1	Epicellular coccidiosis in goldfish
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8	ABSTRACT: In a goldfish stock held in a pet fish pond, heavy coccidian infection, caused by

9 an epicellularly developing *Goussia* species, appeared in April of three consecutive years. The 10 shape and size of its oocysts resemble to an inadequately described species, Goussia 11 carassiusaurati (Romero-Rodrigez, 1978). In histological sections, gamogonic and 12 sporogonic stages infested mostly the second fifth of the intestine, where almost all epithelial 13 cells became infected. Both gamonts and young oocysts occurred intracellularly but in 14 extracytoplasmal position, seemingly outside the cells. Oocysts were shed unsporulated. Sphaeroid to ellipsoidal unsporulated oocysts measured 12.4×13.5 µm on average, but after 15 16 48 h sporulation in tap water they reached 16×13 µm oocyst size, in which the four elliptical 17 sporocysts of 13×5.4 µm located loosely. The size of oocysts and sporocysts are smaller than 18 those of the better known Goussia species, Goussia aurati (Hoffman, 1965).

19 KEY WORDS: Coccidiosis, Goussia, goldfish, epicellular location, seasonal development

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INTRODUCTION

22 Coccidia of the *Eimeria* and *Goussia* genera are common parasites of fish inhabiting 23 European natural waters and aquaculture farms. The genus *Goussia* was first described and 24 separated from *Eimeria* by Labbé (1896). Epicellular development of a *Goussia* species, *G.* 25 *pigra*, was first demonstrated by Léger & Bory (1932). Dyková & Lom (1981) created a new 26 genus *Epieimeria* for epicellularly developing fish coccidia selecting *Epieimeria anguillae* 27 (Léger et Hollande 1922) Dyková et Lom, 1981 as the type species and revitalized the genus 28 *Goussia* Labbé. Molnár (1989) found seasonally developing coccidia of nodular and 29 epicellular types in cyprinid fishes and described them as Goussia balatonica and G. 30 pannonica, respectively. Molnár (2006) regarded four genera common in fish, Eimeria, 31 Goussia, Crystalospora, Calyptospora as members of the Eimeriidae family. Several authors 32 among them Upton (2012) and Duszynski et al. (2016) still regard Goussia as synonym of 33 Eimeria. In recent papers (Molnár et al. 2012, Rosenthal et al. 2016) where molecular 34 sequences of different fish coccidia were examined, it was, however documented that great 35 differences exist among *Eimeria* and *Goussia* spp, more over within the members of *Goussia* 36 at least four groups can clearly be differentiated. Among Goussia spp. collected from 37 Hungarian fishes Rosenthal et al. (2016) found that molecular sequences of species with small oocysts developing dispersedly in the gut of their hosts throughout the year differ from 38 39 sequences of species with large oocysts developing seasonally in nodules and epicellular sites. 40 Great differences were also found between nodular and epicellular species. Species infecting 41 the kidney composed the fourth group.

42 Four coccidian species have been reported from the gut of goldfish. Goussia aurati 43 was described by Hoffman (1965) as *Eimeria aurati* from the gut of a goldfish shedding fecal casts with non- sporulated oocysts, which, once sporulated, measured $16-24 \times 14-17$ µm and 44 45 containing elliptical sporocysts measuring $11-13 \times 6.5-8 \mu m$. He found this species similar to 46 Goussia subepithelialis (Moroff & Fiebiger, 1906), the nodular coccidium of the common 47 carp, which according to data given by Steinhagen et al. (1990) have similar sizes with round 48 17-19 μ m oocysts and 12-14 \times 3 μ m sporocysts. Romero-Rodriguez (1978) found a similar 49 species, Goussia carassiusaurati (described as Eimeria) in goldfish with $15.2 \times 13.3 \,\mu m$ 50 oocysts and $13.6 \times 5.8 \ \mu m$ sporocysts. The third species Goussia hupehensis described by 51 Chen & Hsieh (1964) as *Eimeria hupehensis* is a typical dispersed coccidium resembling 52 closely in its morphology Goussia carpelli (Léger et Stankovitch, 1921). G. hupehensis (Chen & Hsieh, 1964) cit. by Chen & Li (1973) was found also in Hungary by Molnár et al. (2005) 53 54 who experimentally proved that this species (named only as Goussia sp.) cannot be 55 transmitted to the common carp. A fourth species, described as *Eimeria newchongensis* by 56 Chen (1984) from the gut of the goldfish and *Culter erythropterus* in China, by its 17.3-20 µm 57 sporulated oocysts, may not differ from G. auratii.

