

# IS THERE A HORMONAL REGULATION OF PHAGOCYTOSIS AT UNICELLULAR AND MULTICELLULAR LEVELS? A CRITICAL REVIEW

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Phagocytosis is an ancient cell function, which is similar at unicellular and multicellular levels. Unicells synthesize, store, and secrete multicellular (mammalian) hormones, which influence their phagocytosis. Amino acid hormones, such as histamine, serotonin, epinephrine, and melatonin stimulate phagocytosis, whereas peptide hormones, such as adrenocorticotrophic hormone (ACTH), insulin, opioids, arginine vasopressin, and atrial natriuretic peptide decreased it, independently on their chemical structure or function in multicellulars. Macrophage phagocytosis of multicellulars is also stimulated by amino acid hormones, such as histamine, epinephrine, melatonin, and thyroid hormones, however, the effect of peptide hormones is not uniform: prolactin, insulin, glucagon, somatostatin, and leptin have positive effects, whereas ACTH, human chorionic gonadotropin, opioids, and ghrelin have negative ones. Steroid hormones, such as estrogen, hydrocortisone, and dexamethasone are stimulating macrophage phagocytosis, whereas progesterone, aldosterone, and testosterone are depressing it. Considering the data and observations there is not a specific phagocytosis hormone, or a hormonal regulation of phagocytosis neither unicellular, nor multicellular level, however, hormones having specific functions in multicellulars also influence phagocytosis at both levels universally (in unicellulars) or individually (in macrophages). Nevertheless, the hormonal influence cannot be neglected, as phagocytosis (as a function) is rather sensitive to minute dose of hormones and endocrine disruptors. The hormonal influence of phagocytosis by macrophages can be deduced to the events at unicellular level.

**Keywords:** phagocytosis, *Tetrahymena*, macrophage, evolution, hormonal effects

Phagocytosis is a basic life function for unicellulars. Corpuscular elements as nourishments or hostile other unicellulars are engulfed by it. Phagocytosis does not a universal function of all cells in the multicellular world, however, it is

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absolutely needed as a step of innate immunity, which initiates the adaptive immune response. Macrophages, neutrophil granulocytes, and dendritic cells recognize the large particles (at least 0.5  $\mu\text{m}$ ) and engulf them by using pseudopods [1, 2], which included the phagocytized cell or material. Actin filaments have a role in transporting the phagocytic vesicles, which finished their way after fusing with lysosomes. This is a complex process, which is spontaneously executed, however, it could take or tolerate hormonal regulation.

In multicellular organisms, hormones are at the service of chemical regulation and phagocytic cells are hormonally influenced [3]. Unicellulars synthesize, store, and secrete amino acid- and polypeptide-type hormones characteristic to multicellular animals (mammals) [4–14]. In addition, the cells have mammalian receptor-like structures in the plasma membrane, which bind these hormones and the cells react to them [15–19], as they have signal transducer pathways [20–23]. Many cell functions of the unicellular ciliate *Tetrahymena* are influenced by the hormones and their effect sometimes seems to be specific [19]. The first encounter with an artificially given hormone causes hormonal imprinting, which provokes a quantitatively altered reaction and this inherited to the progenies up to the thousandth generations [24, 25].

As hormones influence phagocytosis at both phylogenetic levels, it seems worth to study the character of the effect as well as the similarities and differences.

## Facts at Unicellular Level

### *Amino acid hormones*

Single histamine treatment increased the phagocytic activity of *Tetrahymena pyriformis* [26]. Chronic histamine treatment was more intense and the intensified activity remained high after some time in histamine-free medium [27]. The action of histamine was dose-dependent. Serotonin also stimulated phagocytosis in *Tetrahymena* [28] as well, as epinephrine [29]. In *Tetrahymena thermophila* histamine was ineffective, whereas the antihistamine, diphenhydramine increased it [30]. In other experiments, H1 and H2 antagonists were studied and these substances did not influence phagocytosis in *T. pyriformis*. However, H1 antagonist phenindamine counteracted the phagocytosis stimulating effect of histamine, whereas H2 antagonist metiamide was ineffective [31]. Serotonin and catecholamine stimulated phagocytosis in *T. thermophila* [32]. Serotonergic antagonists spiperone and metergoline also stimulated the process, whereas propranolol, alprenolol, and ergocryptine, which are beta and alpha adrenergic antagonists were ineffective or inhibitors [32]. In *Paramecium aurelia*, beta adrenergic

agonists (isoproterenol and norepinephrine) enhanced phagocytosis stereospecifically and dose-dependently. The effect was inhibited by propranolol and alprenolol [33]. Histidine, the basic amino acid for histamine formation also stimulated phagocytosis in *T. pyriformis* even stronger than histamine itself [34].

The pineal hormone, melatonin, between  $10^{-6}$  and  $10^{-10}$  M concentrations significantly stimulated the *E. coli* phagocytosis of *T. pyriformis* [35, 36].

The effect of amino acid-type hormones is manifested through the adenylate cyclase–adenosine monophosphate (cAMP) system. Treatment with cAMP or cPDE inhibitors (as theophylline) increases phagocytic activity of *T. pyriformis* [37, 38].

### *Peptide hormones*

Adrenocorticotrophic hormone (ACTH) and insulin inhibited the phagocytic activity of *Tetrahymena*. [39]. In addition, insulin antagonized the phagocytosis increasing action of histamine in *T. pyriformis* [40]. Arginine vasopressin decreased the phagocytic activity of *T. pyriformis* [41]. Atrial natriuretic peptide is also a potent inhibitor of phagocytosis in *T. pyriformis* [42].

