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**Single Particle Dynamics at the Intrinsic Surface of
Various Apolar, Aprotic Dipolar and Hydrogen Bonding
Liquids, As Seen from Computer Simulations**

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Running title: Single Particle Dynamics at the Liquid Surface

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Abstract

We investigate the single molecule dynamics at the intrinsic liquid/vapor interface of five different molecular liquids (carbon tetrachloride, acetone, acetonitrile, methanol and water). After assessing that the characteristic residence times in the surface layer are long enough for a meaningful definition of several transport properties within the layer itself, we characterize the dynamics of the individual molecules at the liquid surface by analyzing their normal and lateral mean square displacements and lateral velocity autocorrelation functions and, in the case of the hydrogen bonding liquids (i.e., water and methanol), also the properties of the hydrogen bonds. Further, dynamical properties as well as the clustering of the molecules residing unusually long in the surface layer are also investigated. The global picture emerging from this analysis is that of a noticeably enhanced dynamics of the molecules at the liquid surface, with diffusion coefficients up to four times larger than in the bulk, and the disappearance of the caging effect at the surface of all liquids but water. The dynamics of water is dominated by the strong hydrogen bonding structure also at the liquid surface.

1. Introduction

The description of soft or fluid interfaces at the molecular level has become the focus of intensive scientific investigation in the past few decades. Understanding the properties of soft interfaces is of great importance both from the fundamental point of view and also from the point of view of applications. Thus, since the molecules located at such interfaces experience a markedly different local environment from those inside the bulk fluid phase, the structural and dynamical properties and even the reactivity of these interfacial molecules are also characteristically different from those in the bulk phases. As a consequence, soft interfaces play a key role in a number of processes, many of which are also of industrial importance, from catalysis to extraction or from adsorption to surface micellization.

However, in spite of their importance both in pure and in applied science, a meaningful investigation of soft interfaces on the molecular level was hindered for a long time by the lack of experimental methods that are able to selectively probe the interfacial molecules. The development of such methods, like nonlinear spectroscopy techniques¹ (e.g., second harmonic generation²⁻⁴ or sum frequency generation^{5,6} spectroscopies), X-ray^{7,8} and neutron^{7,9} reflection methods, or time resolved fluorescence anisotropy measurements^{10,11} at the end of the last century have led to a rapid increase of studies in this field since then. Further, the rapid development of the routinely available computing capacities in the past decade enabled us to meaningfully investigate the molecular level properties of soft interfaces also by computer simulation methods.¹² As a consequence, our understanding of the properties of soft interfaces improved considerably in the past two decades.

In investigating soft interfaces by computer simulation methods one has to face the problem that these interfaces are corrugated, on the molecular length scale, by thermal capillary waves. As a consequence, finding the accurate location of the interface at every point along its macroscopic plane (or, equivalently, finding the full list of molecules that are located right at the interface) is not a simple task at all if the system is seen at atomistic resolution. In the early years the majority of the simulation studies simply neglected this problem, and defined the interface in a reference frame fixed to the simulation box as the region of intermediate densities between the two bulk phases. Neglecting the effect of the capillary waves was later shown to introduce a systematic error of unknown magnitude of any interfacial properties calculated this way,¹³⁻¹⁷ and this error can even propagate to the thermodynamic properties of the system.¹⁸ The origin of this systematic error is simply the

misidentification of a large number of molecules located at the boundary of the two phases as non-interfacial, as well as that of molecules located in a bulk-like local environment as interfacial ones. Further, any meaningful comparison of simulation results with those of surface sensitive experiments, probing selectively the molecules that are located at the boundary of the two phases, requires the unambiguous identification of these molecules also in the computer simulation.

Although the importance of locating the true, capillary wave corrugated, so-called intrinsic interface in computer simulations was already realized in the first interfacial simulations,^{19,20} the first method that was able to accurately locate the intrinsic surface of a fluid phase was only proposed more than a decade later, in the pioneering work of Chacón and Tarazona.²¹ In their Intrinsic Sampling Method (ISM) the intrinsic surface is found as the surface of minimum area that covers a set of pre-selected pivot atoms, the list of which is determined in a self-consistent way.^{22,23} Since then a number of intrinsic surface analyzing methods, based on detecting the outermost molecules in slabs parallel with the macroscopic surface normal axis,^{24,25} on the vicinity of molecules of the opposite phase,^{26,27} or on the accessibility by a spherical probe from the opposite phase¹³ have been proposed, several of which being even free from the assumption that the interface is macroscopically planar.²⁸⁻³⁰ Among these methods, the Identification of the Truly Interfacial Molecules (ITIM)¹³ turned out to be an excellent compromise between accuracy and computational cost.²⁷

Having the intrinsic surface of the fluid phase detected, the variation of a number of physical properties, e.g., density,^{21-25,31,32} energy,³² solvation free energy,^{33,34} electrostatic potential,³⁵ or lateral pressure^{32,36} can be calculated across the interface, either as a profile relative to the position of the intrinsic surface or in a layer-by-layer manner.³² The calculation of such intrinsic profiles proved to be essential in understanding a number of soft interface-related phenomena in the past few years, such as the molecular level explanation of the surface tension anomaly of water,^{37,38} the determination of how the subsequent molecular layers beneath the surface contribute to the surface tension,³⁶ investigation of Newton black films³⁹ and the immersion depth of various surfactants into the liquid phase,⁴⁰ the molecular level description of the surface of various ionic⁴¹⁻⁴⁵ and molecular liquids^{13-15,46-48} and liquid mixtures^{15-17,49-51} as well as aqueous electrolyte solutions.³⁵

In spite of the relative abundance of recent simulation studies concerning the structural and thermodynamic properties of soft interfaces, little care has been taken to understand their dynamical properties. In fact, although several studies focused on the dynamics of interfacial molecules,⁵²⁻⁶³ only a handful of them were considering the true, capillary wave corrugated,

intrinsic liquid surface.⁶⁰⁻⁶³ Duque et al. studied the surface of a Lennard-Jones liquid,⁶⁰ whereas in a recent study we addressed several questions concerning the dynamics of the molecules at the free water surface.⁶³ These two studies revealed important differences between the surface dynamics of these liquids, among which the most interesting one is probably that while the water molecules stay, on average, considerably longer at the liquid surface than the characteristic time of their diffusion within the surface layer,⁶³ these two time scales are equal in the case of the Lennard-Jones surface.⁶⁰ It was also found that the water molecules staying longest at the surface are rather weakly bound to the molecules forming the second layer.⁶³

The different surface dynamics observed for Lennard-Jones particles and water naturally gives rise to the question how the surface dynamics of the molecules depend on the intermolecular interactions characteristic of the liquid phase. To address this question and further improve our understanding of the surface dynamics of liquids we present here a detailed investigation of the dynamics of the molecules located at the intrinsic liquid-vapor interface of five molecular liquids, namely carbon tetrachloride, acetone, acetonitrile, methanol, and water. This set of molecules covers the range of interactions from weak van der Waals through dipolar yet aprotic to hydrogen bonding ones. Since a similar study was recently published for water,⁶³ it is largely regarded here as a reference system, to which the properties of the other systems can be compared. In this paper we focus our attention to the following questions: (i) how the mean surface residence time of the molecules is related to the time scale of various dynamical processes of the surface molecules (e.g., diffusion, vibrational motion, H-bond lifetime in the case of hydrogen bonding liquids); (ii) how the diffusion of the molecules within the surface layer is related to their diffusion in the bulk liquid phase; (iii) how different or similar are single molecular motions at the liquid surface and in the bulk liquid phase; (iv) how, if at all, these dynamical properties of the surface molecules are related to their surface residence; and finally (v) how the answers to the above questions depend on the intermolecular interactions acting in the liquid phase.

The paper is organized as follows. In sec. 2 details of the calculations performed and methods used are given. The obtained results are presented and discussed in detail in sec. 3. Finally, the main conclusions of this study are summarized in sec. 4.

2. Methods

2.1. Molecular Dynamics Simulations. The molecular dynamics (MD) simulations performed are described in detail in our previous paper,³⁶ thus, they are only briefly reminded here. MD simulations of the liquid-vapor interface of five molecular liquids, namely CCl₄, acetone, acetonitrile, methanol, and water have been performed on the canonical (N,V,T) ensemble. The set of molecules considered corresponds to markedly different intermolecular interactions: while in CCl₄ only van der Waals interaction acts between the molecules, acetone and acetonitrile are characterized by dipolar forces, while methanol and water are strongly hydrogen bonding liquids, with the important difference that in water the H-bonds form a space-filling, percolating network,^{64,65} while in methanol they do not.⁶⁶ The simulations have been performed at the temperature of 280 K, with the exception of water for which it has been 300 K. The rectangular basic simulation box has consisted of 4000 molecules in every case. The Y and Z edge lengths of the simulation box have been 50 Å, whereas the length of the X edge, being perpendicular to the macroscopic plane of the liquid surface, has been varied from 300 to 500 Å, depending on the density of the liquid, in order to provide a sufficiently wide vapor layer between the two liquid surfaces present in the basic box. Periodic boundary conditions have been applied in all directions. The CCl₄, acetone, and water molecules have been modeled by the OPLS,⁶⁷ TraPPE,⁶⁸ and SPC/E⁶⁹ potentials, respectively, whereas the acetonitrile and methanol molecules have been described by the potential models proposed by Böhm et al.⁷⁰ and by Walser et al.,⁷¹ respectively. Our previous study on the surface dynamics of water showed that the results are qualitatively insensitive to the particular choice of the potential model.⁶³ According to these potential models, the CH₃ groups of acetone and methanol have been treated as united atoms, whereas the H atoms of the CH₃ group of acetonitrile have been explicitly taken into account. The interaction parameters of the potential models used are collected in Table 1 of Ref. 36. All molecular models used are rigid; the internal geometry of the molecules has been kept fixed by means of the SHAKE algorithm.⁷² The intermolecular potential energy of the system has been calculated as the sum of the contributions of each molecule pairs, the interaction energy of a molecule pair being equal to the sum of the Coulomb and Lennard-Jones contributions of their respective interaction sites. The long range part of the electrostatic interaction has been accounted for using the Particle Mesh Ewald (PME) method in its smooth variant.⁷³

The simulations have been performed using the GROMACS 5.1 program package.⁷⁴ The equations of motion have been integrated in time steps of 1 fs. The temperature of the systems has been controlled using the Nosé-Hoover thermostat.^{75,76} The interfacial systems were created after equilibrating the liquid phase in a basic box the edges Y and Z of which being already set to a length of 50 Å. The X edge length corresponded to the bulk liquid density, and was subsequently increased the X edge length to its final value. The systems have been equilibrated for at least 5 ns, after which 2000 sample configurations per system, separated from each other by 1 ps long trajectories, have been dumped for the calculation of the diffusion coefficients and surface residence times. Further, 1000 sample configurations, separated by 10 fs long trajectories each, have been saved for evaluating the velocity autocorrelation functions. Finally, for methanol and water an additional set of 1000 sample configurations, now separated by 0.1 ps long trajectories, have been saved for the analysis of the hydrogen bond dynamics.

