wide data, with random jitter added for clarity. **c**, Principal component analysis of 990 present-day West Eurasian individuals (grey dots), with previously published (pale yellow) and new ancient samples projected onto the first two principal components. This figure is a zoom of Extended Data Fig 1a.

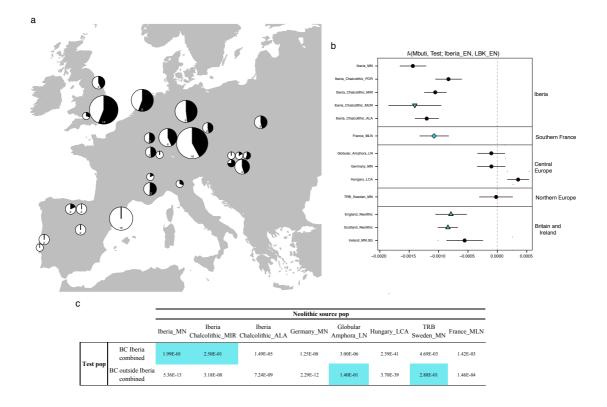


Figure 2. Investigating the genetic makeup of Beaker Complex individuals. a, Proportion of Steppe-related ancestry (shown in black) in Beaker Complex groups, computed with qpAdm under the model Yamnaya_Samara + Anatolia_Neolithic + WHG. The area of the pie is proportional to the number of individuals (shown inside the pie if more than one). See Supplementary Information, section 6 for mixture proportions and standard errors. b, f-statistics of the form f_4 (Mbuti, Test; Iberia_EN, LBK_EN) computed for European populations before the emergence of the Beaker Complex. Error bars represent ± 1 standard errors. c, Testing different populations as a source for the Neolithic farmer ancestry component in Beaker Complex individuals. The table shows the P-values (highlighted if >0.05) for the model: Yamnaya Samara + Neolithic farmer population. BC, Beaker complex.

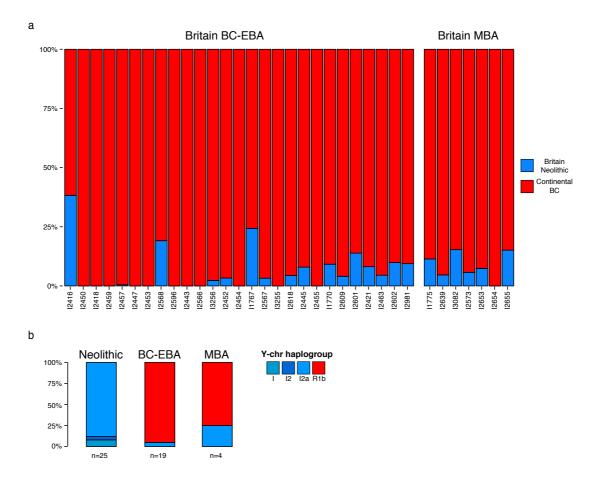


Figure 3. Population transformation in Britain associated with the arrival of the Beaker Complex. a, Modelling Beaker Complex and Bronze Age individuals from Britain as a mixture of continental Beaker Complex (red, represented by Beaker Complex samples from Oostwoud) and Britain_Neolithic (blue). Individuals are ordered chronologically (oldest on the left) and included in the plot if represented by more than 100,000 SNPs. See Supplementary Information, section 6 for mixture proportions and standard errors. **b**, Y-chromosome haplogroup distribution in males from Britain. EBA, Early Bronze Age; MBA, Middle Bronze Age. BC, Beaker complex.

Supplementary Tables

Supplementary Table 1. Ancient individuals included in this study.

Supplementary Table 2. Mitochondrial haplogroup calls for individuals with newly reported data.

Supplementary Table 3. Y-chromosome calls for males with newly reported data .