*Tetrahymena* synthesizes beta-endorphin-like proteins [43] and have receptors for opioid peptides [43], which are suitable for testing opioid peptides of metazoa. These latter inhibit the phagocytotic activity of *T. thermophila* [44]. *Tetrahymena* opioids inhibit phagocytosis of *Tetrahymena* by a naloxone-reversible mechanism [45]. The opioid receptors of *Tetrahymena* are more sensitive to beta-endorphin and most sensitive to morphine [46], which points to the mu-likeness in some pharmacological characters. Chronic treatment with an opioid causes tolerance [47].

### *Steroid hormones*

Dexamethasone and prednisolone stimulated the phagocytosis by *T. pyriformis*, but prednisolone-sodium-succinate and deoxycorticosterone-glucoside inhibited it [48].

## **Facts at Multicellular Level**

### *Amino acid hormones*

Histamine is believed to be the physiological activator of phagocytosis, since the basic works of Jancsó [49]. These data were supported by the

experiments with tubercle bacilli [50] and staphylococci [51]. However, there were studies, which demonstrated neutral or negative effects [52]. Epinephrine stimulated macrophage phagocytosis [53], whereas norepinephrine suppressed phagocytosis of wound neutrophils [54].

Induction of phagocytosis in murine macrophages is positively influenced by thyroid hormones through a glutamine mechanism [55]. Exercise (swimming) increases phagocytosis and thyroid hormones are responsible for it [56]. Triiodo-thyronine (T3) stimulated granulocytes' phagocytic activity [57, 58]. Melatonin increased engulfment of latex beads [59]. Physiological phagocytosis by neutrophil granulocytes seems to be dependent on the presence of nocturnal melatonin surge [60]. Alcohol treatment provokes a drastic decrease in neutrophil phagocytosis, which is restored by melatonin. Stress caused by swimming to exhaustion provoked lower melatonin peak and consequently higher phagocytic activity of macrophages [61]. Melatonin also suppressed phagocytic activity of cultured retinal pigment cells [62].

### *Peptide hormones*

Prolactin increases the *in vitro* phagocytic capacity of macrophages [63] and helps to stimulate the exercise (swimming) induced phagocytosis [56]. Follicle-stimulating hormone (FSH) negatively influences phagocytic activity of Sertoli cells in tissue cultures [64]. ACTH suppresses phagocytosis of murine peritoneal macrophages [65], however, contradictory results are also known [66]. Chorionic gonadotropin suppresses the phagocytic activity of blood leukocytes and peritoneal macrophages [67, 68]. Opioids, such as endorphin and dynorphin, stimulate phagocytosis of mouse macrophages [66]. Insulin supports the onset of phagocytosis in inflammatory macrophages by a glutamine-transmitted mechanism [55] and restores neutrophil phagocytosis in diabetic patients [69]. Chronic treatment with insulin strongly depressed the macrophage phagocytosis in rats [70]. The phagocytic activity is low in type 2 diabetes and improves after metabolic improvement [71]. Insulin inhibits phagocytosis of normal human neutrophils [72]. It also enhances immunological phagocytosis by macrophages [72]. Glucagon and somatostatin stimulated macrophage phagocytosis [70]. Met-enkephalin, leu-enkephalin, and beta-endorphin reduced phagocytosis of *Candida albicans* by human monocytes [73]. Beta-endorphin also suppressed the phagocytic activity of splenic phagocytes and this was antagonized by opioid receptor antagonist, naltrexone [74]. Ghrelin decreased the phagocytic activity of cold-restraint stress exposed rats [75]. Leptin, the adipocyte hormone activates mononuclear phagocytes by a JAK/STAT signaling pathway [76].

### *Steroid hormones*

Estrogen (E2) or progesterone significantly enhanced the phagocytosis of rat peritoneal macrophages [77]. Phagocytosis by human mononuclear cells was stimulated by dexamethasone or hydrocortisone [78]. In tissue culture, murine Sertoli cell phagocytosis was stimulated by hydrocortisone [79]. Prednisolone inhibited the latex phagocytic capacity of human granulocytes, by a receptor-mediated manner [80]. When prednisolone depressed phagocytic function, vitamin D3 or vitamin E partially restored it [81]. Progesterone reduced *E. coli* phagocytosis of cultured human decidual cells [82]. Gonadectomy in both sexes (in mice) significantly reduced phagocytic activity of peritoneal macrophages. In females, estradiol supplementation restored the normal condition, however, dihydrotestosterone treatment in males was insufficient [83]. In freshwater snake, *Natrix piscator* testosterone depressed phagocytic activity of splenic macrophages [84]. In common carp, *Cyprinus carpio*, beta-estradiol, 11 ketotestosterone, and progesterone suppressed phagocytosis of kidney macrophages in a dose-dependent manner [85, 86]. In tilapia, cortisol and dexamethasone decreased phagocytosis, and aldosterone had a weaker effect [87]. A chronic treatment with estradiol, testosterone, or dihydrotestosterone in chicks significantly depressed the phagocytic activity of macrophages [88].

### **Conclusions**

*Tetrahymena* produce amino acid-type hormones as well as peptide ones. However, at unicellular level, there was not systematic investigation of these hormones in case of phagocytosis, but typical hormones were studied which allows the drawing of some conclusions. At multicellular level, more experiments and observation were performed and practically all important mammalian hormones were studied, sometimes with contradictory results, mainly depending on the used methods and subject species. This is understandable, as multicellular phagocytosis is part of the immune process, which have decisive role in the manifestation or healing of human diseases. Nevertheless, although phagocytosis is a rather complex process, mostly the engulfment of neutral particles or bacteria was studied under the effect of hormones and hardly are data on the behavior of actin network, on the encounter and fusion of endocytotic vesicles, etc. However, this has not importance from the aspect of our evaluation as phagocytosis, as a function is studied irrespective of details.