It should finally be noted that, for reference purposes, the bulk liquid phases of the systems considered have also been simulated, without the interface and the vapor phase, in exactly the same way as the corresponding interfacial systems, at the density equal to that of the bulk liquid phase of the corresponding interfacial system.

2.2. ITIM Analyses. In the ITIM analysis the detection of the full set of the truly interfacial molecules can be described as if one would move a spherical probe along test lines parallel with the macroscopic interface normal, starting from the bulk opposite phase, towards the liquid surface to be analyzed. Once the probe touches the first molecule of the phase of interest, the touched molecule is marked as being interfacial, and the algorithm continues with moving the probe along the next test line. The probe is regarded as being in contact with a given atom if the distance of their centers is equal to the sum of their radii. Having all test lines considered the full list of the truly interfacial molecules is detected.¹³

In this study a probe sphere of the radius of 2 Å has been used for CCl_4 , acetone, and acetonitrile, while that of 1.25 Å for methanol and water, in accordance with the results of previous studies concerning the dependence of the results on the probe size.^{13,27,77} The radius of the atoms has been defined as half of their Lennard-Jones distance diameter, σ . In determining whether a given molecule belongs to the liquid or to the vapor phase, a cluster analysis algorithm¹⁸ has been employed. Thus, two molecules have been defined as being in contact with each other if the distance between any of their atoms was smaller than a pre-defined cut-off value. Two molecules belong to the same cluster if they are connected through

a chain of contact molecules. The largest cluster found in the simulation box is regarded as the liquid phase itself, whereas the molecules belonging to any other cluster are regarded as being part of the vapor phase.^{18,33} The cut-off distance used in defining the contact position of the molecules has been set equal to the smallest of the minima positions of the atom-atom partial pair correlation functions, excluding the ones involving OH hydrogens. This way, the cut-off values of 8.0, 5.0, 3.6, 5.8, and 3.5 Å have been used for CCl₄, acetone, acetonitrile, methanol, and water, respectively. Test lines have been arranged in a 100 × 100 grid along the *YZ* plane (i.e., the macroscopic plane of the liquid surface), thus, two neighboring test lines have been separated by 0.5 Å from each other. Figure 1 shows an equilibrium snapshot of the surface region of the systems simulated, illustrating the surface layers of the liquid phases as determined by the ITIM method.

2.3. Calculation of the Mean Surface Residence Time. The survival probability of the molecules at the liquid surface, $L(t)$, can simply be defined as the probability that a molecule that belongs to the surface layer at time t_0 remains at the liquid surface until time $t_0 + t$. In order to distinguish between the cases when a molecule leaves the surface layer permanently, and when it only leaves it temporarily due to an oscillatory move, and returns to the surface immediately, a departure from the surface between t_0 and $t_0 + t$ is conventionally allowed if the molecule returns within a short time window of Δt . However, since the 1 ps length of the trajectories separating two subsequent sample configurations is already larger than/comparable with the time scale of these oscillations, here we have not allowed such departures of the molecules from the liquid surface; once a molecule has not been found in the surface layer it has been regarded as having left the surface. Since the departure of the molecules from the liquid surface is governed by first order processes, the $L(t)$ survival probability is of exponential decay, and, in the simplest case, it can be fitted by the function $\exp(-t/\tau_{\text{surf}})$, where τ_{surf} is the mean residence time of the molecules at the liquid surface. However, since some of the molecules leave and rejoin the surface layer due to a fast oscillatory move, the $L(t)$ data can be fitted by the sum of two exponentials, and has two characteristic time values, the first of which corresponds to this fast oscillation, while the second one to the permanent departure of the molecules from the surface.

2.4. Calculation of the Diffusion Coefficient and Characteristic Time of Surface Diffusion. The self-diffusion coefficient, D , of homogeneous, isotropic liquids can be estimated by comparing the second moment of the probability distribution function $P(\mathbf{r}, t; \mathbf{r}_0)$

of finding a molecule at time t at position \mathbf{r} , given that at $t = 0$ its position was \mathbf{r}_0 with that of the solution of the Fokker-Planck equation:

$$\frac{\partial}{\partial t} P(\mathbf{r}, t; \mathbf{r}_0) = D \nabla^2 P(\mathbf{r}, t; \mathbf{r}_0) . \quad (1)$$

At the practical level, this is usually done by sampling directly the second moment of the distribution, i.e., the mean square displacement of the molecules within the time t ,

$$MSD = \left\langle (\mathbf{r}_i(t_0 + t) - \mathbf{r}_i(t_0))^2 \right\rangle, \quad (2)$$

along the trajectory of the system simulated. In the above equation $\mathbf{r}_i(t_0)$ and $\mathbf{r}_i(t_0+t)$ stand for the position vectors of the i th molecule at time t_0 and $t_0 + t$, respectively; and the brackets $\langle \dots \rangle$ denote ensemble averaging. The solution of the Fokker-Planck equation in a homogeneous, isotropic system is

$$P(\mathbf{r}, t; \mathbf{r}_0) \propto \exp \left(- \frac{(\mathbf{r}_i(t_0 + t) - \mathbf{r}_i(t_0))^2}{kDt} \right), \quad (3)$$

and its second moment is simply $MSD = kDt$, where k is a parameter related to the dimensionality of the diffusive motion, its value being 2, 4, and 6 in the case of one-, two-, and three-dimensional diffusion, respectively. Therefore, the diffusion coefficient can simply be calculated through the Einstein relation:¹²

$$D = \frac{MSD}{kt}, \quad (4)$$

from the steepness of a straight line fitted to the MSD vs. t data. This fitting should, however, be done in a limited time range in order to ensure that the molecules exit the ballistic regime and lose correlation. In the present study, the time range above 2 ps has turned out to be sufficient for this purpose in every case. One should, of course, make sure that, in presence of periodic boundary conditions, the continuous trajectory of particles is reconstructed before calculating the MSD . It should also be noted that in calculating the diffusion coefficient of the molecules within the surface layer each molecule contributes to the MSD only in the time range it is part of the surface layer.

In confined systems, which lose both homogeneity and isotropy, there are several important changes to be taken into account in these equations, namely (i) a position-

dependent diffusion tensor $\mathbf{D}(\mathbf{r})$ in place of the diffusion coefficient, (ii) a full Fokker-Planck equation that includes the gradient of the position-dependent diffusion coefficient, and (iii) the fact that the solution for the probability distribution is not any more eq. 3, and, as a consequence, the *MSD* will also not have the simple form of kDt . Two examples for the application of this formalism are the investigation of the diffusion of water in proximity of a protein⁷⁸ and in the interstitial space between two periodic copies of a lipid bilayer.⁵⁶ Since here we are interested in the diffusion within the surface layer, and we update the statistics for the *MSD* only when a molecule is in that layer, the problem can be expressed in terms of an effective Fokker-Planck equation with reflecting boundary conditions along the layer normal, between $X = X_0$ and $X = X_0 + L_{\text{eff}}$, where L_{eff} is the effective width of the layer. In this case, the diffusion tensor takes the form $\mathbf{D} = \text{diag}(D_{\perp}, D_{\parallel}, D_{\parallel})$, where D_{\perp} and D_{\parallel} are the diffusion constants along the macroscopic surface normal axis, X , and within the macroscopic plane of the surface, YZ , respectively. Here we further assume that within the layer the diffusion tensor is not position dependent. The solution can therefore be expressed in terms of marginal probabilities to diffuse parallel to the layer, or perpendicular to it along X (still within the layer itself). The *MSD* for the lateral diffusion has still the Einstein form, $4D_{\parallel}t$, whereas the expression for the perpendicular *MSD*, once averaged over the initial position X_0 , is the series⁵⁶

$$\frac{1}{L_{\text{eff}}} \int \langle (X - X_0)^2 \rangle dX_0 = \frac{L_{\text{eff}}^2}{6} - L_{\text{eff}}^2 \sum \frac{16}{(2n+1)^4 \pi^4} \exp\left(-D_{\perp}t \left[\frac{(2n+1)\pi}{L_{\text{eff}}}\right]^2\right). \quad (5)$$

Due to the presence of boundaries, the asymptotic perpendicular (average) *MSD* is a constant (i.e., $L_{\text{eff}}^2/6$) rather than a linearly growing quantity, and the effective perpendicular diffusion coefficient, D_{\perp} , has to be estimated via a best fit of the sampled *MSD* to eq. 5. The series is quickly converging due to the presence of the $1/n^4$ term, and only few terms are needed to obtain an accurate approximation. It is interesting to note that, since both the series in eq. 5 and the Taylor expansion of the exponential function are absolutely convergent, it is possible to exchange the two sums and obtain, for small times, that the (average) perpendicular *MSD* is $2D_{\perp}t$. This is seemingly recovering the result of the Einstein equation. However, this approximation is correct only for times small enough for the diffusing particles not to feel the presence of the boundaries.

The characteristic time of the diffusion can be defined as the time after which the positions visited by a molecule follow a Gaussian distribution with the width of L_m , $\sqrt{A_m}$, and $\sqrt[3]{V_m}$ in the case of one-, two-, and three-dimensional diffusion, respectively, given that the diffusive motion can indeed be regarded as a random walk (i.e., it is not biased by any external force).^{63,79} Here L_m , A_m , and V_m , stand for the section, area, and volume per molecule in the one-, two-, and three-dimensional cases, respectively. Thus, the characteristic time of the (two dimensional) diffusion of the molecules within the surface layer of the liquid phase, τ_D , can simply be given as

$$\tau_D = \frac{A_m}{4D_{||}}, \quad (6)$$

where

$$A_m = \frac{2YZ}{\langle N_{\text{surf}} \rangle}, \quad (7)$$

$\langle N_{\text{surf}} \rangle$ stands for the average number of the surface molecules in the system; and the factor 2 in the numerator of eq. 7 accounts for the two liquid surfaces present in the basic box. It is important to point out that the parallel and perpendicular diffusion coefficients in the first molecular layer are still calculated in the global reference frame, and not along the local tangent plane or normal direction to the curved interface. Beside the added complexity of projecting the motion on the local reference frame, this approach raises a conceptual problem related to the fact that the surface is changing in time, and it would be probably difficult to avoid an ambiguous definition of the distance travelled along the surface itself.

