

## New Data About the Virus Susceptibility of Some *Chenopodium* Species

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The aim of our study was to investigate the susceptibility of some *Chenopodium* species (*Chenopodium album*, *C. glaucum*, *C. berlandieri*, *C. ugandae*) to six viruses (*Alfalfa mosaic virus*, *Cucumber mosaic virus*, *Obuda pepper virus*, *Potato virus Y*, *Sowbane mosaic virus*, *Zucchini yellow mosaic virus*). Fourteen plants of each species were mechanically inoculated and virus susceptibility was evaluated on the basis of symptoms and back inoculation. A series of new host-virus relations were determined.

Keywords: Virus susceptibility, *Chenopodium* species.

The genus *Chenopodium* involves 200 species of great economical and virological importance. *Chenopodium quinoa* Willd. is an important food of certain ethnic groups, while *C. album* L. is one of the worst weeds of the world (Holm et al., 1977; Hegi, 1979). So far susceptibility of nearly 40 *Chenopodium* species is known to different viruses. The most known are in this respect *Chenopodium quinoa* Willd. and *C. amaranticolor* Coste et Reyn., which could be used in separation of viruses and for virus diagnosis as local assay and propagative hosts (Edwardson, 1974; Horváth, 1976, 1983; Schmelzer and Wolf, 1977). *Chenopodium album* is known as artificial and natural host of over 60 viruses (Schmelzer and Wolf, 1977; Horváth, 1986). *Chenopodium glaucum* L. has less virological importance with susceptibility to nine viruses [*Alfalfa mosaic virus* (AMV), *Bean yellow mosaic virus* (BYMV), *Beet yellows virus* (BYV), *Grapevine fanleaf virus* (GFLV), *Potato virus M* (PVM), *Robinia mosaic virus* (RMV), *Strawberry latent ringspot virus* (SLRSV), *Tobacco mosaic virus* (TMV), *Turnip mosaic virus* (TuMV)] (Schmelzer and Wolf, 1977; Horváth, 1983). So far the virological importance of *Chenopodium berlandieri* Moq. and *C. ugandae* Aellen was not known at all. Therefore the aim of our study was to investigate the reaction of *C. album*, *C. berlandieri*, *C. glaucum* and *C. ugandae* to different viruses.

### Materials and Methods

Seeds of *Chenopodium album*, *C. berlandieri*, *C. glaucum* and *C. ugandae* were sown in sterile boxes in vector free glasshouse. The seedlings were planted in plastic pots (12 cm in diameter) containing a soil mixture of sand (pH: 6.96, humus %: 0.27): peat

(pH: 6.78, humus %: 9.98) in a ratio of 1:3. Fourteen plants of each species were mechanically inoculated at 6–8 leaves stage with six viruses using Sørensen phosphate buffer (pH 7.2) in the ratio of 1:1 (Table 1).

**Table 1**

Viruses (strains and isolates) used for inoculation and their propagative hosts

Viruses	Strains or isolates	Acronyms	Propagative hosts
<i>Cucumber mosaic virus</i>	U/246	CMV-U/246	<i>Nicotiana tabacum</i> 'Xanthi-nc'
<i>Alfalfa mosaic virus</i>	Asc	AMV-Asc	<i>Chenopodium amaranticolor</i>
<i>Sowbane mosaic virus</i>	H	SoMV-H	<i>Chenopodium quinoa</i>
<i>Zucchini yellow mosaic virus</i>	10	ZYMV-10	<i>Cucumis sativus</i> 'Delicatesse'
<i>Obuda pepper virus</i>	pepper	ObPV-pepper	<i>Nicotiana tabacum</i> 'Samsun'
<i>Potato virus Y</i>	NTN	PVY <sup>NTN</sup>	<i>Nicotiana tabacum</i> 'Xanthi-nc'

The inoculated plants were symptomatologically tested for infection. In order to confirm the results of symptomatology, back inoculations were also carried out to *N. tabacum* 'Xanthi-nc', *N. tabacum* 'Samsun', *C. amaranticolor*, *C. quinoa* and *C. sativus* 'Delicatesse' as indicator plants.

## Results and Discussion

A lot of new host-virus relations were determined. *Chenopodium album*, as local and systemic experimentally host of SoMV-H and AMV-Asc was described (Fig. 1). Other viruses induced only local symptoms on *C. album* (Table 2).