It is indisputable on basis of the data that at both levels of phylogeny hormones can influence phagocytosis. However, it is not known whether it is a

physiological interaction (regulation), which is needed for the normal execution of the function or coincidental because of the chemical structure of molecules. From evolutionary aspect, the problem seems to be more simple at unicellular level, as all of the amino acid hormones studied positively influenced phagocytosis (Table I), whereas peptide hormones affected it negatively (Table II). As the chemical structures inside the group given are very different and the modified amino acid molecule is easily distinguishable from a peptide chain this could mean that amino acid hormones – or may be amino acids, which were not fully studied from this point of view – are stimulating phagocytosis and polypeptide hormones – or may be peptides, which also have not studied from this point of view – are influencing negatively the process. This is supported by a study, in which histamine and serotonin enhanced the adsorption of fluorescein isothiocyanate (FITC)-labeled bovine serum albumin (BSA) to the plasma membrane of *Tetrahymena*, while a similar action by insulin was not significant. The degree of BSA binding was similar to the degree of phagocytosis [89]. However, the effect of epinephrine on BSA binding was also not significant which weakens the conclusion.

Steroid hormones positively influenced phagocytosis in *T. pyriformis* (Table III), however, it is questionable whether steroids are used for

**Table I.** Hormone-influenced phagocytosis in unicellulars: amino acid hormones

Hormone	Species	Effect +/-
Histamine	TP	+
Histamine/chronic	TP	+
Serotonin	TP	+
Serotonin	TT	0
Serotonin	TT	+
Epinephrine	TT	+
Epinephrine	P	+
Melatonin	TP	+
Histidine (amino acid)	TP	+

Note: TP: *Tetrahymena pyriformis*, TT: *Tetrahymena thermophila*, P: paramecium.

**Table II.** Hormone-influenced phagocytosis in unicellulars: peptide hormones

Hormone	Species	Effect +/-
ACTH	TP	–
ANP	TP	–
Opioids	TP	–
Insulin	TP	–
Vasopressin	TP	–

Note: TP: *Tetrahymena pyriformis*.

**Table III.** Hormone-influenced phagocytosis in unicellulars: steroid hormones

Hormone	Species	Effect +/-
Dexamethasone	TP	+
Prednisolone	TP	+
Prednisolone-sodium-succinate	TP	-
Deoxycorticosterone-glucoside	TP	-

Note: TP: *Tetrahymena pyriformis*.

communication at all, as the unicells are living in a watery milieu in which steroids are not dissolved, in addition they have not steroid receptors and their induced steroid receptors are not individual hormone-specific [90].

The ideal phagocyte models in multicellulars are the macrophages as they are the “professional” phagocytes. The effect of amino acid hormones on macrophages is identical with the unicellular ones (Tables IV and VII). However, the effect of peptide hormones is not so clear from this point of view (Table V). At a rough estimate the same amount of them is observed with positive, as negative

**Table IV.** Hormone-influenced phagocytosis in multicellulars: amino acid hormones

Hormone	Cell type	Effect +/-
Histamine	Macrophage/granulocyte	+
Epinephrine	Macrophage	+
Nor-epinephrine	Macrophage	-
Thyroid	Macrophage	+
Melatonin	Macrophage	+
Melatonin	Retinal pigment	-

**Table V.** Hormone-influenced phagocytosis in multicellulars: peptide hormones

Hormone	Cell type	Effect +/-
Prolactin	Macrophage	+
ACTH	Macrophage	-
HCG	Macrophage/granulocyte	-
FSH	Sertoli cell	-
Opioids	Macrophage	+
Endorphin	Macrophage	-
Insulin	Macrophage	+
Insulin/chronic	Macrophage	-
Insulin	Granulocyte	-
Glucagon	Macrophage	+
Somatostatin	Macrophage	+
Ghrelin	Macrophage	-
Leptin	Macrophage	+

**Table VI.** Hormone-influenced phagocytosis in multicellulars: steroid hormones

Hormone	Cell type	Effect +/-
Estrogen	Macrophage	+
Estrogen/chronic	Macrophage	–
Progesterone	Macrophage	–
Progesterone	Macrophage (carp)	–
Testosterone	Macrophage	–
Ketotestosterone	Macrophage (carp)	–
Aldosterone	Macrophage (carp)	–
Hydrocortisone	Macrophage	+
Hydrocortisone	Sertoli cell	+
Dexamethasone	Macrophage	+
Prednisolone	Granulocyte	–
Testosterone	Splenic macrophage (snake)	–

effects. This could mean that in the case of amino acid hormones the amino acid character is dominant, whereas in the case of polypeptide hormones the specificity of the peptide chain. Considering immunity, phagocytosis belongs to the innate immunity, which is absolutely needed for the expression of adaptive immunity, however, this type of phagocytosis is not identical with the unicellulars' phagocytosis, from hormonal aspect. In addition, in multicellulars, the same hormone can participate in the control of innate and adaptive immunity alike [91, 92]. In the case of steroid hormones – and in multicellulars these are working – again their individual character has the decisive role (Table VI). “Glucocorticoids are the main effectors” which are bound by glucocorticoid receptors of immune cells (in macrophages included) [93].