2.5. Calculation of the Velocity Autocorrelation Function. The autocorrelation function of the molecular center of mass velocity, \mathbf{v}^{cm} , defined as

$$\Psi(t) = \frac{1}{N} \left\langle \sum_i \mathbf{v}_i^{\text{cm}}(t_0 + t) \mathbf{v}_i^{\text{cm}}(t_0) \right\rangle, \quad (8)$$

where N is the total number of molecules, is a useful tool for understanding the dynamical behavior of single molecules, providing information on which time scales the memory of the initial velocity of a particle is lost due to interaction with neighboring molecules. The typical traits of the velocity autocorrelation functions in dense fluids are an initial parabolic decay,

related to the average force acting on the molecule, followed by a steep decay imposed by collisions with nearest neighbors. At relatively high densities, the velocity autocorrelation function can become negative because of strong repulsion from the cage of neighboring molecules, and its long time behavior can be characterized by hydrodynamics in the form of an algebraic decay to zero. The velocity autocorrelation function carries similar information on the dynamics of the molecules as the *MSD*. In practical terms, however, the short-time dynamics is more easily accessible from the velocity autocorrelation function, and for this reason we introduce a velocity autocorrelation function, $\Psi_{\parallel}(t)$, which is the analogue of the *MSD* of the first layer:

$$\Psi_{\parallel}(t) = \left\langle \frac{1}{N(t_0)} \sum_{i=1}^{N(t_0)} \mathbf{v}_{\parallel i}^{\text{cm}}(t_0 + t) \mathbf{v}_{\parallel i}^{\text{cm}}(t_0) \theta_i(t_0 + t, t_0) \right\rangle, \quad (9)$$

where $N(t)$ is the number of molecules in the first layer at time t , and the function $\theta_i(t_2, t_1)$ is equal to 1 if molecule i has been residing continuously in the first layer from time t_1 to t_2 , and zero otherwise.

3. Results and Discussion

The profiles of the molecular number density, ρ , of the five systems simulated along the macroscopic surface normal axis, X , are shown in Figure 2, along with those of the first molecular layer at the liquid surface. The different positions of the surface region along the X axis simply reflect the different sizes of the molecules and the different densities of the liquid phases considered. As is clearly seen, the X range of the surface layer largely overlaps with the constant density region of the system, while the intermediate density part of the overall profile is far from being fully accounted for by the contribution of the surface layer in every case. In other words, the definition of the surface region of the systems in the usual, non-intrinsic way as the X range of intermediate density would indeed cause an erroneous identification of a surprisingly large set of molecules, both interfacial and non-interfacial ones, and ultimately lead to the analysis of an *ad hoc* set of molecules rather than that of the real, capillary wave corrugated, intrinsic liquid surface layer.^{13,14} It is also seen that, as it is expected,⁸⁰ the density profile of the surface layer is of Gaussian shape in every case. (The Gaussian functions fitted to these distributions are also shown in Fig. 2.) The width parameter

of the Gaussian function fitted to such a density profile, δ , can serve as a measure of the width of the surface layer.

The $L(t)$ survival probabilities of the molecules at the liquid surface are shown in Figure 3. In the following, this function is used to define the set of the longest residing 10% of the surface molecules in every sampled configuration, in order to analyze to what extent the properties of these long residing molecules differ from those of the entire set of the surface molecules. We could fit the $L(t)$ data well with the sum of two exponential functions in every case, as shown also in the figure. The characteristic times of these two processes are collected in Table 1. The shorter of these two characteristic times never exceeds 2.5 ps, indicating that the corresponding process is probably related to the fast librational motion of the molecules. This process usually does not lead to the permanent departure of the molecules from the surface layer; instead, they only leave the surface layer due to this librational oscillation, but come back shortly thereafter due to the same mechanism. On the other hand, the second process corresponds to the real departure of the molecules from the surface layer. The characteristic time of this second process, τ_{surf} , falls in the range of about 15-25 ps, being the largest for CCl_4 , and being rather similar in the other four systems considered. The value of τ_{surf} sets the time scale of all molecular processes occurring in the surface layer of the corresponding liquid phase.

3.1. Surface Diffusion. The perpendicular and parallel (relative to the macroscopic surface plane) *MSDs* of the molecules within the surface layer are shown as a function of time in Figure 4. For comparisons, the full, three dimensional *MSDs*, obtained in the corresponding bulk liquid phases, are shown in the inset of Fig. 4. The D_{\perp} and D_{\parallel} diffusion coefficient values corresponding to all surface molecules as well as to the longest residing 10% of them, and also the bulk phase diffusion coefficients, obtained from the best fits of eqs. 5 (D_{\perp}) and 4 (D_{\parallel} and bulk phase D) are collected in Table 2. Further, the characteristic times of the parallel diffusion within the surface layer, τ_{D} , obtained through eq. 6, are included in Table 1.

As is seen, the characteristic diffusion time, τ_{D} , is considerably smaller (i.e., by a factor of 3-5) than the mean surface residence time, τ_{surf} , indicating that the surface diffusion of the molecules can indeed be meaningfully discussed, as it occurs well within the time scale of the molecules remaining part of the surface layer. This finding is illustrated in Figure 5, showing the trajectory, projected to the macroscopic surface plane, YZ , both of a surface molecule that is among the longest residing 10%, and also that of one having a surface

residence time close to the average value for all the five systems simulated. The $\tau_{\text{surf}}/\tau_{\text{D}}$ ratio is the largest for the strongly dipolar but aprotic molecules, which can diffuse faster than the hydrogen bonding ones, as their diffusion is not hindered by the H-bonds formed with their neighbors (the dipole moment of the molecular models used are also collected in Table 1). This ratio, on the other hand, is as small for CCl_4 as for methanol and water, primarily due to the large characteristic time of its surface diffusion. The finding that the $\tau_{\text{surf}}/\tau_{\text{D}}$ ratio decreases, in general, with decreasing dipole moment is in clear accordance also with the earlier finding of Duque et al. that this ratio is around 1 for the totally apolar Lennard-Jones system.⁶⁰ It might seem surprising that, contrary to Duque et al., we obtained a considerably larger τ_{surf} than τ_{D} value for the apolar CCl_4 molecules. However, it should be emphasized that although the CCl_4 molecules do not have a net dipole moment, their atoms still bear (at least, in the molecular model used) non-negligible fractional charges, and hence they, unlike the Lennard-Jones spheres, still interact via a considerable multipolar interaction.

As is seen from Table 2, the surface residence time of the individual molecules is not correlated to their surface mobility, as the calculation of D_{\perp} and D_{\parallel} for all the surface molecules or for only the longest residing 10% of them results in very similar values. Further, it is also found that the molecules diffuse considerably faster at the liquid surface, both in the parallel and perpendicular directions, than inside their bulk liquid phase. Similar relation was found earlier by Duque et al. for the diffusion of the Lennard-Jones system.⁶⁰ This is understandable in the light of the fact that at the liquid surface the molecules lose a part of their attractive interactions with respect to the bulk liquid phase. It can also be well understood that the ratio of the surface and bulk diffusion coefficients is the smallest in water, since it is known that water molecules adopt such orientations at the liquid surface that they can preserve about 75% of their hydrogen bonds as compared to the bulk liquid phase.^{13,81} On the other hand, it is somewhat surprising that this ratio is considerably larger for methanol than water, considering that methanol molecules can be aligned at the surface in such a way that they preserve all of their hydrogen bonds. The reason for this enhanced surface diffusion for methanol could be related to the hindrance of the mobility of the bulky methyl groups inside the liquid phase due to their accumulation around each other.⁸²⁻⁸⁴ This hindrance can be dramatically reduced at the liquid surface by the very strong preference of the molecules for sticking their methyl groups straight out to the vapor phase.¹⁵

Besides the D_{\perp} value itself, the fitting of the perpendicular *MSD* data by eq. 5 also yields the effective width of the surface layer, L_{eff} . The values of L_{eff} are collected in Table 3,

along with the width parameter of the surface layer density profiles, δ , as obtained for the five liquids considered. As is seen, these values indeed correlate well with each other, their ratio falling between about 1.4 and 1.8 in every case. Integration of the Gaussian-shape density profile of the surface molecules (Fig. 2) in the distance range of the width L_{eff} around its center reveals that L_{eff} is representative of an effective width that includes 83-92% of the surface molecules for the different system, as detailed in Table 3.

Figure 6 a and b show the *MSD* of the surface molecules along the macroscopic surface normal axis, X , as a function of time on two different time scales, normalized by the mean surface residence time, τ_{surf} , and by the characteristic time of surface diffusion, τ_{D} , respectively. The obtained *MSD* deviates downwards from linearity not only on the real time scale up to 25 ps, but also on the scale of the surface residence time of the molecules in every case. More precisely, the simulated data points start deviating from linearity at around 20-40% of τ_{surf} . This finding demonstrates that although the molecules can seemingly freely diffuse also along the macroscopic surface normal in a non-negligible fraction of their surface lifetime, they start feeling the presence of the boundaries still well within their lifetime at the liquid surface, τ_{surf} . On the other hand, as seen from Fig. 6.b, the *MSDs* are indeed linear up to τ_{D} , i.e., within the characteristic time scale of the lateral surface diffusion. To interpret this finding, however, we have to emphasize that τ_{D} is the upper limit of the time range within which the molecules can still have memory of their initial position (i.e., they might still not exhibit an uncorrelated random walk). The observed linearity of the perpendicular *MSD* can thus either be related to the fact that, within this time scale, the molecules do not feel yet the constraint of being in the surface layer and diffuse freely along the macroscopic surface normal, or it can also be an artifact of the limited time window. Further investigation of the possible physical meaning of the observed linearity of *MSD* within this time scale can be done by analyzing the velocity autocorrelation function of the surface molecules, which is presented in a subsequent sub-section.

3.2. Spatial Correlation between Long-Residing Surface Molecules. The diffusion of the molecules that stay at the liquid surface for unusually long times did not turn out to be markedly different from that of the other surface molecules in any case. To further investigate whether long surface residence times of certain molecules simply occur randomly, or they are related to certain properties of these molecules, we investigate how strongly the positions of these molecules are correlated with each other at the liquid surface. In other words, we

address the question whether long-residing surface molecules are distributed randomly at the liquid surface, or they form relatively dense patches, leaving large empty spaces between them. For this purpose, we have projected the centers of the longest residing 10% of the surface molecules to the macroscopic plane of the surface, YZ , and have calculated the Voronoi cells⁸⁵⁻⁸⁷ around each of these projections. If these projections are randomly distributed, the area (A) of the Voronoi cells is expected to follow approximately a gamma distribution^{88,89}

$$P(A) = a A^{\nu-1} \exp(-\nu\rho A) \quad (10)$$

where ν and ρ are free parameters, while a is a normalization factor. On the other hand, in the case of correlated arrangement of these projections the $P(A)$ distribution deviates from eq. 10, exhibiting a long tail of exponential decay at the large area side of its peak.⁹⁰

The $P(A)$ Voronoi cell area distributions are shown in Figure 7 as obtained in the five systems simulated, together with their best fits by eq. 10. The exponential decay of all these data sets (transformed to a linear decrease by the use of a logarithmic scale) as well as the deviation from eq. 10 is clearly seen from the figure in every case. This finding indicates that the long-residing molecules are distributed in a correlated way at the liquid surface, i.e., they prefer to stay in the vicinity of each other. It is also apparent that this correlation is the weakest for the hydrogen bonding liquids, in particular, for water, and strongest for CCl_4 . The observed correlated arrangement of the long residing surface molecules at the liquid surface is illustrated also in Figure 8, showing the projections of the centers of these molecules to the macroscopic plane of the surface, YZ , in an equilibrium snapshot of both the CCl_4 and the water system.

3.3. Hydrogen Bonding at the Intrinsic Liquid Surface. In this sub-section we address the point how the properties of the hydrogen bonds are affected by the liquid surface in the two H-bonding liquids considered, i.e., methanol and water. Also, to further study the question how unusually long surface residence time is related to other properties of the molecules, we compare the properties of the H-bonds of the longest residing 10% of the surface molecules with those of all surface molecules.