Here, we describe a coccidiosis in the gut of goldfish caused by epicellularly developingcoccidian stages.

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MATERIALS AND METHODS.

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64 Goldfish (Carassius auratus gibelio (L) are frequently cultured in small hobby ponds in 65 Hungary, as well as in aquaria. From one of these ponds where about 100 specimens of 1 to 5 years old goldfish and their about 1000 offspring was cultured some fish were sent to 66 laboratory for health control at about in monthly intervals. Fish were collected from the pond 67 68 by a hand net, carried to the laboratory alive in oxygenated plastic bags, kept in aerated 69 aquaria. They were sedated with 20 ppm clove oil added to the water and exterminated by cutting 70 their head. Coccidian infection of varying intensity with small sized oocysts of the dispersed 71 G. hupehensis species was recorded throughout the examination period. In addition in Aprils 72 of three consecutive years heavy infection with large sized oocysts resembling epicellular and 73 nodular coccidia, common at this time in natural water fishes (Molnár 1989), were recorded. 74 In April, 2016 a specific survey for coccidian infections was performed on twenty five specimens of 1 to 3 years old goldfish specimens measuring 4 to 13 cm. The gut was opened 75 76 lengthwise under a dissecting microscope and cut into five equal parts. From the epithelial 77 surface, pieces of clean mucus were removed, placed under coverslip, and studied with a 78 Nomarski differential interference contrast with an Olympus BH2 microscope. Some non-79 sporulated oocysts were placed in tap water into a small Petri dish and left to sporulate for 48 80 hours. Histological sections were prepared in 2016. In April 5, 15, and 25 the gut of the 81 smallest fish, about 4 cm in length, was separated and coiled into a roll, fixed in Bouin's 82 solution for four hours, embedded in paraffin wax, cut to 4-5 µm sections, and stained by 83 haematoxylin and eosin. Non-sporulated and sporulated oocysts were studied using Nomarski 84 differential interference contrast with an Olympus BH2 microscope. The oocysts and 85 histological sections were photographed with an Olympus DP 20 digital camera. From 86 oocysts digitised images were obtained and measurements were taken with the IMAGO® 87 software.

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RESULTS

94 Heavy infections with non-sporulated oocysts were found in April in each year. At this time the gut of these fish was empty or contained only very little faeces. The fish were in poor 95 96 condition but no mortality was recorded. No mortality was recorded on such fish subsequently 97 held in laboratory aquarium for about a month. All of the examined fish proved to be infected. 98 When freshly dissected, the second and fifth sections of the intestine were found to be most 99 heavily infected with gamogonic and sporogonic stages. Relatively little infection was 100 recovered in the first and third sections of the gut. From the last two sections, only non-101 sporulated oocysts were recovered and these were located free in the lumen of the gut or in 102 faces. Oocysts were shed unsporulated. Most of the unporulated oocysts had a granulated 103 structure and spheroid or short ellipsoidal shape (Fig. 1). In the first half of April, mostly 104 developing gamonts were found in the first part of the intestine (Fig. 2). In the middle of April 105 only non-sporulated oocysts were found in infected sections of the gut. By the end of April 106 most oocysts had left the gut and only sparse oocysts were found in the rectum among faces. 107 In tap water at 22-23 °C, non-sporulated oocysts became sporulated in two days. Sporulated 108 oocysts had also spheroid or short ellipsoidal shape (Fig. 3), a very thin, single layered and colourless wall. The oocysts (Table 1) measured 12.4±0.86 (12.8-15.2) µm in length and 109 110 13.5 ± 1.8 (12-14.6) µm in width. There was no oocyst residuum, but a small polar body existed in oocysts. The four thin-walled, elongated ellipsoidal sporocysts located loosely in 111 112 the oocysts, measured 13 ± 0.8 (11.2-15.4) µm in length and 5.4 ± 0.34 (4.5-5.6) µm in width. In 113 each sporocyst there were two vermiform sporozoites arranged head to tail. Sporozoites 114 measured 10.3±0.73 (9.6-11.2) µm in length and 2.5±0.32 (2.4-2.8) µm in width. In freshly sporulated sporocysts, the sporocyst residuum covered most parts of the sporozoites, after 48 115 116 hours sporulation, this residuum became short and ellipsoidal, an about 2.5×1.3 compact 117 body, and it located in the sporocysts among sporozoites.