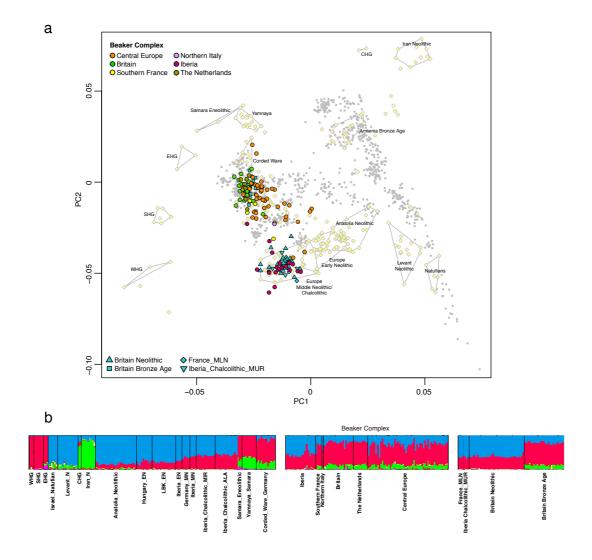
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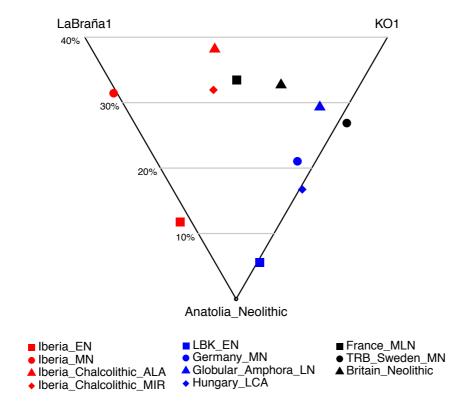
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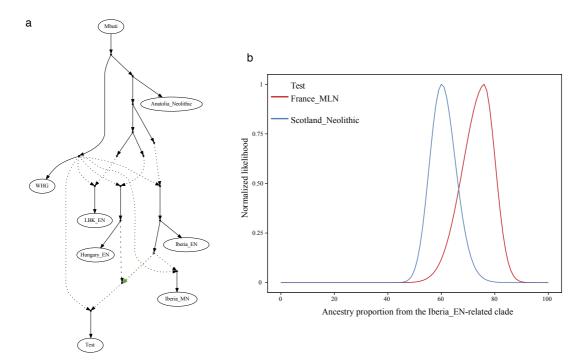


Extended Data Figure 1. Population structure. a, Principal component analysis of 990 present-day West Eurasian individuals (grey dots), with previously published (pale yellow) and new ancient samples projected onto the first two principal components. **b,** ADMIXTURE clustering analysis with k=8 showing ancient individuals. E/M/MLN, Early/Middle/Middle Late Neolithic; W/E/S/CHG, Western/Eastern/Scandinavian/Caucasus hunter-gatherers.

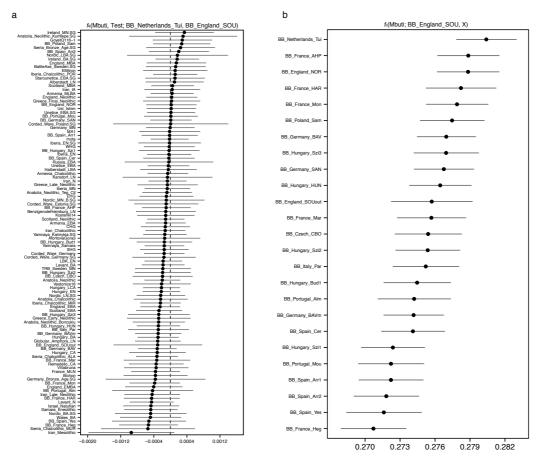


Extended Data Figure 2. Hunter-gatherer affinities in Neolithic/Copper Age Europe.

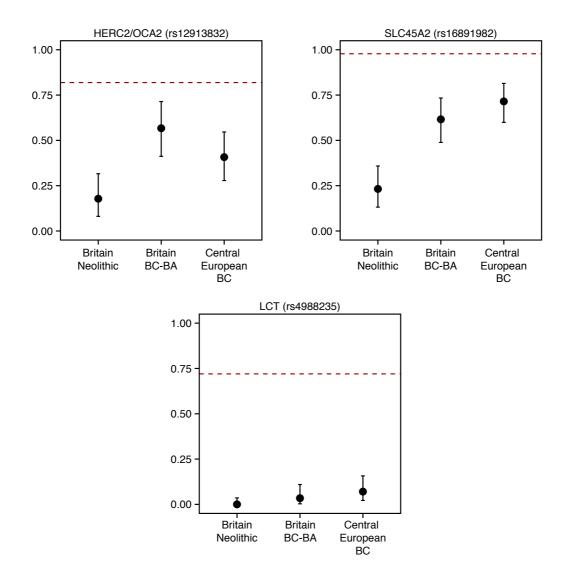
Differential affinity to hunter-gatherer individuals (LaBraña1 36 from Spain and KO1 39 from Hungary) in European populations before the emergence of the Beaker Complex. See Supplementary Information, section 6 for mixture proportions and standard errors computed with qpAdm.



Extended Data Figure 3. Modelling the relationships between Neolithic populations. a, Admixture graph fitting a *Test* population as a mixture of sources related to both Iberia_EN and Hungary_EN. b, Likelihood distribution for models with different proportions of the source related to Iberia_EN (green admixture edge in (a)) when *Test* is Great Britain_Neolithic or France_MLN.