The average lifetime of a hydrogen bond can be defined in a similar way as the mean surface residence time. Thus, the survival probability of a H-bond, $L_{\text{HB}}(t)$, is the probability that a H-bond existing at time t_0 will persist up to the time t_0+t . Again, the breaking of a H-

bond is a process of first order kinetics, hence, $L_{\text{HB}}(t)$ is a function of exponential decay. Therefore, the mean H-bond lifetime, τ_{HB} , can simply be estimated by fitting the function $\exp(-t/\tau_{\text{HB}})$ to the simulated $L_{\text{HB}}(t)$ data. Similarly to the survival probability at the liquid surface, $L(t)$, the short time part of $L_{\text{HB}}(t)$ can also deviate from the exponential decay; this transient part of the $L_{\text{HB}}(t)$ data, covering the first 0.1-0.5 ps of the time range, has thus been left out from the exponential fit (see Figure 9). The τ_{HB} values corresponding to the H-bond between two surface molecules, included also in Table 1, are typically an order of magnitude smaller for both H-bonding liquids considered than the mean surface residence time of the molecules. Therefore, the H-bonds formed specifically by surface molecules can be distinguished from those involving also bulk phase molecules, and thus their properties can indeed be meaningfully discussed. It should be noted that the average lifetime of a H-bond at the liquid surface is considerably, i.e., 25-40%, shorter than in the bulk liquid phase for both H-bonding liquids considered: the τ_{HB} values obtained in the bulk liquid phase of methanol and water have turned out to be 5.22 ps and 2.01 ps (the corresponding surface values being 3.22 ps and 1.54 ps), respectively. On the other hand, the surface residence time of the molecules is not related to the lifetime of their H-bonds, as the τ_{HB} values corresponding to the H-bonds between two long-residing surface molecules are 3.28 ps in methanol and 1.52 ps in water.

The average number of the H-bonds formed by a surface molecule, $\langle n_{\text{HB}} \rangle$, as well as the average interaction energy of such a H-bonded molecule pair, $\langle U_{\text{HB}}^{\text{pair}} \rangle$, are collected and compared to the respective bulk phase values in Table 4. Values corresponding specifically to the longest residing 10% of the surface molecules are also included in the table. Furthermore, the $\langle n_{\text{HB}} \rangle$ and $\langle U_{\text{HB}}^{\text{pair}} \rangle$ values corresponding to the interfacial molecules are also decomposed according to the location of the H-bonding partner molecule (i.e., whether it also belongs to the surface layer or not). As is clear from the data, the liquid surface does not have a considerable influence on the H-bonding structure of the molecules in methanol. The average number of the H-bonded neighbors of a surface methanol molecule is only 3% less than that of a bulk phase one and, correspondingly, the interaction energy of such a molecule pair agrees within 0.3 kJ/mol for molecule pairs being at the liquid surface and in the bulk liquid phase. By contrast, interfacial water molecules have, on average, about 15% less H-bonded neighbors than the bulk phase ones, while their pair interaction energy is, on average, 0.7 kJ/mol deeper than in the bulk liquid phase. This difference can be related to the preferred

surface orientations of these molecules. Namely, both of these molecules can easily be oriented at the liquid surface in such a way that three of their tetrahedrally aligned H-bonding (i.e., O-H and lone pair) directions are turned flatly towards the bulk liquid phase.¹³⁻¹⁵ Since methanol molecules have only three H-bonding directions, they can efficiently maintain all of their H-bonds even at the macroscopically flat liquid surface by sticking the fourth of the tetrahedrally aligned electron pairs of their O atom (i.e., the O-CH₃ bond) straight out to the vapor phase.¹⁵ On the other hand, in water this fourth electron pair around the O atom also represents a H-bonding direction. Therefore, alignments of the surface water molecules in which three of the H-bonding directions are turned flatly inward involves the “sacrifice” of the fourth of these directions, which is then turned straight towards the vapor phase.^{13,14} All four H-bonding directions can only be turned towards the bulk liquid phase at strongly curved portions of the liquid surface,^{13,14,91} such as at the tips of the ripples of the molecularly wavy surface.

The energy loss corresponding to the fewer number of H-bonding neighbors is partly compensated by a certain ordering of the H-bonding arrangement of the water molecules at the liquid surface, which results, on average, in somewhat stronger H-bonds at the surface than in the bulk phase (see Table 4). The observed small, about 4% strengthening of the H-bonds at the surface of water is also in accordance with earlier results.^{82,92,93} It is interesting to note that although the bulk phase H-bonds are, on average, slightly weaker in water, and are about the same strength in methanol than the interfacial ones, H-bonds live considerably longer in the bulk phase than at the interface of both liquids. This finding is again in accordance with earlier claims that the strength and lifetime of the H-bonds are independent from each other.^{93,94} Instead of their strength, the shorter lifetime of the surface H-bonds can be explained by the enhanced mobility of the surface molecules, as compared to that of the bulk phase ones (see Table 2), due to their lack of attractive interactions at the vapor side of the interface.

When comparing the properties of the long-residing molecules with those of all the surface molecules, it is seen that long residing molecules form, on average, slightly, by 3-4% less H-bonds than all the surface molecules. When decomposing these numbers into the values corresponding to in-layer and off-layer H-bonds, it turns out that the average number of H-bonding neighbors of the long residing surface molecules within the surface layer is somewhat (i.e., by 11% in methanol and 3% in water) larger, while that of their non-surface H-bonding neighbors is considerably (i.e., 38% in methanol and 25% in water) smaller than the values corresponding to all surface molecules. The observed increase of the number of in-

layer H-bonds is in accordance with our previous finding that long residing molecules prefer to stay in the vicinity of each other. However, the most striking feature of the long residing surface molecules is clearly that they form much less hydrogen bonds with the subsurface molecules than the value corresponding to all of the surface molecules. This fact can also explain their long stay in the surface layer. Namely, having less off-layer H-bonded neighbors, these molecules are better separated from the subsurface region, and hence can not leave the surface as easily as the other ones.

3.4. Velocity Autocorrelation Function at the Intrinsic Liquid Surface. In Figure 10 we report the autocorrelation function of the in-plane molecular center of mass velocity for the molecules belonging to the first layer, $\Psi_{\parallel}(t)$, and, for comparison, also the autocorrelation function $\Psi(t)$ of the molecular center of mass velocity in the corresponding bulk liquid phases. The common trait, shared by all systems, is that the in-plane velocity of surface molecules is always more correlated during the initial, rapid decay, which takes place within the first 0.1-0.3 ps. Of all considered liquids, only CCl_4 and acetone show, in the bulk, no presence of the cage effect, and $\Psi(t)$ is always positive, whereas $\Psi_{\parallel}(t)$ is considerably larger at all times, with values clearly different from zero, also for time lags where $\Psi(t)$ has already vanished. In the case of acetonitrile, the two autocorrelation functions are different both qualitatively and quantitatively from each other, as the negative part of $\Psi(t)$ is not present any more in $\Psi_{\parallel}(t)$. The latter function decays smoothly, resembling a memoryless process. In both methanol and water the two autocorrelation functions share some common features, namely an oscillation at 0.25 and 0.13 ps, respectively, which is the signature of hydrogen bonding.⁹⁵ In methanol, however, the in-plane correlation function of the surface molecules is, again, always positive, and the cage effect, which characterizes the bulk phase dynamics, is not present within the surface layer. While methanol molecules retain the majority of their hydrogen bonds at the liquid surface, the outward pointing arrangement of the CH_3 groups at the surface results in a much less crowded environment of the molecules. As a consequence, the cage effect disappears, in accordance with the strongly enhanced surface diffusion discussed previously. Water is the only case in which the in-plane correlation of the surface molecules becomes negative, showing that the hydrogen bond network is strong enough to influence the dynamics of the water molecules even within the surface layer. Our results suggest that water behaves in a rather unique way in this respect, as no such behavior is seen for the other liquids considered. Still, the in-plane velocity correlation of the surface

molecules is always larger than its bulk counterpart in the region of positive values, and smaller in that of negative ones, showing therefore a larger mobility of the molecules, and a less pronounced cage effect, which again explains the larger diffusion coefficient in the surface, with respect to that in the bulk. This effect is, however, less pronounced here than in the case of methanol, where caging is completely eliminated at the liquid surface.

4. Summary and Conclusions

In this paper, we have analyzed the single particle dynamical properties of the molecules in the first molecular layer of five molecular liquids, ranging from apolar through aprotic dipolar to hydrogen bonding ones. Such an analysis is clearly enabled by performing an intrinsic analysis of the liquid surface. The analysis of the molecular residence times in the first layer has shown that the dynamics of escape from the first layer is dictated by two characteristic time scales. The first, fast process of escape takes place on the time scale of about 2 ps for all liquids considered here, and is most likely representative of molecules leaving the layer for short times due to librational motions. The other process dominates after the first few picoseconds and takes place on the much longer scale of 15-25 ps, and is found to be considerably larger than the characteristic time of in-layer diffusion and hydrogen bond lifetime (for methanol and water), which are therefore meaningful observables for this set of molecular liquids and thermodynamic points. We investigated the diffusion in the first layer by sampling both the mean square displacement and the velocity autocorrelation function of the molecular centers of mass. The mean square displacement parallel to the macroscopic plane of the interface and perpendicular to it shows two qualitatively different behaviors, namely, the common Einstein linear dependence of bulk systems (for the parallel diffusion), and saturation behavior that fits extremely well with the diffusion between two reflecting walls.^{56,78} The diffusion coefficients estimated from these two separate fits are markedly different from the diffusion coefficient obtained in the bulk liquid phase, showing, in all cases, a much (i.e., typically 3-4 times) larger surface diffusion with respect to the bulk. The analysis of the in-plane velocity autocorrelation function confirmed this finding, showing that, at the picosecond scale, molecules at the surface are in all cases more free to move than in the bulk. At the surface, excluding the case of water, no trace of the cage effect is found, if this was present in the bulk. In those cases, which did not present a cage effect even in the bulk liquid phase (i.e., CCl₄ and acetone), sizeable correlation with the initial velocity is found at time lags, where the bulk counterpart shows no correlation any more. The analysis of the

spatial distribution of the long-residing surface molecules has revealed that they are clearly characterized by some degree of clustering. The analysis of the hydrogen bonded neighbors has shown that, in contrast to water, methanol is retaining practically all of its hydrogen bonded neighbors at the liquid surface. This result might be surprising in the light of the much higher diffusion coefficient of the methanol molecules found at the liquid surface than in the bulk liquid phase, however, it is consistent with the pronounced tendency of methanol to expose the bulky CH_3 group to the vapor side of the interface,¹⁵ which also helps eliminating the cage effect. This shows that the main factor in building up the internal friction for bulk methanol is, in fact, presence of the CH_3 groups rather than that of the hydrogen bonds (which, unlike in liquid water, do not form a percolating network in bulk methanol⁶⁶). The opposite happens in water, where the dynamics of the molecules is almost completely dominated by the hydrogen bond networking, both in the bulk liquid phase and at its surface, resulting also in its very high surface tension, with respect to all other molecular liquids considered here.