In an earlier study Horváth (1983) experienced only local symptoms on *C. album* var. *centrorubrum* due to AMV infection. It shows that great differences may be in virus susceptibility of subspecific taxa. *Chenopodium album* belongs to the most serious and widespread weeds of the world (Holm et al., 1977). In Hungary, on the basis of the four

**Table 2**Symptoms on *Chenopodium* species due to virus infections

Viruses used for inoculation	<i>Chenopodium</i> species			
	<i>C. album</i>	<i>C. berlandieri</i>	<i>C. glaucum</i>	<i>C. ugandae</i>
	Symptoms (local/systemic)*			
CMV-U/246	Chl/-	Chl, NI/-	-/-	Ni
AMV-Asc	Chl/YeMo	Chl, NI/YeMo, Led	-/-	-/-
SoMV-H	Chl/Led, Chl	Chl, NI/Led, Mo	-/-, L	-/-, L
ZYMV-10	Chl/-	Chl/-	-/-	Ni
ObPV-pepper	NI/-	Chl, NI/Mo, Led	Chl, NI/-	Chl/-
PVY <sup>NTN</sup>	Chl, NI/-	Chl, NI/-	-/-	Ni

\*- Symptomless; Chl: chlorotic lesions; NI: necrotic lesions; YeMo: yellow mosaic; Led: leaf deformation; Mo: mosaic; L: latent infection; Ni: not investigated.

National Weed Surveys (1947–1997) it occupies the 3rd, 3rd, 2nd and 4th positions, respectively (Tóth and Spilák, 1998; Hunyadi et al., 2000). *Chenopodium album* is known as artificial and natural host of more than 60 viruses (Schmelzer and Wolf, 1977; Horváth, 1986). In Hungary, *C. album* samples collected from different agro- and water ecosystems were naturally infected only with three viruses [*Cucumber mosaic virus*, *Potato virus S* (PVS), *Potato leafroll virus* (PLRV)] (Kiss et al., 2002; Kazinczi, 2003). Its virological importance was satisfied on the evidence that it serves as food plant of *Frankliniella occidentalis*, which is the vector of *Tomato spotted wilt virus* (TSWV) (Cho et al., 1989).

As far as we know, so far no data was available about virological role of *C. berlandieri* and *C. ugandae*. In our experiments in case of *C. berlandieri* three local and three local and systemic host-virus relations were determined. Systematization of ObPV-pepper is very interesting, because earlier *Chenopodium* species as local hosts of the most *tobamoviruses* were described (Brunt et al., 1996) (Table 2, Fig. 2). *Chenopodium ugandae* and *C. glaucum* as local hosts of ObPV-pepper and systemic latent host of SoMV-H became known (Table 2). Neither local nor systemic symptoms could be seen due to SoMV-H infection, but back inoculation of systemic leaves to test plants (*C. amaranticolor* and *C. quinoa*) proved systemic latent host-virus relation. In spite the fact that AMV has been described from *C. glaucum* earlier (Schmelzer and Wolf, 1977; Horváth, 1983) our inoculation was unsuccessful. *Chenopodium glaucum* as local host of ObPV-pepper, non-host of CMV-U/246, PVY<sup>NTN</sup> and ZYMV-10, and latent host of SoMV-H are described here for the first time.

All the studied four *Chenopodium* species have proved as hosts (local and systemic or systemic latent ones) of SoMV-H. Most *Chenopodium* species as artificial hosts of SoMV-H are known (Bennett and Costa, 1961; Kado, 1971). Natural occurrence of SoMV on *Chenopodium hybridum* L. and *Chenopodium murale* L. was described in Hungary (Horváth et al., 1993) and in the former Yugoslavia (Juretic, 1976). Our investigation confirmed the fact that *Chenopodium* species could be potential infection sources in potato agro-ecosystems, regarding the fact that SoMV is easily transmitted by seeds, pollen and mechanical way (Bos and Huijberts, 1996; Kazinczi and Horváth, 1998; Kazinczi et al., 2000), and it is also well known that ‘Puebla’ potato variety as natural host of SoMV is already known in Mexico (Salazar, 1996).

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## Literature

- Bennett, C. W. and Costa, A. S. (1961): Sowbane mosaic caused by a seed transmitted virus. *Phytopathology* 51, 546–550.
- Bos, L. and Huijberts, N. (1996): Occurrence and transmission of sowbane mosaic virus in seed from naturally infected plants of spinach (*Spinacea oleracea*). *Eur. J. Plant Pathol.* 102, 707–711.

- Brunt, A. A., Crabtree, K., Dallwitz, M. J., Gibbs, A. J. and Watson, L. (1996): Viruses of Plants. Descriptions and Lists from the VIDE Database. CAB International, Wallingford, 1484. pp.
- Cho, J. J., Mau, R. F., German, T. L., Hartmann, R. W., Yudin, L. S., Gonsalves, D. and Provvidenti, R. (1989): A multidisciplinary approach to management of tomato spotted wilt virus in Hawaii. *Plant Dis.* 73, 375–383.
- Edwardson, J. R. (1974): Host ranges of viruses in the PVY-group. *Florida Agr. Exp. Stat. Monograph. Ser. 5*, 1–225.
- Hegi, G. (1979): *Illustrierte Flora von Mitteleuropa III*. Verlag Paul Parey, Berlin.
- Holm, L. G., Plucknett, D. L., Pancho, J. V. and Herberger, J. P. (1977): *The World's Worst Weeds*. Univ. Hawaii Press, Honolulu, pp. 1–609.
- Horváth, J. (1976): *Vírus-gazdanövénykörök és vírusdifferenciálás. (Virus host range and differentiation of viruses.)* Akadémiai Doktori Értekezés, Budapest–Keszthely, pp. 1–607.
- Horváth, J. (1983): New artificial hosts and non-hosts of plant viruses and their role in the identification and separation of viruses. XVIII: Concluding remarks. *Acta Phytopath. Acad. Sci. Hung.* 18, 121–161.
- Horváth, J. (1986): Újabb adatok a növények vírusfogékonyságáról. 5. *Chenopodiaceae (Chenopodium) fajok. [New data on the virus susceptibility of plants. 5. Chenopodiaceae (Chenopodium species).]* *Bot. Közlem.* 73, 229–242.
- Horváth, J., Juretic, N., Wolf, P. and Pintér, Cs. (1993). Natural occurrence of sowbane mosaic virus on *Chenopodium hybridum* L. in Hungary. *Acta Phytopath. Entomol. Hung.* 28, 379–389.
- Hunyadi, K., Béres, I. and Kazinczi, G. (2000): *Gyomnövények, gyomirtás, gyombiológia. (Weeds, weed control, weed biology.)* Mezőgazda Kiadó, Budapest, pp. 1–630.
- Juretic, N. (1976): Some data on sowbane mosaic virus isolated from *Chenopodium murale* in Yugoslavia. *Acta Bot. Croat.* 35, 33–39.
- Kado, C. I. (1971): Sowbane mosaic virus. *CMI/AAB Descriptions of Plant Viruses* 64, 1–4.
- Kazinczi, G. (2003): A vírusok alternatív gazdái: gyomnövények. (Alternative hosts of viruses: weeds.) *Akadémiai Doktori Értekezés, Keszthely*, pp. 1–122.
- Kazinczi, G. and Horváth, J. (1998): Transmission of sowbane mosaic sobemovirus by seeds of *Chenopodium* species and viability of seeds. *Acta Phytopath. Entomol. Hung.* 33, 237–242.
- Kazinczi, G., Horváth, J. and Lukács, D. (2000): Germination characteristics of *Chenopodium* seeds derived from healthy and virus infected plants. *Z. PflKrankh. PflSchutz, Sonderh.* 17, 63–67.
- Kiss, E., Kazinczi, G., Horváth, J., Kobza, S., Baranyi, T., Varga, M., Havasréti, B. and Fehér, A. (2002): Virus disease problems on field cucumber in Hungary with some international aspects. *Acta Phytopath. Entomol. Hung.* 37, 317–327.
- Salazar, L. F. (1996): *Potato Viruses and Their Control*. IPC Press, Lima, pp. 1–214.
- Schmelzer, K. and Wolf, P. (1977): Wirtspflanzen und ihre Viren, Viroseen und Mykoplasmosen. In: M. Klinowski (ed.): *Pflanzliche Virologie. Registerband Verzeichnisse und Übersichten zu den Viroseen in Europa*. Akademie Verlag, Berlin, pp. 53–189.
- Tóth, Á. and Spilák, K. (1998): A IV. országos gyomfelvételezés tapasztalatai. (Results of the fourth national weed survey in Hungary.) 8. *Növényvédelmi Fórum, Keszthely, 1998. Abstr. p.* 49.