118 In histological preparations fixed in the first half of April, heavy infection with young 119 gamogonic stages was found in the second and third fifths of the intestine, while developing oocysts were only rarely found in the first fifth, and they were not found in the fourth and fifth
segments of the gut. At this time, most of the gamonts were young, measuring about five µm
in diameter. In some segments of the intestine, they infected almost every epithelial cell (Fig.
Less frequently, microgamonts were also found (Fig. 5). Development of gamogonic and
sporogonic stages was not synchronous, as mature, unsporulated oocysts were found among

125 developing gamonts (Fig. 6).

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DISCUSSION

128 Although the genus *Goussia* is still regarded by many authors synonym to *Eimeria* (Upton 129 1912, Duszynski et al. 2016), others working on eimerians of fish and anurans accept its 130 validity (Lukeš & Dyková 1990, Steinhagen et al. 1990, Lukeš 1992, Jirkú et al. 2002, 2009, 131 Dogga et al. 2015). Recent phylogenetic works (Molnár et al. 2012, Rosenthal et al. 2016) 132 have validated deep evolutionary distinctions among the coccidian parasites of fish, including 133 (but not limited to) a monophyletic assemblage limited to true members of the genus *Eimeria*; 134 subdivisions within Goussia correspond to morphological and developmental attributes that 135 likely warrant further taxonomic revision. Several Goussia species develop in nodules in the 136 gut like G. subepithelialis, while others develop in epicellular position in the enterocytes like 137 G. pigra. Molnár (1989) who described G. balatonica and G. pannonica spp. from the white 138 bream, remarked that in several cyprinids a mixed infection with nodular and epicellular type 139 coccidia occur in the time of oocyst formation in April. Morphologically similar oocysts 140 which, however, differ in size also occur in non-cyprinids; these include Goussia aculeati, 141 and G. zarnowskii in Gasterosteus aculeatus (L.) and G. acipenseris and G. vargai in 142 Acipenser ruthenus (L.) (Jastrzebski 1984, Molnár 1986). It is not excluded therefore, that 143 epicellular and nodular coccidia exist concurrently also in the common carp and the goldfish. Both fish species is of Chinese origin which introduced only a smaller part of their parasite 144 145 fauna to Europe. The smaller (in average 16 µm) oocyst size measured for samples of the 146 goldfish in the present investigation resembles to epicellular G. pannonica studied by Molnár 147 (1989), and G. janae described by Lukeš & Dyková (1990), while G. aurati, with its 16×20 148 µm large oocysts described by Hoffman (1965), correspond better to a nodular species. 149 Although the above facts might suggest the nodular nature of G. aurati and the epicellular 150 nature of G. carassiusaurati present data give not enough basis for drawing this conclusion.

151 Similar problems exist at the common carp. To date, only a nodular species, *G.* 152 *subepithelialis*, has been reported from this fish. It cannot be ruled out, however, that a more 153 detailed examination in China, in the original biotope of the common carp and the gold fish 154 both nodular and epicellular species could be recorded.

155 Epicellular development characterizes a particular lineage of coccidia, which adds weight to 156 the validity of the genus Epieimeria Dyková & Lom, 1981. On the other hand, the great 157 difference in morphology, development and molecular sequences between Goussia type 158 epicellularly developing coccidia and the type specimen Epieimeria anguillae contradicts a 159 relationship between the two eimerian groups. Differences in the development (permanently developing group vice versa annually developing groups), intracellular location 160 161 (intracytoplasmal and epicytoplasmal), more over differences in the molecular sequences 162 among dispersed, nodularly and epicellularly developing groups (Rosenthal et al. 2016) would 163 favour creation of new coccidian genera in the future, among them a genus with a tentative 164 name Epigoussia.

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- 174 *Conflict of interest*
- 175 The authors declare that they have no conflict of interest.
- *Ethical approval* All applicable institutional, national and international guidelines for the care and use of animals
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LITERATURE CITED

182

181

- 183 Chen, CL (1984) Sporozoa of fishes from Liao He (Liaoho River) of China. In: Parasitic
 184 Organisms of Freshwater Fish of China, Edited by the Institute of Hydrobiology
 185 Academia Sinica, Agricultural Publishing House, Beijing. pp. 3-21.
- Chen CL, Li, WW (1973) An Illustrated Guide to the Fish Disease and Causative Pathogenic
 Fauna and Flora in the Hupei Province. Publishing House Science, Institute of
 Hydrobiology, Wuhan, China. 456 pp.