In conclusion, the analysis of single particle dynamical properties at the intrinsic liquid surface has proven to be very informative on the microscopic dynamics at liquid/vapor interfaces, showing that mass transport properties are markedly different at the surface, with respect to the bulk. In fact, these difference are surprising, considering the fact that, from the structural point of view, the first molecular layer is not so much different (e.g., in terms of density or hydrogen bonded neighbors) from the subsequent ones. The two- to fourfold increase in mobility at the surface draws a picture of a much more fluid surface layer, sharing some traits with those of rarefied fluids in case of non-hydrogen bonding liquids, which can have important implication for diffusion-limited reactions occurring at interfaces.

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Tables

TABLE 1. Characteristic Times of Various Molecular Processes Occurring in the Surface Layer of the Liquids Studied (in ps Units). Values in Parenthesis Correspond to the Initial, Fastly Decaying Process of Leaving the Liquid Surface. Error Bars Are Always Below 1%. The Dipole Moment of the Molecular Models Used (μ) Is Also Included in the Table.

system	τ_{surf}	τ_{D}	τ_{HB}	μ/D
CCl ₄	26.2 (2.5)	7.20	-	0.00
Acetone	16.1 (2.0)	2.97	-	2.50
Acetonitrile	14.5 (1.8)	3.39	-	4.14
Methanol	16.4 (2.0)	4.34	2.27	2.28
Water	15.0 (1.7)	4.11	1.36	2.35

TABLE 2. Diffusion Coefficients within the Surface Layer (in $\text{\AA}^2/\text{ps}$ Units), Both Along with and Perpendicular to the Macroscopic Plane of the Surface, and Inside the Bulk Liquid Phase of the Systems Studied. Values in Parenthesis Correspond to the Longest Residing 10% of the Surface Molecules. Error bars Are Always Below 3%.

		CCl ₄	Acetone	Acetonitrile	Methanol	Water
Surface	D_{\perp}	0.70 (0.73)	1.52 (1.66)	1.09 (1.18)	0.74 (0.76)	0.51 (0.51)
	D_{\parallel}	0.99 (1.03)	2.04 (2.19)	1.46 (1.56)	0.74 (0.76)	0.52 (0.53)
Bulk	D	0.24	0.55	0.42	0.28	0.27

TABLE 3. Parameters Describing the Width of the Surface Layer in the Different Systems. x_{eff} is the Fraction of Surface Molecules Within L_{eff} (See the Text).

	CCl ₄	Acetone	Acetonitrile	Methanol	Water
$L_{\text{eff}}/\text{\AA}$	11.5	11.8	8.8	8.4	5.6
$\delta\text{\AA}$	6.5	6.6	5.6	6.2	3.7
x_{eff}	0.92	0.93	0.89	0.83	0.88

TABLE 4. Average Number of Hydrogen Bonded Neighbors Around, and Average Interaction Energy of a Hydrogen Bonded Molecule Pair Involving a Surface Molecule, a Long-Residing Surface Molecule, and a Bulk Phase Molecule in Methanol and Water. Error Bars for $\langle n_{\text{HB}} \rangle$ and $\langle U_{\text{HB}}^{\text{pair}} \rangle$ Are Always Below 0.1% and 1%, Respectively.

system		$\langle n_{\text{HB}} \rangle$			$\langle U_{\text{HB}}^{\text{pair}} \rangle / \text{kJ mol}^{-1}$		
		total	in-layer	off-layer	total	in-layer	off-layer
methanol	interfacial	1.86	1.34	0.52	-20.58	-20.75	-20.16
	long-residing	1.81	1.49	0.32	-20.68	-20.87	-19.97
	interfacial	1.92	-	-	-20.30	-	-
water	bulk	3.30	2.42	0.88	-19.22	-19.60	-18.17
	interfacial	3.17	2.50	0.67	-19.34	-19.74	-17.84
	long-residing	3.79	-	-	-18.53	-	-

Figure legend

Figure 1. Equilibrium snapshot of the surface portion of the five systems simulated. Molecules belonging to the surface layer are shown enlarged. C, Cl, O, N, and H atoms are marked by light blue, green, red, dark blue, and white color, respectively.

Figure 2. Molecular number density profile of the five systems simulated (dashed lines) and those of their surface layer (open circles) along the macroscopic surface normal axis, X . The Gaussian functions fitted to the surface layer profiles are shown by solid lines. All profiles shown are symmetrized over the two liquid-vapor interfaces present in the basic box. CCl_4 : black, acetone: red, acetonitrile: orange, methanol: blue, water: green.

Figure 3. Survival probability of the molecules within the surface layer of their liquid phase, shown on a semi-logarithmic scale, as obtained in the five systems simulated (full circles). The sums of the two exponentially decaying functions, fitted to these data sets, are shown by solid lines. Color coding of the systems is the same as in Fig. 2.

Figure 4. Mean square displacements of the surface molecules along the macroscopic surface normal axis, X (top panel), and within the macroscopic plane of the surface, YZ (bottom panel) as a function of time, as obtained in the five systems simulated. The inset shows the MSD vs. t data obtained in the bulk liquid phase of these liquids. Fits of the one dimensional MSD vs. t data by eq 5 as well as linear fits of the two and three dimensional data are shown by solid lines. Color coding of the systems is the same as in Fig. 2.

Figure 5. Trajectory of a surface molecule that belongs to the longest residing 10% at the liquid surface (open circles), and of a surface molecule the surface residence time of which roughly equals to its mean value (full circles), projected to the macroscopic plane of the surface, YZ , as taken out from all five systems simulated. Color coding of the systems is the same as in Fig. 2.

Figure 6. One dimensional mean square displacement of the molecules belonging to the surface layer along the macroscopic surface normal axis, X , as obtained in the five systems simulated, shown on the time scales corresponding to the characteristic times of (a) the surface residence time, and (b) the lateral diffusion of the surface molecules. Color coding of the systems is the same as in Fig. 2.

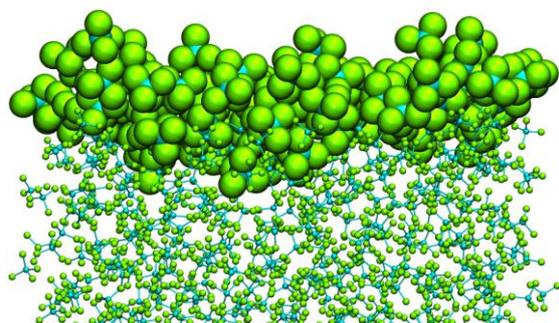
Figure 7. Distribution of the area of the Voronoi cells of the projections of the surface molecules onto the macroscopic plane of the surface, YZ as obtained in the five systems simulated. To emphasize the exponential decay of the distributions at large area values, the data are shown on a semi-logarithmic scale. Best fits of the data by eq. 10 are shown by solid lines. Color coding of the systems is the same as in Fig. 2.

Figure 8. Equilibrium snapshot of the liquid surface of CCl_4 (left) and water (right) from the top view, showing the projection of the centers of the longest residing 10% of the surface molecules to the macroscopic plane of the surface, YZ .

Figure 9. Survival probability of the hydrogen bonds formed by two molecules at the liquid surface (full circles), two long-residing surface molecules (open circles), and two molecules in the bulk liquid phase (asterisks), as obtained in methanol (blue) and water (green). To emphasize their exponential decay, the data are shown on a semi-logarithmic scale. The exponentially decaying functions fitted to the long time part of the data are shown by solid lines.

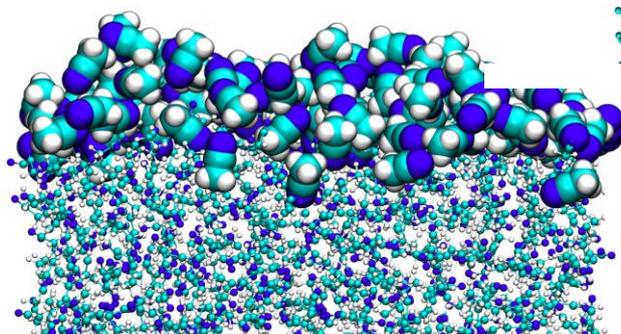
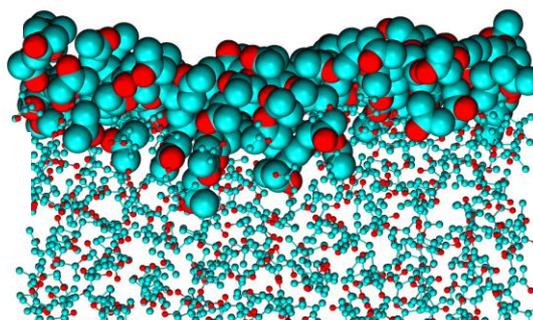
Figure 10. Normalized velocity autocorrelation function of the centers of mass of the surface molecules in the macroscopic plane of the surface, YZ (solid lines), and that of the three dimensional velocity of the molecules in the bulk liquid phase (dashed lines) of the five molecular systems studied. The insets show the relevant part of the functions on an enlarged scale. Color coding of the systems is the same as in Fig. 2.

Figure 1.
Fábián et al.



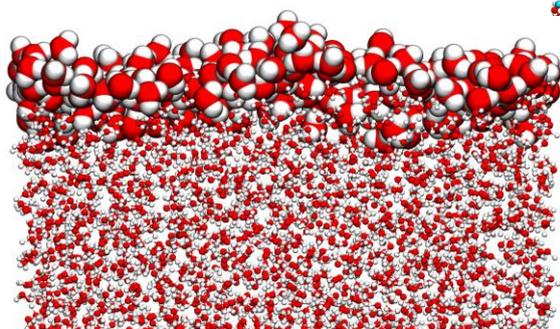
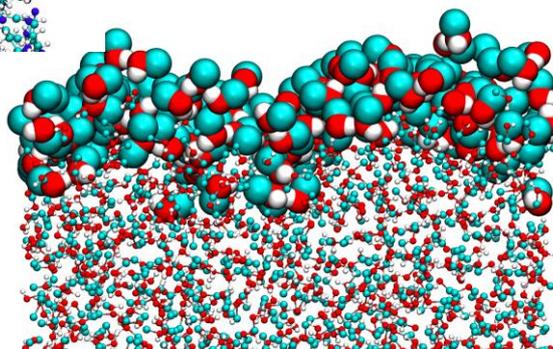
CCl₄

Acetone



Acetonitrile

Methanol



Water

Figure 2.
Fábián et al.