Extended Data Figure 4. Genetic affinity between Beaker complex individuals from southern Great Britain and the Netherlands. a, f-statistics of the form f_4 (Mbuti, Test; BB_Netherlands_Tui, BB_Great Britain_SOU). Negative values indicate that Test is closer to BB_Netherlands_Tui than to BB_Great Britain_SOU, and the opposite for positive values. Error bars represent ± 3 standard errors. b, Outgroup- f_3 statistics of the form f_3 (Mbuti; BB_Great Britain_SOU, X) measuring shared genetic drift between BB_Great Britain_SOU and other Beaker Complex groups. Error bars represent ± 1 standard errors.



Extended Data Figure 5. Derived allele frequencies at three SNPs of functional importance. Error bars represent 1.9-log-likelihood support interval. The red dashed lines show allele frequencies in the 1000 Genomes GBR population (present-day people from Great Britain). BC, Beaker Complex; BA, Bronze Age.

Extended Data Table 1. 62 Newly reported radiocarbon dates

Sample	Date	Location	Country
I4145	2279–2033 calBCE (3740±35 BP, Poz-84460)	Kněževes	Czech Republic
I1392	2832–2476 calBCE (4047±29 BP, MAMS-25935)	Hégenheim Necropole, Haut-Rhin	France
I4144	2572–2512 calBCE (3955±35 BP, Poz-84553)	Osterhofen-Altenmarkt	Germany
E09537_d	2471–2300 calBCE (3909±29 BP, MAMS 29074)	Unterer Talweg 58-62, Augsburg, Bavaria	Germany
I4249	2336-2141 calBCE (3802±26 BP, BRAMS1217)	Irlbach LKR	Germany
E09538	2464–2212 calBCE (3870±30 BP, MAMS 29075)	Unterer Talweg 58-62, Augsburg, Bavaria	Germany
I3592	2458-2204 calBCE (3844±33 BP, BRAMS-1218)	Alburg-Lerchenhaid, Spedition Häring, Bavaria	Germany
I4250	2434-2150 calBCE (3825±26 BP, BRAMS1219)	Irlbach LKR	Germany
I3593	2398-2146 calBCE (3817±26 BP, BRAMS-1215)	Alburg-Lerchenhaid, Spedition Häring, Bavaria	Germany
I3590	2339-2143 calBCE (3802±26 BP, BRAMS-1217)	Alburg-Lerchenhaid, Spedition Häring, Bavaria	Germany
12657	3952–3781 calBCE (5052±30 BP, SUERC-68701)	Macarthur Cave	Great Britain
I2633	3766-3642 calBCE (4911±32 BP, SUERC-68634)	Tulloch of Assery B	Great Britain
12659	3762–3644 calBCE (4914±27 BP, SUERC-68702)	Distillery Cave	Great Britain
I2691	3701–3640 calBCE (4881±25 BP, SUERC-68704)	Distillery Cave	Great Britain
12796	3706–3536 calBCE (4856±33 BP, SUERC-69074)	Point of Cott, Orkney	Great Britain
I2634	3704–3535 calBCE (4851±34 BP, SUERC-68638)	Tulach an t'Sionnach	Great Britain
I2635		Tulloch of Assery A	Great Britain
I2636	3653–3390 calBCE (4796±37 BP, SUERC-68639)		Great Britain
	3520–3362 calBCE (4651±33 BP, SUERC-68640)	Holm of Papa Westray North	
I2988	3517–3362 calBCE (4645±29 BP, SUERC-68711)	Clachaig	Great Britain
I2660	3514–3353 calBCE (4631±29 BP, SUERC-68703)	Distillery Cave	Great Britain
I2650	3500–3360 calBCE (4754±36 BP, SUERC-68642)	Holm of Papa Westray North	Great Britain
I2637	3510–3340 calBCE (4697±33 BP, SUERC-68641)	Holm of Papa Westray North	Great Britain
I2605	3632–3373 calBCE (4710±35 BP, Poz-83483)	Eton Rowing Course	Great Britain
I2980	3361–3102 calBCE (4530±33 BP, SUERC-69073)	Point of Cott, Orkney	Great Britain
I2651	3330–3090 calBCE (4525±36 BP, SUERC-68643)	Holm of Papa Westray North	Great Britain
I3085	3339–3027 calBCE (4471±29 BP, SUERC-68724)	Isbister, Orkney	Great Britain
I2978	3336–3024 calBCE (4464±29 BP, SUERC-68725)	Isbister, Orkney	Great Britain