As it was mentioned above, the unicellular *Tetrahymena* synthesize, store, and secrete hormones characteristic to multicellular animals (mammals). These

**Table VII.** Comparison of hormone-influenced phagocytosis in unicellulars and multicellulars

	Effect in unicellulars	Effect in multicellulars
Histamine	+	+
Epinephrine	+	+
Melatonin	+	+
ACTH	–	–
Insulin	–	+
Opiates	–	Uncertain
Dexamethasone	+	+
Prednisolone	+	Uncertain



unicellular hormones act to cells of metazoa similar to hormones of their own [94]. The unicell also has receptors for mammalian hormones and the receptors' structure is similar to mammalian ones [15, 16, 95], and can transmit hormonal information into the cells, which provokes response. However, the hormones which are classified to a group (e.g. amino acid type or peptide) are provoking similar (identical) reactions (positive by amino acid and negative by peptide). This means that there is not an individual hormone-specificity, but a hormone-type specificity which is not suitable for a hormonal regulation, but enough for being the base of a phylogenetic development of later hormonal control. This makes likely that the regulation by polypeptide hormones and steroids can be deduced to this type of group regulation. It is interesting that in the professional phagocytes (macrophages) of multicellular organisms, the amino acid hormones affect phagocytosis similar to the unicellulars (group-like), but peptide hormones have individual effect. This could mean that the influence of phagocytosis in the frame of the evolution of immune mechanisms [96] runs parallel with the differentiation of hormones [93] and many components of unicellular phagocytosis have been conserved in higher ranked animals [97].

Phagocytosis is a form of endocytosis, when corpuscular elements are engulfed by the cell and it is a very ancient process [98]. The other form is pinocytosis when dissolved materials are taken up. These materials could be amino acids and these are utilized by the phagocyte for building up proteins. This means that the presence of amino acids in the environment is a positive signal for endocytosis which is also studied by the engulfment of particles (phagocytosis). This could also mean that amino acid hormones stimulate phagocytosis by their amino acid character, and this could explain the uniform effect of them.

The unicellular animal is composed of one cell, however, it is also a complete organism, which has all of the organs (organelles) which are needed for life. It synthesizes all of the water-soluble hormones which have been studied and also can react to them if these materials are present around it in the watery milieu. The macrophages of the multicellular organisms are able to synthesize hormones, however, these are produced by professional hormone producing organs or cells, as there is a division of labor in the organism. Steroid hormones are also produced and transported to the site of the effect. This means that in both cases there is a possibility of hormonal regulation of the phagocytic function [19].

Answering the question in the title of this paper: in our present knowledge, there is not a hormonal regulation of phagocytosis, neither unicellular, nor multicellular level. However, hormones are synthesized, stored, and secreted by unicells which can influence phagocytosis [99, 100] and there is a similar situation in multicells, where hormones which has specific functions (e.g., regulation of sugar metabolism, blood calcium level, ovarian cycle, etc.) are also influencing the

phagocytosis of professional cells (macrophages and neutrophil granulocytes). There is a similar situation if other cell functions would be studied, as specific hormones always have side effects on cells which are not in the mainstream of the hormonal effect. I cannot be excluded the existence of a specific phagocytosis hormone, or hormone-like molecule with specific phagocyte regulating activity at any levels of phylogeny, however, it is not known. At the same time, the hormones presently have been studied permits some evolutionary conclusions: their effects on phagocytosis of macrophages can be deduced to the effects on unicells.

Although in our present knowledge, there is not exist a hormonal regulation of phagocytosis, or specific phagocytosis hormone, the influence of hormones to phagocytosis cannot be undervalued. This is especially very important in our modern world, where hormone-like endocrine disruptors are present in increasing number and increasing amount. There are many data that these materials can alter normal immune functions, phagocytosis included [101–105] and the functional alterations of phagocytosis could cause or promote diseases in the present and future human generations [106–108].

### Conflict of Interest

The author declares no conflict of interest.

### References

1. Flannagan, R. S.: The cell biology of phagocytosis. *Annu Rev Pathol* **7**, 6198 (2012).
2. Aderem, A.: Mechanism of phagocytosis in macrophages. *Annu Rev Immunol* **17**, 593–623 (1999).
3. Hunt, I. S., Miller, L., Platt, J. S.: Hormonal regulation of uterine macrophages. *Dev Immunol* **6**, 105–110 (1998).
4. Lajkó, E., Pállinger, É., Csaba, G.: Effect of glucose on the insulin production and insulin binding of *Tetrahymena*. *Acta Microbiol Immunol Hung* **59**, 461–468 (2012).
5. Rodriguez, E., Lazaro, M. I., Renaud, F. L., Marino, M.: Opioid activity of beta-endorphin-like proteins from *Tetrahymena*. *J Eukaryot Microbiol* **51**, 60–65 (2004).
6. LeRoith, D., Liotta, A. S., Roth, J., Shiloach, J., Lewis, M. E., Pert, C. B., Krieger, D. T.: Corticotropin and beta-endorphin-like materials are native to unicellular organisms. *Proc Natl Acad Sci USA* **79**, 2086–2090 (1982).
7. Berelowitz, M., LeRoith, D., von Schenk, H., Newgard, C., Szabo, M., Frohman, L. A., Shiloach, J., Roth, J.: Somatostatin-like immunoreactivity and biological activity is present in *Tetrahymena pyriformis*, a ciliated protozoan. *Endocrinology* **110**, 1939–1944 (1982).