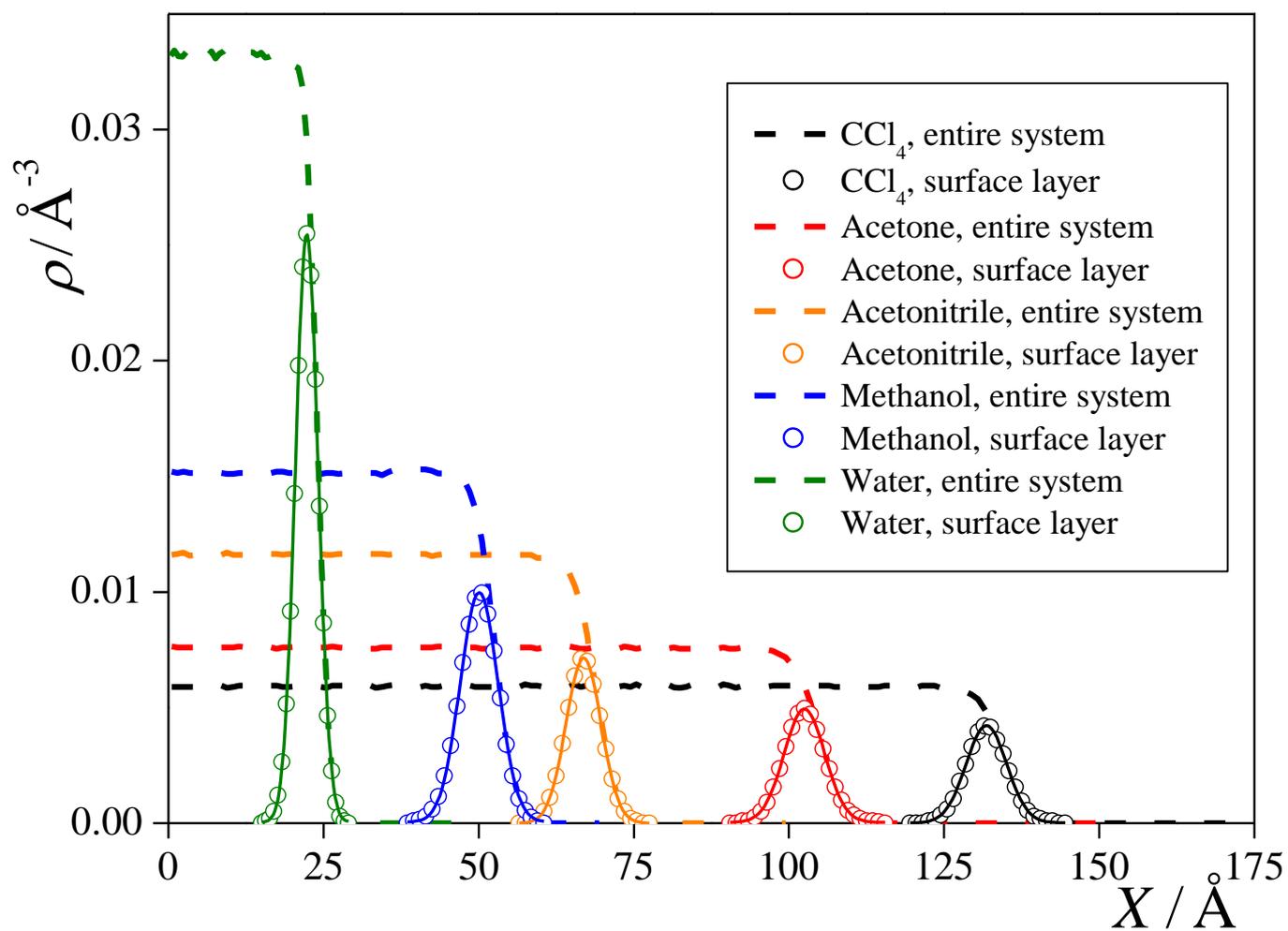


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Fabián et al.

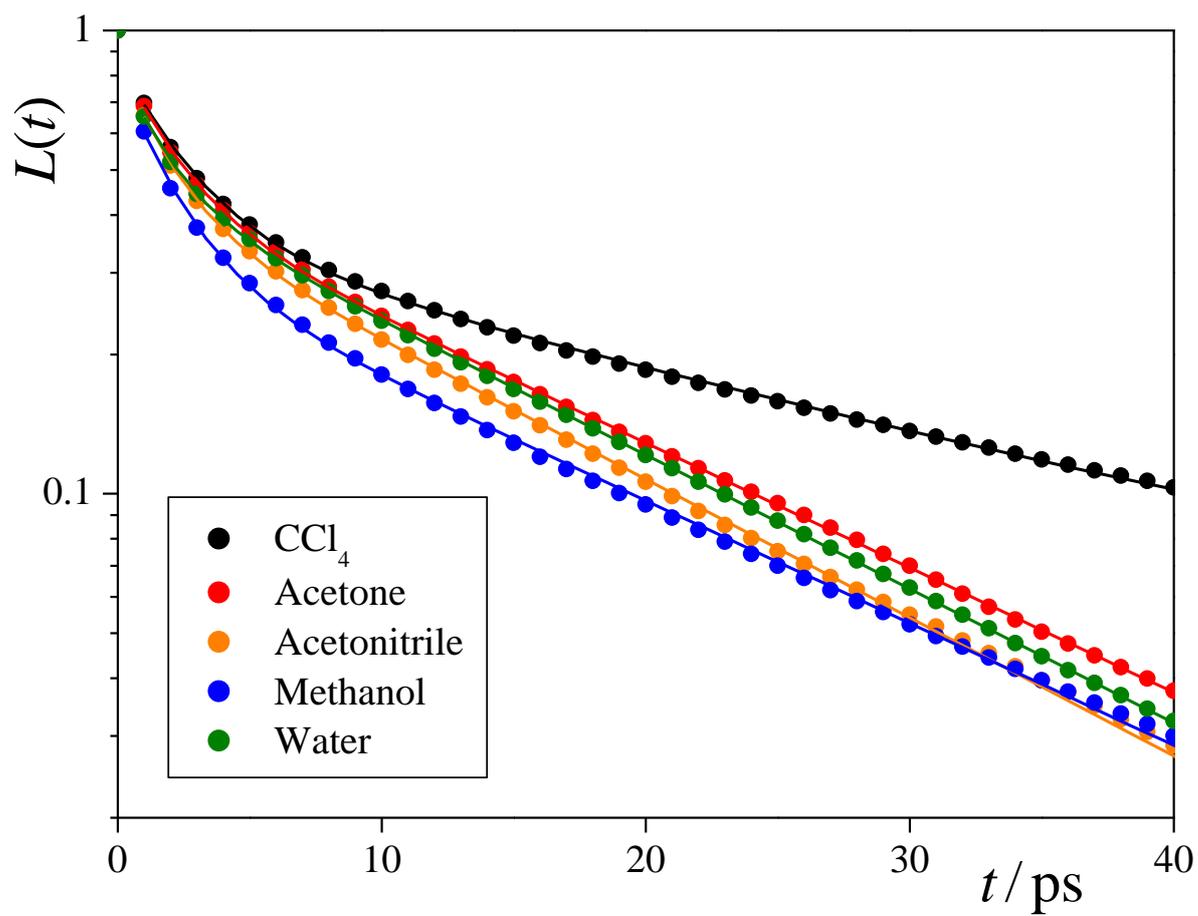


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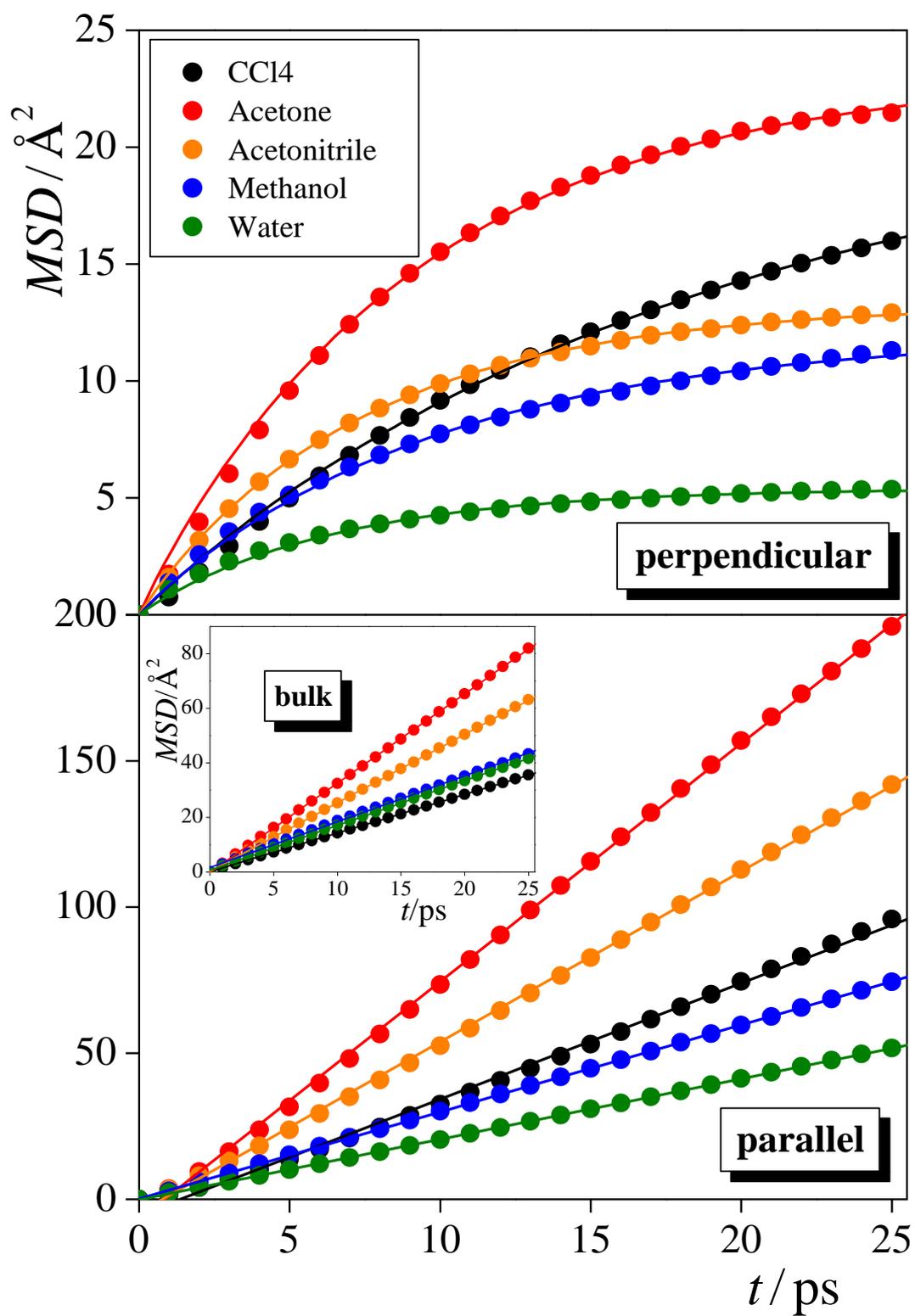