I2934	3327–3036 calBCE (4466±33 BP, SUERC-69071)	Isbister, Orkney	Great Britain
I2935	3336-3012 calBCE (4451±29 BP, SUERC-68723)	Isbister, Orkney	Great Britain
I2979	3334–2942 calBCE (4447±29 BP, SUERC-68726)	Isbister, Orkney	Great Britain
I2631	3098-2907 calBCE (4384±36 BP, SUERC-68633)	Quoyness	Great Britain
I2933	3011–2886 calBCE (4309±29 BP, SUERC-68722)	Isbister, Orkney	Great Britain
I2977	3009–2764 calBCE (4275±33 BP, SUERC-69072)	Isbister, Orkney	Great Britain
12630	2581–2464 calBCE (3999±32 BP, SUERC-68632)	Isbister, Orkney	Great Britain
I2932	2571–2348 calBCE (3962±29 BP, SUERC-68721)	Isbister, Orkney	Great Britain
I2612	2465–2209 calBCE (3865±35 BP, Poz-83492)	Hasting Hill, Sunderland, Tyne and Wear	Great Britain
I2416	2470-2285 calBC (3830±30 BP, Beta-432804)	Amesbury Down, Wiltshire	Great Britain
I2418	2440–2200 calBCE (3835±25 BP, NZA-32788)	Amesbury Down, Wiltshire	Great Britain
I2565	2470–2140 calBCE (3829±38 BP, OxA-13562)	Amesbury Down, Wiltshire	Great Britain
I2459	2460–2140 calBCE (3829±30 BP, SUERC-54823)	Amesbury Down, Wiltshire	Great Britain
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I2457	2480-2280 calBCE (3890±30 BP, SUERC-36210)	Amesbury Down, Wiltshire	Great Britain
I2457	2200-2031 calBCE (3717±28 BP, SUERC-69975)	Amesbury Down, Wiltshire	Great Britain
I2453	2289–2041 calBCE (3760±35 BP, Poz-83404)	West Deeping	Great Britain
I2445	2137–1930 calBCE (3650±35 BP, Poz-83407)	Yarnton	Great Britain
I2596	2280–2030 calBCE (3739±30 BP, NZA-32484)	Amesbury Down, Wiltshire	Great Britain
I2566	2210–2030 calBCE (3734±25 BP, NZA-32490)	Amesbury Down, Wiltshire	Great Britain
I2452	2195–1920 calBCE (3700±30 BP, Beta-444979)	Dairy Farm, Willington	Great Britain
I2452	2277–2030 calBCE (3735±35 BP, Poz-83405)	Dairy Farm, Willington	Great Britain
12598	2140–1940 calBCE (3664±30 BP, NZA-32494)	Amesbury Down, Wiltshire	Great Britain
I2460	2030–1820 calBCE (3575±27 BP, SUERC-53041)	Amesbury Down, Wiltshire	Great Britain
I2609	2023-1772 calBCE (3560±40 BP, Poz-83423)	Hexham Golf Course, Northumberland	Great Britain
I2610	1936–1746 calBCE (3515±35 BP, Poz-83498)	Summerhill, Blaydon, Tyne and Wear	Great Britain
I1775	1693–1600 calBCE (3344±27 BP, OxA-14308)	Great Orme Mines, Llandudno, North Wales	Great Britain
I2574	1415–1228 calBCE (3065±36 BP, SUERC-62072)	North Face Cave, Llandudno, North Wales	Great Britain
I2786	2459–2206 calBCE (3850±35 BP, Poz-83639)	Szigetszentmiklós,Felső Ürge-hegyi dűlő	Hungary
I2787	2458–2202 calBCE (3840±35 BP, Poz-83640)	Szigetszentmiklós, Felső Ürge-hegyi dűlő	Hungary
I2741	2458–2154 calBCE (3835±35 BP, Poz-83641)	Szigetszentmiklós, Felső Ürge-hegyi dűlő	Hungary
I4229	2289–2135 calBCE (3775±25 BP, PSU-1750)	Cova da Moura	Portugal
I0826	2833–2480 calBCE (4051±28 BP, MAMS-25940)	Paris Street, Cerdanyola, Barcelona	Spain
I0020	2571–2350 calBCE (3965±29 BP, MAMS-25937)	Paris Street, Cerdanyola, Barcelona	Spain
I0257 I0462	2566–2346 calBCE (3950±29 BP, MAMS-25937)	Arroyal I, Burgos	Spain Spain
I0462 I0825			-
10023	2474–2300 calBCE (3915±29 BP, MAMS-25939)	Paris Street, Cerdanyola, Barcelona	Spain

Extended Data Table 2. Sites with new genome-wide data reported in this study.

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