8. Schwabe, C., LeRoith, D., Thompson, R. P., Shiloach, J., Roth, J.: Relaxin extracted from protozoa (*Tetrahymena pyriformis*). Molecular and immunological properties. *J Biol Chem* **258**, 2778–2781 (1983).
9. Deftos, L. J., LeRoith, D., Shiloach, J., Roth, J.: Salmon calcitonin-like immunoactivity in extracts of *Tetrahymena pyriformis*. *Horm Metab Res* **17**, 82–85 (1985).
10. LeRoith, D., Shiloach, J., Heffron, R., Rubinovitz, C., Tanenbaum, R., Roth, J.: Insulin-related material in microbes: Similarities and differences from mammalian insulins. *Can J Biochem Cell Biol* **63**, 839–849 (1985).
11. de Pablo, F., Lesniak, M. A., Hernandez, E. R., LeRoith, D., Shiloach, J., Roth, J.: Extracts of protozoa contain materials that react specifically in the immunoassay for guinea pig insulin. *Horm Metab Res* **18**, 82–87 (1986).
12. Csaba, G., Pállinger, É.: Thyrotropin (TSH) regulates triiodothyronine (T3) production in the unicellular *Tetrahymena*. *Acta Biol Hung* **62**, 228–234 (2011).
13. Lajkó, E., Pállinger, É., Csaba, G.: Investigations on the triiodothyronine (T3)-specificity of thyrotropic (TSH) and gonadotropic (HCG) hormone in the unicellular *Tetrahymena*. *Acta Microbiol Immunol Hung* **58**, 85–91 (2011).
14. Csaba, G., Pállinger, É.: Is there a hormonal network in *Tetrahymena*? A systematic investigation of hormonal effects on the hormone content. *Cell Biochem Funct* **26**, 303–308 (2008).
15. Christopher, G. K., Sundermann, C. A.: Isolation and characterization of the insulin binding sites of *Tetrahymena pyriformis*. *Biochem Biophys Res Commun* **212**, 515–523 (1995).
16. Christensen, S. T., Guerra, C. F., Awan, A., Wheatley, D. N., Satir, P.: Insulin receptor-like proteins in *Tetrahymena thermophila* ciliary membranes. *Curr Biol* **13**, R50–R52 (2003).
17. Chiesia, R., Silva, W. I., Renaud, F. L.: Pharmacological characterization of an opioid receptor in the ciliate *Tetrahymena*. *J Eukaryot Microbiol* **40**, 800–804 (1993).
18. Csaba, G., Kovács, P.: Oxytocin and vasopressin change the activity of the contractile vacuole in *Tetrahymena*: Newer contributions to the phylogeny of hormones and hormone receptors. *Comp Biochem Physiol Comp Physiol* **102**, 353–355 (1992).
19. Csaba, G.: The hormonal system of the unicellular *Tetrahymena*: A review with evolutionary aspects. *Acta Microbiol Immunol Hung* **59**, 131–156 (2012).
20. Shpakov, A. O., Derkach, K. V., Uspenskaia, Z. I.: Effect of natural amino acids and sugars on cyclase activities in infusoria *Tetrahymena pyriformis* and *Dileptus anser*. *Zh Evol Biokhim Fiziol* **47**, 128–135 (2011).
21. Kőhidai, L., Barsony, J., Roth, J., Marx, S. J.: Rapid effects of insulin on cyclic GMP location in an intact protozoan. *Experientia* **15**, 476–481 (1992).
22. Shpakov, A. O., Derkach, K. V., Uspenskaya, Z. I.: Glucose and cyclic adenosine monophosphate stimulate activities of adenylate cyclase and guanylate cyclase of *Tetrahymena pyriformis* infusoria. *Bull Exp Biol Med* **152**, 427–430 (2012).
23. Plattner, H.: Signalling in ciliates: Long-and short-range signals and molecular determinants for cellular dynamics. *Biol Rev Camb Philos Soc* **92**, 60–107 (2017).
24. Csaba, G.: Hormonal imprinting in the unicellular *Tetrahymena*: The proto-model of epigenetics. *Acta Microbiol Immunol Hung* **59**, 291–310 (2012).

25. Kőhidai, L., Lajkó, E., Pállinger, É., Csaba, G.: Verification of epigenetic inheritance in a unicellular model system: Multigenerational effects of hormonal imprinting. *Cell Biol Int* **36**, 951–959 (2012).
26. Csaba, G., Nagy, S. U., Lantos, T.: Are biogenic amines acting on *Tetrahymena* through a cyclic AMP mechanism? *Acta Biol Med Ger* **35**, 259–261 (1976).
27. Csaba, G., Lantos, T.: An attempt to differentiate selection and amplification in hormone receptor development: The unicellular model. *Differentiation* **8**, 57–59 (1977).
28. Darvas, Z., Csaba, G.: Dose-dependent impact of pretreatment (imprinting) with histamine and serotonin on the phagocytic activity of *Tetrahymena*. *Acta Microbiol Hung* **37**, 285–287 (1990).
29. Csaba, G.: Biogenic amines at a low level of evolution: Production, functions and regulation in the unicellular *Tetrahymena*. *Acta Microbiol Immunol Hung* **62**, 93–108 (2015).
30. Buduma, N., Balabanian, J., Dalvi, P., Chia, S.-K., Dhaliwal, A., Eliya, D., Boothby, J., Bros-Seemann, S., Kibler, L., Khurt, S., Veregge, S.: Modulation of phagocytosis in *Tetrahymena thermophila* by histamine and antihistamine diphenhydramine. *Acta Protozool* **52**, 317–323 (2013).
31. Csaba, G., László, V., Darvas, Z.: Effects of H1 and H2 antagonists on *Tetrahymena*. *Acta Biol Med Ger* **37**, 161–163 (1978).
32. Quinones-Maldonado, V., Renaud, F. L.: Effect of biogenic amines on phagocytosis in *Tetrahymena thermophila*. *J Protozool* **34**, 435–438 (1987).
33. Wyroba, E.: Beta-adrenergic stimulation of phagocytosis in the unicellular eukaryote *Paramecium aurelia*. *Cell Biol Int Rep* **13**, 667–678 (1989).
34. Csaba, G., Darvas, Z.: Receptor-level interrelationships of amino acids and the adequate amino acid type hormones in *Tetrahymena*: A receptor evolution model. *Biosystems* **19**, 55–59 (1986).
35. Kőhidai, L., Vakkuri, O., Keresztesi, M., Pállinger, É., Leppaluoto, J., Csaba, G.: Impact of melatonin on the cell division, phagocytosis and chemotaxis of *Tetrahymena pyriformis*. *Acta Protozool* **41**, 85–89 (2002).
36. Csaba, G.: Presence in and effects of pineal indoleamines at very low level of phylogeny. *Experientia* **49**, 627–634 (1993).
37. Csaba, G., Nagy, S. U., Lantos, T.: Cyclic AMP and its functional relationship in *Tetrahymena*: A comparison between phagocytosis and glucose uptake. *Acta Biol Med Ger* **37**, 505–507 (1978).
38. Csaba, G., Lantos, T.: Effect of cyclic AMP and theophylline on phagocytotic activity of *Tetrahymena pyriformis*. *Experientia* **32**, 321–322 (1976).
39. Kőhidai, L., Lovas, B., Csaba, G.: Effect of adrenocorticotrophic hormone (ACTH) and insulin on the phagocytic capacity of *Tetrahymena*. *Zoolog Sci* **12**, 277–281 (1995).
40. Csaba, G., Darvas, Z.: Insulin antagonizes the phagocytosis stimulating action of histamine in *Tetrahymena*. *Biosci Rep* **12**, 23–27 (1992).
41. Jahn, I., Csaba, G.: The influence of arginine vasopressin (AVP) on phagocytosis in the unicellular *Tetrahymena*. *Acta Protozool* **26**, 39–44 (1987).
42. Kőhidai, L., Csaba, G., Karsa, J.: Effects of atrial natriuretic peptide on the unicellular *Tetrahymena pyriformis* model. *Microbios* **82**, 27–40 (1995).
43. Rodriguez, E., Lazaro, M. I., Renaud, F. L., Marino, M.: Opioid activity of beta-endorphin-like proteins from *Tetrahymena*. *J Eukaryot Microbiol* **51**, 60–65 (2004).

44. DeJesus, S., Renaud, F. L.: Phagocytosis in *Tetrahymena thermophila*: Naloxone reversible inhibition by opiates. *Comp Biochem Physiol C Comp Pharmacol* **92**, 139–142 (1989).
45. Renaud, F. L., Colon, I., Lebron, J., Ortiz, N., Rodriguez, F., Cadilla, C.: A novel opioid mechanism seems to modulate phagocytosis in *Tetrahymena*. *J Eukaryot Microbiol* **42**, 205–207 (1995).
46. Chiesa, R., Silva, W. I., Renaud, F. L.: Pharmacological characterization of an opioid receptor in the ciliate *Tetrahymena*. *J Eukaryot Microbiol* **40**, 800–804 (1993).
47. Salaman, A., Roman, M., Renaud, F. L., Silva, W. I.: Effect of chronic opioid treatment on phagocytosis in *Tetrahymena*. *Neuropeptides* **16**, 115–120 (1990).
48. Csaba, G., Lantos, T., Nagy, S. U., Arányi, P., Náray, A.: Effects of steroids on *Tetrahymena*. *Acta Biol Med Ger* **37**, 1377–1380 (1978).
49. Jancsó, M.: Histamine as a physiological activator of the reticulo-endothelial system. *Nature* **159**, 227 (1947).
50. Gözsy, B., Kató, L.: Studies on the effects of phagocytic stimulation on microbial disease: Stimulation of phagocytic activity of monocytes against tubercle bacilli, strain BCG. *Can J Biochem Physiol* **34**, 571–579 (1956).
51. Northover, J. B.: The effect of histamine and 5-hydroxytryptamine on phagocytosis of staphylococci in vitro by polymorphs and macrophages. *J Pathol Bacteriol* **82**, 355–361 (1961).
52. Azuma, Y., Shinohara, M., Wang, P. L., Hidaka, A., Ohura, K.: Histamine inhibits chemotaxis, phagocytosis, superoxide anion production, and the production of TNF alpha and IL-12 by macrophages via H2-receptors. *Int Immunopharmacol* **1**, 1867–1875 (2001).
53. Zhou, J., Yan, J., Liang, H., Jiang, J.: Epinephrine enhances the response of macrophages under LPS stimulation. *Biomed Res Int* **2014**, Article ID 254686 (2014).
54. Gosain, A., Gamelli, R. L., Di Pietro, L. A.: Norepinephrine mediated suppression of phagocytosis by wound neutrophils. *J Surg Res* **152**, 311–318 (2009).
55. Costa Rosa, L. E., Cury, Y., Curi, R.: Hormonal control of macrophage function and glutamine metabolism. *Biochem Cell Biol* **69**, 309–312 (1991).
56. Ortega, E., Rodriguez, M. J., Barriga, C., Forner, M. A.: Corticosterone, prolactin and thyroid hormones as hormonal mediators of stimulated phagocytic capacity of peritoneal macrophages after high-intensity exercise. *Int J Sports Med* **17**, 149–155 (1996).
57. De Vito, P., Balducci, V., Leone, S., Percario, Z., Mangino, G., Davis, P. J., Davis, F. B., Alfabis, E., Luly, P., Pedersen, J. Z., Incerpi, S.: Nongenomic effects of thyroid hormones on the immune system cells: New targets, old players. *Steroids* **77**, 988–995 (2012).
58. Balázs, C., Leövey, A., Szabó, M., Bakó, G.: Stimulating effect of triiodothyronine on cell-mediated immunity. *Eur J Clin Pharmacol* **17**, 19–23 (1980).
59. Kanchev, I. N., Baichev, J., Kamenov, I., Baikov, B., Hallak, A. K.: Melatonin, corticosterone, stress and phagocytic activity. *Bulg J Vet Med* **9**, 257–264 (2006).
60. Hriscu, M. I.: Modulatory factors of circadian phagocytic activity. *Ann N Y Acad Sci* **1057**, 403–430 (2005).
61. Barriga, C., Martin, M. I., Tabla, R., Ortega, R., Rodriguez, A. B.: Circadian rhythm of melatonin, corticosterone and phagocytosis: Effect of stress. *J Pineal Res* **30**, 180–187 (2001).

62. Ogino, N., Matsumura, M., Shirakawa, H., Tsukahara, I.: Phagocytic activity of cultured retinal pigment epithelial cells from chick embryo: Inhibition by melatonin and cyclic AMP, and its reversal by taurine and cyclic GMP. *Ophthalmic Res* **15**, 72–89 (1983).
63. Ortega, E., Ferner, M. A., Barriga, C.: Effect of prolactin on the in vitro phagocytic capacity of macrophages. *Comp Immunol Microbiol Infect Dis* **19**, 139–146 (1996).
64. Filippini, A., Russo, M. A., Palombi, F., Bertalot, G., De Cesaris, P., Stefanini, M., Ziparo, E.: Modulation of phagocytic activity in cultured Sertoli cells. *Gamete Res* **23**, 367–375 (1989).
65. Ichinose, M., Savada, M., Maeno, T.: Suppression of phagocytosis by adrenocorticotrophic hormone in murine peritoneal macrophages. *Regul Pept* **54**, 457–466 (1994).
66. Xiaojing, W.: The effects of different opiate peptides and ACTH on the phagocytosis of mouse macrophages. *Chin J Immunol* **4** (1987).
67. Shirshev, S. V., Gorbunova, O. L.: Various mechanisms of the direct and indirect effects of chorionic gonadotropin on the phagocytic activity of leukocytes. *Izv Akad Nauk Ser Biol* **1**, 24–29 (2008).
68. Perntelewa-Rostaing, E., Fontagné, J., Adolphe, M., Engellman, P., Morin, P., Lechat, P.: Effect of human chorionic gonadotropin on phagocytic activity and proliferative capacity of rat peritoneal macrophages in culture. *Acta Endocrinol (Copenh)* **92**, 187–192 (1979).
69. Yano, H., Kinoshita, M., Fujino, K., Nakashima, M., Yamamoto, Y., Miyazaki, H., Mamada, K., Ono, S., Iwaya, K., Saitoh, D., Seki, S., Tanaka, Z.: Insulin treatment directly restores neutrophil phagocytosis and bactericidal activity in diabetic mice and thereby improves surgical site *Staphylococcus aureus* infection. *Infect Immun* **80**, 4409–4416 (2012).
70. Cornell, R. P., McClellan, C. C.: Modulation of hepatic reticuloendothelial system phagocytosis by pancreatic hormones. *J Reticuloendothel Soc* **32**, 397–407 (1982).
71. Lecube, A., Pechon, G., Petriz, J., Hernandez, C., Simo, R.: Phagocytic activity is impaired in Type 2 diabetes mellitus and increases after metabolic improvement. *PLoS One* **6**, e23366 (2011).
72. Lima, A. O., Queiroz, M., Brascher, H. M., Vargens, J.: Effect of insulin on immunological phagocytosis by macrophages. *Experientia* **35**, 119–120 (1979).
73. Prieto, J., Subira, M. L., Castilla, A., Arroyo, J. L., Serrano, M.: Opioid peptides modulate the organization of vimentin filaments, phagocytic activity, and expression of surface molecules in monocytes. *Scand J Immunol* **29**, 391–398 (1989).
74. Kumar, S., Ghorai, S. M., Rai, U.: Beta-endorphin inhibits phagocytic activity of lizard splenic phagocytes through mu receptor-coupled adenylate cyclase–protein kinase A signaling pathway. *Gen Comp Endocrinol* **171**, 301–308 (2011).
75. Turner, C., Bilgin, H. M., Obay, B. D., Diken, H., Tasdemir, E., Sermet, A.: Effect of ghrelin administration on phagocytic activity in acute cold restrained rats. *Regul Pept* **138**, 113–117 (2007).
76. Shirsev, S. V., Orlova, E. G.: Molecular mechanisms of regulation of functional activity of mononuclear phagocytes by leptin. *Biochemistry (Mosc)* **70**, 841–847 (2005).
77. Chao, T. C., Phuangsab, A., Van Alten, P. J., Walter, R.: Steroid sex hormones and macrophage function: Regulation of chemoluminescence and phagocytosis. *Am J Reprod Immunol* **35**, 106–113 (1996).
78. Kay, J., Czop, J. K.: Enhancement of human monocyte beta-glucan receptors by glucocorticoids. *Immunology* **81**, 96–102 (1994).