- 7
- 189 Dogga, SK, Bartosova-Sojkova P, Lukes J, Soldati-Favre D (2015) Phylogeny, Morphology,
 190 and Metabolic and Invasive Capabilities of Epicellular Fish Coccidium *Goussia* 191 *janae*. PROTIST 166:659-676.
- 192 Duszynski DW, Couch L, Upton SJ (2016) Coccidia of the World. Available at:
 193 http://biology.unm.edu/biology/coccidia/home.html
- 194 Dyková I, Lom J (1981) Fish coccidia: critical notes on life cycle, classification and
 195 pathogenicity. J Fish Dis 4:487–505.
- Hoffman GL (1965) *Eimeria aurati* n. sp. (Protozoa: Eimeriidae) from Goldfish (*Carassius auratus*) in North America. J Protozool 12:273-275.
- Jastrzebski M (1984) Coccidiofauna of cultured and feral fishes in farms. Wiadomosci
 Parazytologiczne 30:144-160.
- Jirkú M, Jirkú M, Obornik M, Lukeš J, Modry D (2009) *Goussia* Labbé, 1896 (Apicomplexa,
 Eimeriorina) in Amphibia: Diversity, Biology, Molecular Phylogeny and Comments
 on the Status of the Genus Protist 160:123-136.
- Jirkú M, Modry D, Slapeta JR, Koudela B, Lukeš J (2002 The phylogeny of *Goussia* and
 Choleoeimeria (Apicomplexa; Eimeriorina) and the evolution of excystation
 structures in coccidia. Protist 153:379-390.
- 206 Labbé A (1896) Recherches Zoologiques, Cytologiques et Biologiques sur les Coccidies.
 207 Arch Zool Exp Gen 4:517–654.
 208
- Léger L, Bory T, (1932) *Eimeria pigra* n. sp. nouvelle coccidie juxtaépitheliale parasite du gardon rouge. C R Hebd Séanc Acad Sci 194:1710–1712.
- Lukeš J (1992) Life cycle of *Goussia pannonica* (Molnár, 1989) (Apicomplexa, Eimeriorina),
 An extracytoplasmic coccidium from the white bream *Blicca bjoerkna* J Protozool
 39:484-494.
- Lukeš J, Dyková I (1990) *Goussia janae* n-sp (Apicomplexa, Eimeriorina) in dace *Leuciscus leuciscus* and chub *Leuciscus cephalus*. Dis Aquat Org 8:85-90.
- Molnár K (1986) Occurrence of two new *Goussia* species in the intestine of the sterlet
 (Acipenser ruthenus). Acta Vet Hung 34:169-174.
- Molnár K (1989) Nodular and epicellular coccidiosis in the intestine of cyprinid fishes. Dis
 Aquat Org 7:1-12.
- Molnár K (2006) Phylum Apicomplexa. In: P.T.K. Woo (ed.): Fish Diseases and Disorders.
 Vol. 1: Protozoan and Metazoan Infections. Second edition. CAB International.
 Wallingford. pp. 183-204.
- 223 Molnár K, Ostoros Gy, Baska F (2005) Cross-infection experiments confirm the host

224 225 226	specificity of <i>Goussia</i> spp. (Eimeriidae: Apicomplexa) parasitizing cypridid fish. Acta Protozool 44: 43-49.	
220 227 228 229	Molnár K, Ostoros Gy, Dunams-Morel D, Rosenthal BM (2012) <i>Eimeria</i> that infect fish are diverse and are related to, but distinct from, those that infect terrestrial vertebrates. Infect Gen Evol 12:1810-1815.	
230 231	Romero-Rodrigez J (1978) Coccidiopatias de peces, estudio del Protozoa Eimeriidae: <i>Eimeria carassiusaurati</i> , n. sp. Rev Iber Parasitol 38:775-781.	
232 233 234	Rosenthal BM, Dunams-Morel D, Ostoros Gy, Molnár K (2016) Coccidian parasites of fish encompass profound phylogenetic diversity and gave rise to each of the major parasitic groups in terrestrial vertebrates. Infect Gen Evol 40: 219-227.	
235 236 237	Upton SJ (2012) Suborder Eimeriorina Léger, 1911. In: Lee, J.J., Leedale, G.F., Bradbury, P. (Eds.), The Illustrated Guide to the Protozoa, second ed. Soc Protozoolog, Laurence, Kansas, pp. 318–339.	
238 239 240	Steinhagen D, Lukeš J, Körting W (1990) Ultrastructural observations on gamogonic stages of <i>Goussia subepithelialis</i> (Apicomplexa, Coccidia) from common carp <i>Cyprinus</i> <i>carpio</i> . Dis Aquat Org 9:31-36.	
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