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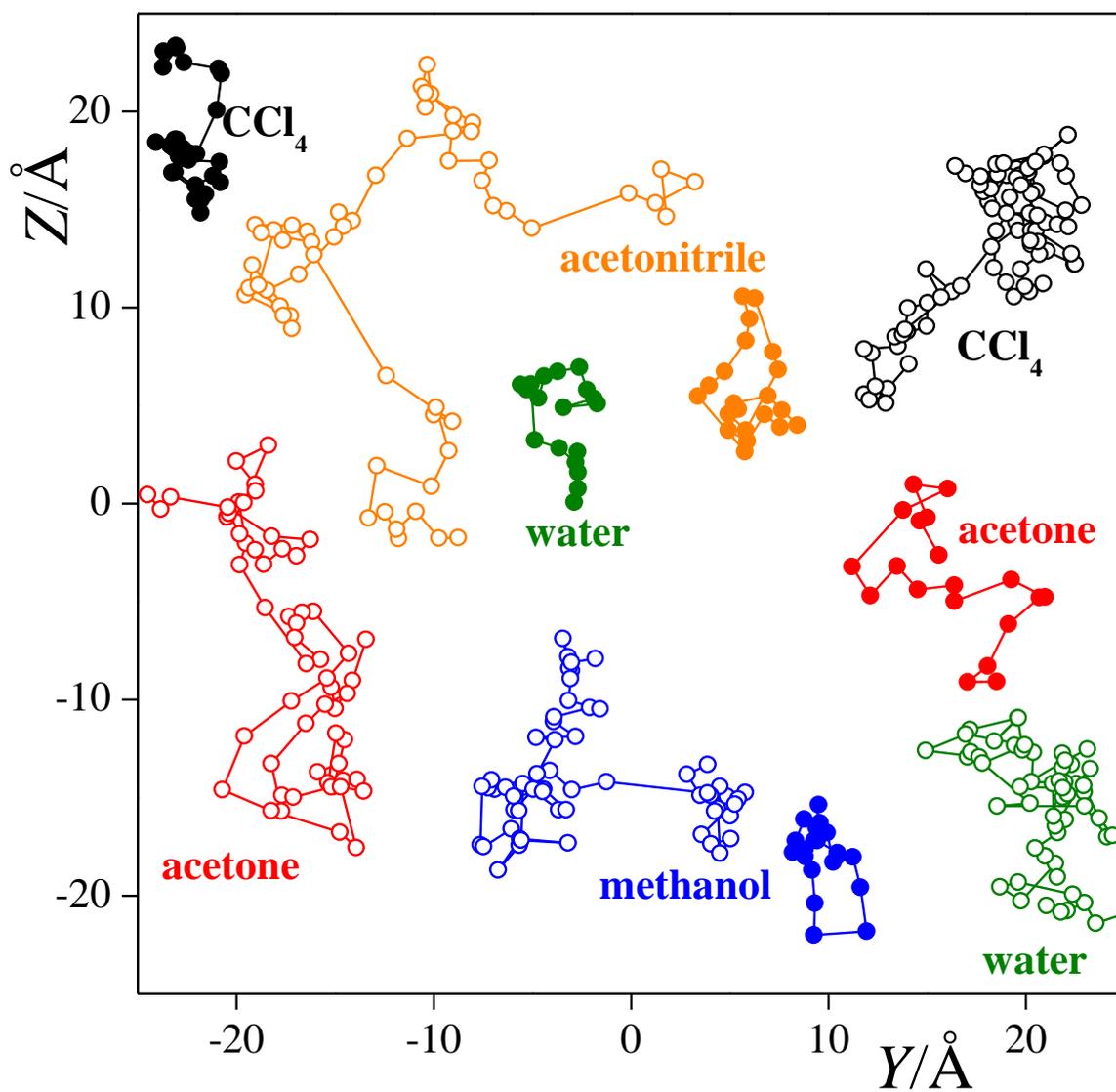


Figure 6.
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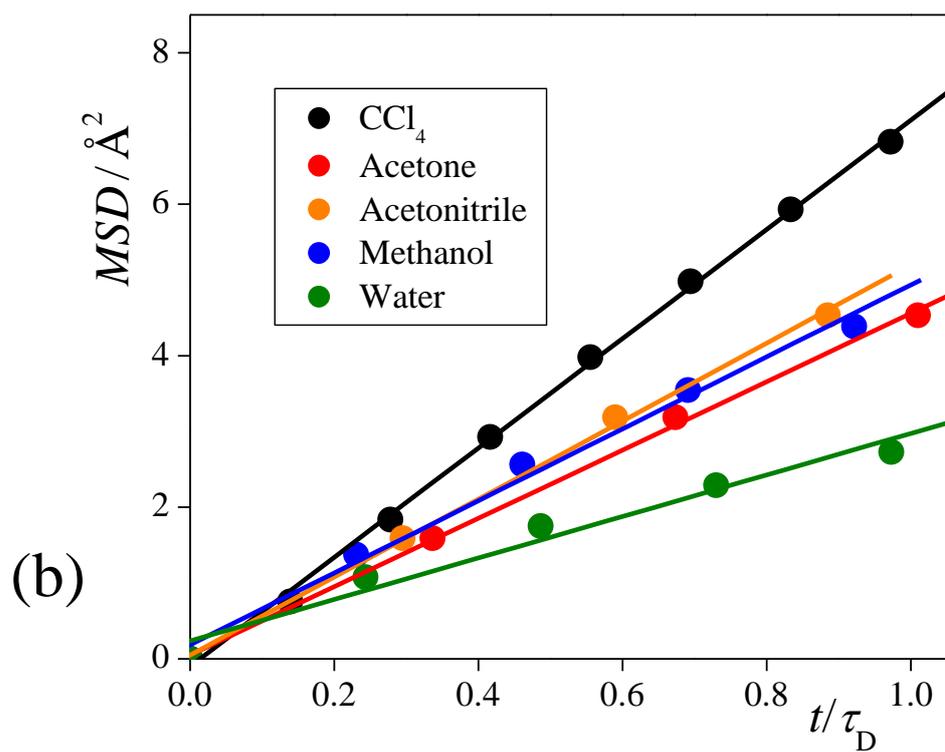
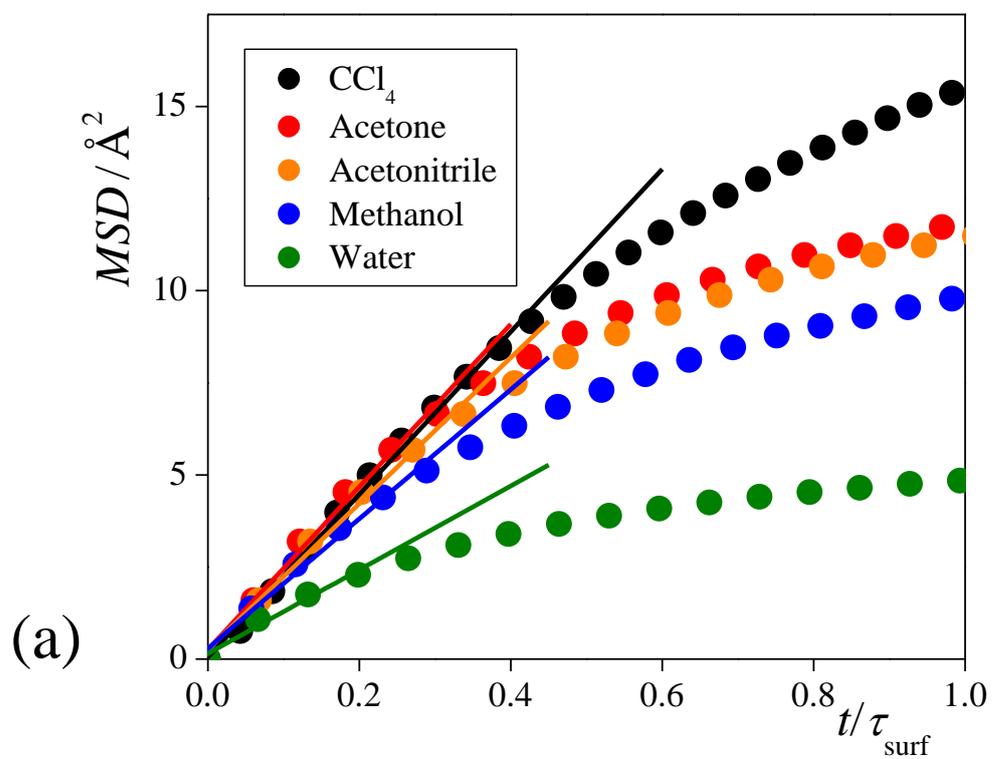


Figure 7.
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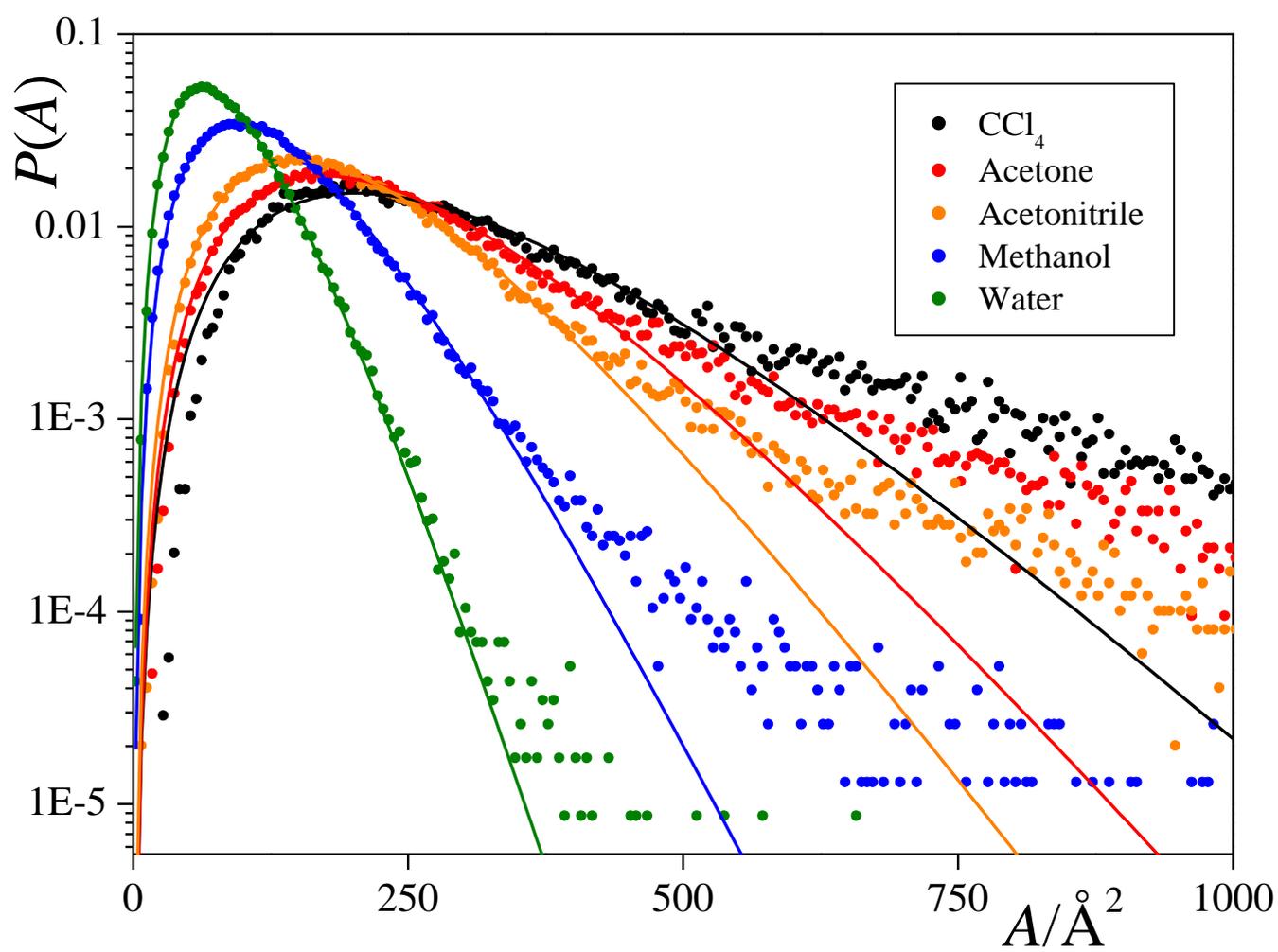


Figure 8.
Fábián et al.