79. Tokuda, N., Mano, T., Levy, R. B.: Phagocytosis by the murine testicular TM4 Sertoli cell line in culture. *J Urol* **147**, 278–282 (1992).
80. Jones, C. J. P., Morris, K. J., Malcolm, I., Jayson, V.: Prednisolone inhibits phagocytosis by polymorphonuclear leucocytes via steroid receptor mediated events. *Ann Rheum Dis* **42**, 56–62 (1983).
81. Shimansky, O., Lisakovska, O. O., Mazanova, A. O., Riasniy, V. M., Veliky, M. M.: Effect of vitamin D3 and vitamin E on prednisolone-induced alterations of phagocyte function. *Eur Rev Med Pharmacol Sci* **20**, 1379–1383 (2016).
82. Ruiz, C., Montes, M. J., Abadia-Molina, A. C., Olivares, E. G.: Phagocytosis by fresh and cultured human decidual stromal cells: Opposite effects of interleukin-1 alpha and progesterone. *J Reprod Immunol* **33**, 15–26 (1997).
83. Baranau, R. I., Tenenbaum, A., Rumi, L. S.: Effects of sexual steroid hormones on the functionality of murine peritoneal macrophages. *Steroids* **56**, 481–485 (1991).
84. Tripathi, M. K., Singh, R.: Differential suppressive effects of testosterone on immune function in fresh water snake, *Natrix piscator*: An in vitro study. *PLoS One* **9**, e104431 (2014).
85. Yamaguchi, T., Watanuki, H., Sakai, M.: Effects of estradiol, progesterone and testosterone on the function of carp, *Cyprinus carpio*, phagocytes in vitro. *Comp Biochem Physiol C Toxicol Pharmacol* **129**, 49–55 (2001).
86. Watanuki, H., Yamaguchi, T., Sakai, M.: Suppression in function of phagocytic cells in common carp *Cyprinus carpio* L. injected with estradiol, progesterone or 11-ketotestosterone. *Comp Biochem Physiol C Toxicol Pharmacol* **132**, 407–413 (2002).
87. Law, W. Y., Chen, W. H., Song, Z. L., Dufour, S., Chang, C. F.: Differential in vitro suppressive effects of steroids on leukocyte phagocytosis in two teleosts, tilapia and common carp. *Gen Comp Endocrinol* **121**, 163–172 (2001).
88. Jutel, M., Blaser, K., Akdis, C. A.: The role of histamine in regulation of immune responses. *Chem Immunol Allergy* **91**, 174–187 (2006).
89. Kovács, P., Csaba, G., Csöreghe, É.: Influence of endocytosis-stimulating hormones on the protein binding capacity of the cell membrane. *Acta Physiol Hung* **61**, 213–216 (1983).
90. Csaba, G., Inczeffi-Gonda, Á.: Specificity of the dexamethasone-induced steroid receptor in *Tetrahymena*. *Experientia* **45**, 174–175 (1989).
91. Ferstl, R., Akdis, C. A., O'Mahony, L.: Histamine regulation of innate and adaptive immunity. *Front Biosci* **17**, 40–53 (2012).
92. O'Mahony, L., Akdis, M., Akdis, C. A.: Regulation of the immune response and inflammation by histamine and histamine receptors. *J Allergy Clin Immunol* **128**, 1153–1162 (2011).
93. Webster, J. I., Tonelli, L., Sternberg, E. M.: Neuroendocrine regulation of immunity. *Annu Rev Immunol* **20**, 125–163 (2002).
94. Csaba, G., Gaál, A., Kovács, P., Simon, G., Kőhidai, L.: Prolonged elevation of insulin content in the unicellular *Tetrahymena* after insulin treatment: Induction of insulin production or storage? *Cell Biochem Funct* **17**, 165–173 (1999).
95. Ferreira de Souza, A. M., Lopez, J. A.: Insulin or insulin-like studies on unicellular organisms: A review. *Braz Arch Biol Technol* **47**, 973–981 (2004).
96. Danilova, N.: The evolution of immune mechanisms. *J Exp Zool B Mol Dev Evol* **306**, 496–520 (2006).

97. Jacobs, M. E., DeSouza, L. V., Samaranayake, H., Perlman, R. I., Sui, W. M., Klobutcher, L. A.: The *Tetrahymena thermophila* phagosome proteome. *Eukaryote Cell* **5**, 1990–2000 (2006).
98. Boulais, J., Trost, M., Landry, C. R., Dieckmann, R., Levy, E. D., Soldati, T., Michnick, S. W., Thibault, P., Desjardins, M.: Molecular characterization of the evolution of phagosomes. *Mol Syst Biol* **6**, 423 (2010).
99. Csaba, G.: Hormonal actions in the Protozoan stress. *Acta Microbiol Immunol Hung* **62**, 331–339 (2015).
100. Csaba, G.: Lectins and *Tetrahymena* – A review. *Acta Microbiol Immunol Hung* **63**, 279–291 (2016).
101. Couleau, N., Falla, J., Beillerot, A., Battaglia, E., D’Innozenzo, M., Plancon, S., Laval-Gilly, P., Bennisroune, A.: Effects of endocrine disruptors on human macrophage-like THP-1 cell response. *PLoS One* **10**, e0131428 (2015).
102. Csaba, G.: Immunoendocrinology: Faulty hormonal imprinting in the immune system. *Acta Microbiol Immunol Hung* **61**, 89–106 (2014).
103. Dunbar, B., Patel, M., Fahey, J., Wira, C.: Endocrine control of mucosal immunity in the female reproductive tract: Impact of environmental disruptors. *Mol Cell Endocrinol* **354**, 85–93 (2012).
104. Liu, P., Wen, W. H., Song, X. X., Yuan, W. H.: [Effects of mixed cypermethrin and methylparathion on endocrine hormone levels and immune functions in rats: I. Dose-response relationship]. *Wei Sheng Yan Jiu* **35**, 257–260 (2006).
105. Bennisroune, A., Rojas, L., Foucaud, L., Goulaouic, S., Laval-Gilly, P., Fickova, M., Couleau, N., Durandet, C., Henry, S., Falla, J.: Effects of 4-nonylphenol and/or diisononylphthalate on THP-1 cells: Impact of endocrine disruptors on human immune system parameters. *Int J Immunopathol Pharmacol* **25**, 365–376 (2012).
106. Ho, S. M., Cheong, A., Adgent, M. A., Veevers, J., Suen, A. A., Tam, N. N., Leung, Y. K., Jefferson, W. N., Williams, C. J.: Environmental factors, epigenetics, and developmental origin of reproductive diseases. *Reprod Toxicol* **68**, 85–104 (2017).
107. Charles, M. A., Delpierre, C., Bréant, B.: Developmental origin of health and adult diseases (DOHaD): Evolution of concept over three decades. *Med Sci (Paris)* **32**, 15–20 (2016).
108. Csaba, G.: The biological basis and clinical significance of hormonal imprinting, an epigenetic process. *Clin Epigenetics* **2**, 187–196 (2011).