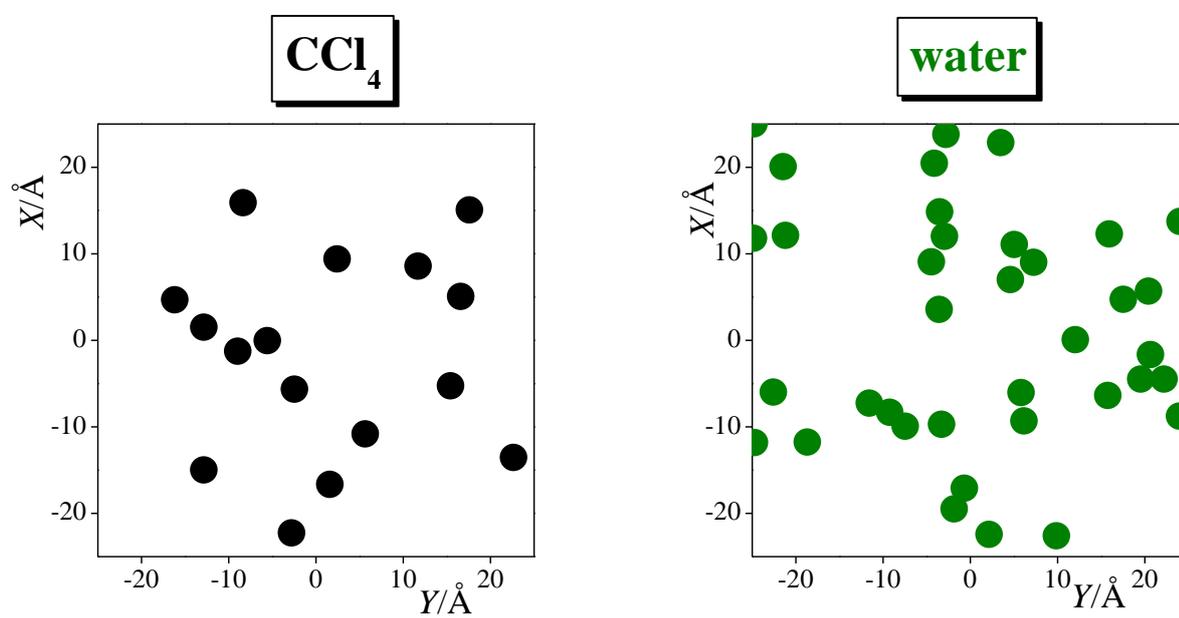


Figure 9.
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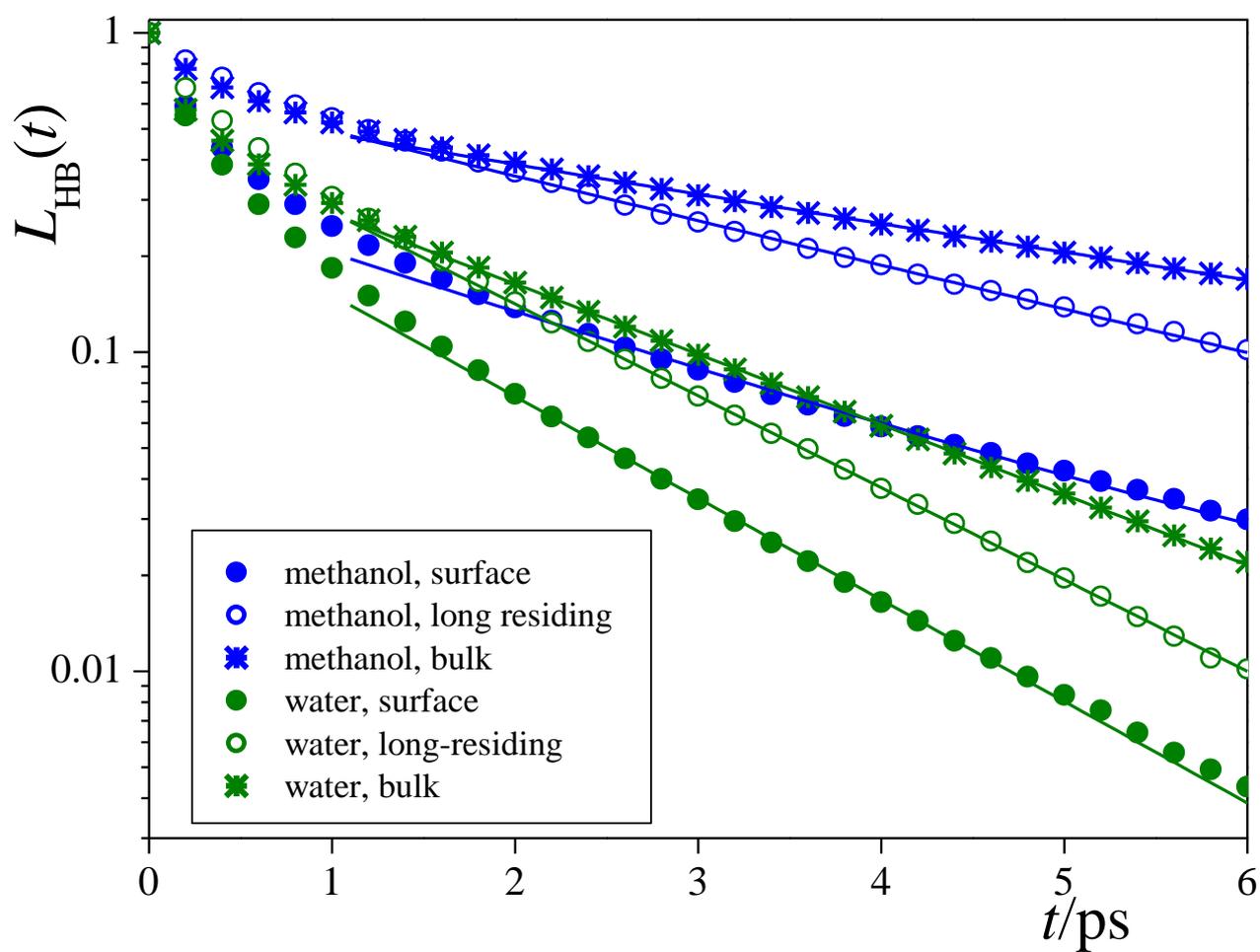


Figure 10.
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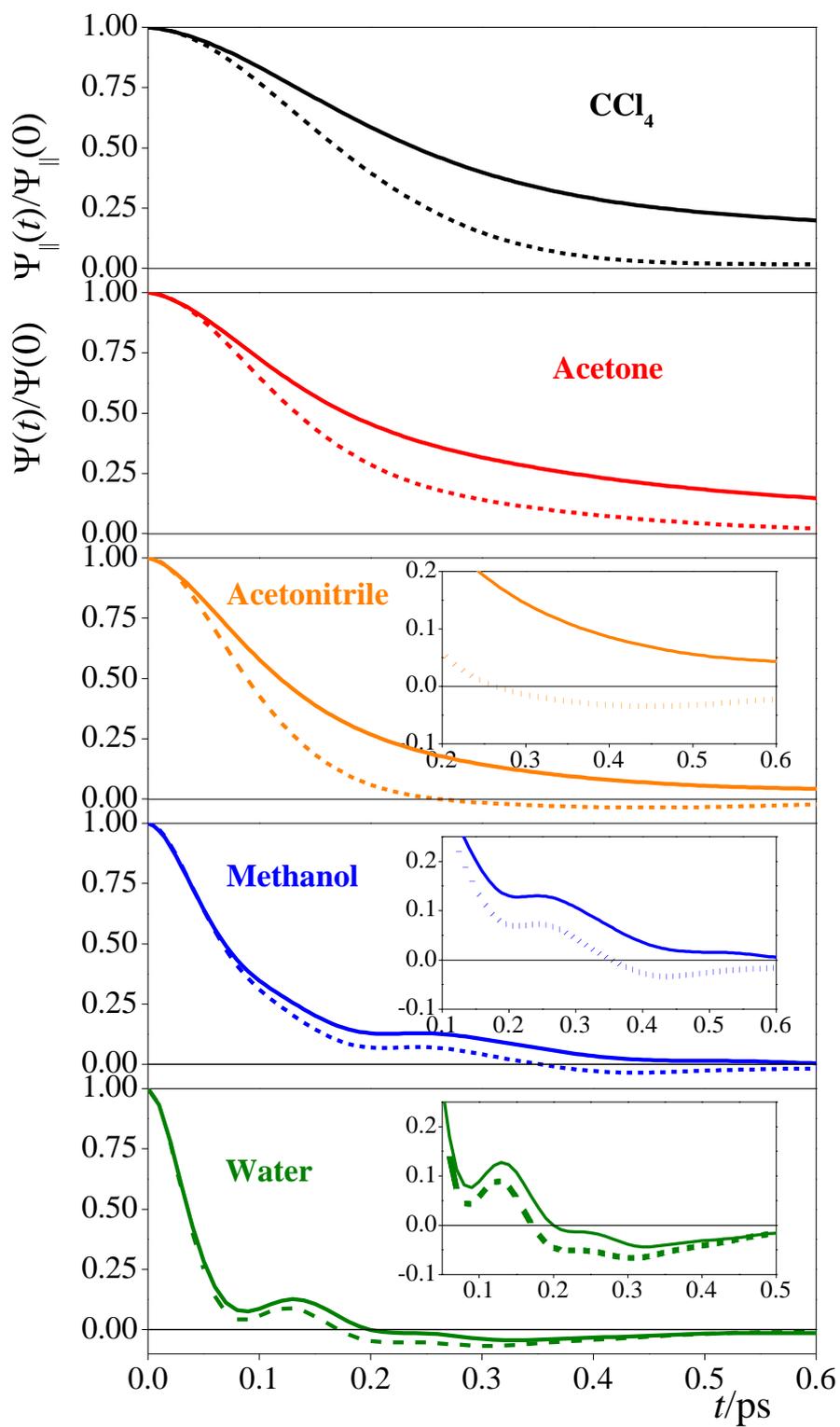


Table of Contents Graphics:

