Egg spotting pattern in common cuckoos and their great reed 1 warbler hosts: a century perspective 2 3 NIKOLETTA GELTSCH^{1,2}, CSABA MOSKÁT^{1*}, ZOLTÁN ELEK¹, MIKLÓS 4 BÁN³, MARTIN STEVENS⁴ 5 6 ¹ MTA-ELTE-MTM Ecology Research Group, a joint research group of the Hungarian 7 Academy of Sciences, the Biological Institute of Eötvös Lóránd University and the Hungarian 8 9 Natural History Museum, Hungary ² Department of Ecology, University of Szeged, Szeged, Hungary 10 ³ MTA-DE 'Lendület' Behavioural Ecology Research Group, Department of Evolutionary 11 Zoology, University of Debrecen, Debrecen, Hungary 12 ⁴ Centre for Ecology and Conservation, College of Life and Environmental Sciences, 13 University of Exeter, Penryn Campus, Penryn, Cornwall, TR10 9FE, UK. 14 15 * Corresponding author. E-mail: moskat@nhmus.hu 16 17 Word count: 7,907 words 18 Short title: changes in egg spottiness 19 20 ADDITIONAL KEYWORDS: brood parasitism, coevolution, common cuckoo, digital image 21 22 analysis, egg pattern, egg spottiness 23 24 The characteristics of common cuckoo (Cuculus canorus) and host eggs are widely thought to 25 have coevolved over time, but few studies have tested this prediction. We compared cuckoo 26 eggs with those of its primary host, the great reed warbler (Acrocephalus arundinaceus) from four time periods spanning >100 years (between 1900 and 2014), and studied if cuckoo 27 eggshell patterns better resembled those of their hosts over time. We used image analysis to 28 compare five eggshell pattern variables, relating to marking size, diversity, contrast, coverage, 29 30 and distribution on the egg surface. Each feature showed different temporal trends. All but

one of these variables ("dispersion" of spots among egg regions) were species-specific and

differed between hosts and parasites. The magnitude of change was greater for hosts than

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cuckoos, which could be a consequence of host eggs' more intensive and variable spottiness. Specifically, the proportion of the egg surface covered with pattern increased marginally over time, and the dispersion of spotting became more even over the egg surface. Egg marking contrast showed a decreasing trend, with species differences also decreasing, suggesting better mimicry. Our results suggest multidirectional evolution of eggshell components in this system, with potential implications for mimicry and rejection over time.

INTRODUCTION

Brood parasitic birds lay their eggs in the nests of other avian species, and these foster parents incubate the foreign eggs and feed and rear the foreign nestlings (Rothstein, 1990). During long-term coevolutionary arms races among brood parasites and their hosts, selection pressures drive adaptations of the brood parasites and counter-adaptations of hosts (Davies, 2000). The common cuckoo (*Cuculus canorus*; hereafter "cuckoo") is a well-studied brood parasite (Schulze-Hagen, Stokke & Birkhead, 2009), comprising several different host-races (often called "gentes"), in which each female cuckoo specializes on a particular host species. Many hosts have evolved the ability to recognize alien eggs and eject them from their nests, or desert their clutch entirely, and so many female cuckoos deceive host parents through laying eggs that mimic those of their host, making recognition of foreign eggs more difficult.

The eggshells of many bird species are highly variable in colour and spotting among individuals (Underwood & Sealy, 2002; Cassey et al., 2010b; Spottiswoode & Stevens, 2010; Hauber, 2014), and brood parasitism is one of the most important selective pressures in modifying these phenotypic traits (Øien, Moksnes & Røskaft, 1995; Soler & Møller, 1996; Kilner et al., 2006). The hosts' main anti-parasite defence mechanism, egg rejection behaviour, is a complex process (e.g. Pozgayová et al., 2011; de la Colina et al., 2012), where background coloration plays a key role in foreign egg recognition by hosts (e.g. Avilés, 2008; Cassey et al., 2008). In contrast, the function of spottiness has less frequently been studied compared to colour, but hosts also likely use various characters of spotting in egg recognition, for example in identifying foreign eggs (Moskát et al., 2008a), or looking for signatures indicating own egg identity (Stoddard, Kilner & Town, 2014), especially those encoded at the blunt pole of the eggs (e.g. Polaciková & Grim, 2010).

Most studies on cuckoo-host coevolution have concentrated on 'snapshots' of the arms race, covering a duration of only a few years, or with highly variable sampling periods. Even when studies are based on eggs stored in museum collections (e.g. Avilés & Møller, 2003; Stoddard & Stevens, 2010), datasets are rarely suitable for detecting longer-term trends (but see Spottiswoode & Stevens, 2012). As such, there is a pressing need for long-term studies on the coevolution of egg colour (e.g. Cherry & Bennett, 2001; Antonov et al., 2010) and pattern mimicry (Medina et al., 2016; Brulez et al., 2016). Previous modelling has predicted that cuckoos should over time benefit from increasing the resemblance of their own eggs to those of the host, and that hosts should be under selection to escape parasitism (Takasu, 2003). Therefore, the selection pressure derived from brood parasitism may promote host eggs in a given population to change phenotypes over time (Takasu et al., 2009; Spottiswoode & Stevens, 2012). Consequently, we cannot simply expect a continuous improvement in cuckoo egg mimicry to host egg phenotypes in a host-brood parasite relationship, but instead might expect more of a Red Queen scenario of both parties evolving changes in phenotype over time; the host to escape cuckoo mimicry, with the cuckoo in turn under selection to keep up.

In Hungary, the great reed warbler (Acrocephalus arundinaceus) is the main host of the cuckoo in wetland habitats (Moskát & Honza, 2002). The eggs laid by great reed warblerspecific cuckoos in Hungary are often cited as one of the most impressive cases of mimicry of spotted, non-plain coloured cuckoo eggs found (e.g. Southern, 1954; Davies & Brooke, 1991; Moksnes & Røskaft, 1995). The high cuckoo parasitism on great reed warblers has been documented in detail from the Hungarian population (Zölei, Bán & Moskát, 2015), with eggs collected from the early 1900's available for study in museums, offering an exceptional possibility for revealing how cuckoo egg mimicry has changed over a more than 100-year period. Here, we used digital images of the eggs of cuckoos and great reed warblers to evaluate how the pattern of spottiness, as an important component of egg mimicry (Stoddard & Stevens, 2010), changed from the 1900's until the present day. In order to quantify egg pattern mimicry between brood parasite and its host, we used the method of Stoddard & Stevens (2010), based on digital image analysis of several components of egg patterns (Table 1). We hypothesized that cuckoos have been under selection pressure to improve their eggshell pattern match to that of their hosts, and so we investigated trends in eggshell pattern components during approximately the last hundred years. We predicted that mimicry (defined as the numerical difference between the egg parameters studied) has improved continuously during this period. As an alternative hypothesis, we predicted that the arms race between the cuckoo and great reed warbler was in a steady state during the studied period, resulting in a

stable relationship or small fluctuations in egg phenotypes. We predicted no significant change in similarity of cuckoo and host eggs in this case.

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We photographed parasitized clutches of great reed warblers collected from the central regions of Hungary contained in the egg collection of the Natural History Museum (Tring, Hertfordshire, UK) and the Mátra Museum (Gyöngyös, Hungary, a branch of the Hungarian Natural History Museum, Budapest). We studied only spotting pattern of eggs stored in dark places, under museum conditions, not exposed to direct sunlight. Changes in colours over a long time could be more relevant when eggshell colours rather than spottiness are studied, as previous studies showed some changes in the background colour of eggshells, mostly in terms of blue-green chroma (Cassey et al., 2010a; Navarro & Lahti, 2014). In addition, we took pictures in the field of parasitized clutches in central Hungary, ca. 50 km south of Budapest, in the vicinity of the village Apaj (47°07'N; 19°06'E). All clutches were photographed on a Kodak Grey Card with a digital camera Olympus E-510 (Olympus Imaging Corp., Tokyo, Japan). Kodak Colour Control Patches were also placed near the eggs on each photo as a reference. We took pictures of clutches containing one cuckoo egg (single parasitism). All eggs were collected from 1901 to 1969 and field images were taken in 2014. Four stages are characterized in this approximately 100-year period: (i) the "1900s" (1901-1903, n = 16), (ii) the "1930s" (1934-1939, n = 16), (iii) the "1960s" (1960-1969, n = 12) and (iv) "2014" (n = 16) 20). We photographed complete clutches (4, 5 or 6 eggs), but randomly chose one host egg per clutch for the following analyses (Fig. 1). To prevent using more than one clutch from the same female, we included different sites and years, and in the recent dataset we took care not to use replacement clutches built after predation. Altogether, 128 eggs from 64 nests were measured and used in the analyses (64 cuckoo and 64 host eggs). All clutches were used in the analysis; no preliminary selection was applied on the similarity of cuckoo eggs to host eggs (in order to attempt to identify cuckoo gentes) so that we could represent the available morphs of cuckoo eggs, similarly to other studies on spotting pattern (e.g. Stoddard & Stevens, 2010). In general, interclutch variation of great reed warblers is high in our study area (Moskát & Honza, 2002), and there are corresponding cuckoo egg morphs for most of

the common and some of the rarer egg types, but cuckoo eggs belonging to other cuckoo gentes may also occur (c.f. Lovászi & Moskát, 2004).

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PATTERN ANALYSIS

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Quantifying spottiness, at least as revealed through human scoring, seems to be an important aspect of cuckoo-host coevolution (Moksnes & Roskaft, 1995). Although some previous studies have used computer programs, e.g. NIH-Image or Scion Image for Windows, for image analysis of eggshell spottiness (Soler J, Soler M, Møller, 2000; Moskát, Szentpéteri & Barta, 2002), these were influenced by subjective decisions by humans and based on uncalibrated images. However, images from most cameras require calibrating before they should be used, to control for differences in light conditions and the camera-specific response to light intensity (see Stevens et al., 2007). Here, we first linearized our images to correct for the camera's non-linear response to light levels (radiance), and normalised the images with regards to the grey standard to control for light conditions, thus deriving images corresponding to reflectance information (Stevens et al., 2007). We then applied the method used by Stoddard & Stevens (2010), based on digital image analysis to quantify several aspects of egg pattern. All calibrations and image analyses were carried out by self-developed computer programmes written in MATLAB (The MathWorks, Inc., MA, USA) and its associated Image Processing toolbox. As with previous work, we took the mediumwave ('green') image channel for analysis because this most closely approximates to avian achromatic vision, which likely primarily underlies pattern perception (see Spottiswoode & Stevens, 2010). Before quantifying the eggs' pattern, we rescaled each image to 50 pixels/mm in ImageJ (Abramoff, Magelhaes & Ram, 2004), because our photographs were taken at slightly variable distances (Spottiswoode & Stevens, 2010). We used the length of the yellow segment of Kodak Colour Control Patches as a reference for this.

During the analyses, each calibrated image of an egg was filtered into a set of new images using Fast Fourier Transformation and seven octave-wide, isotopic band-pass filters (Barbosa et al., 2008; Stoddard & Stevens, 2010; Spottiswoode & Stevens, 2010). These new images include information regarding pattern at different spatial scales, which can be thought of as acting analogous to a set of sieves, each image capturing information for different sized markings from smaller sizes (high spatial frequency) to larger sizes (low spatial frequency). These seven filtered images or 'granularity bands' (Barbosa et al., 2008) contain information about the relative importance of different marking sizes in contributing to the overall pattern

(marking filter size, referred to as "filter size"). From each of the seven granularity bands we calculated the overall pattern 'energy' (total energy), as the sum of the squared pixel values in each image divided by the number of pixels in the image (Chiao et al., 2009; Stoddard & Stevens, 2010). The total energy of the spectrum corresponds to the overall amplitude of the spectrum, and provides a measure of pattern contrast of the egg markings (Stoddard & Stevens, 2010; Spottiswoode & Stevens, 2010). This variable expresses how contrasting the markings are against the egg background colour ("total energy"; see Table 1). These seven values produce a 'granularity spectrum', where the maximum energy value corresponds to the filter size containing the highest energy, and thus the dominant marking filter size ("filter size"). We also calculated the proportion that this maximum value contributes to the total energy of the spectrum ("proportion energy"), which indicates the importance of the dominant marking size to the overall egg pattern. A high value shows that the egg pattern is dominated by just one or a few filter sizes (Stoddard & Stevens, 2010; Spottiswoode & Stevens, 2010).

In addition, we calculated the relative proportion of three main egg regions covered by markings, as opposed to the base colour (simply referred as pattern "coverage"). Methods again closely followed Stoddard & Stevens (2010). First, we thresholded each image into a binary format, whereby markings are encoded by one (1.00) and background egg colour encoded by a zero (0.00). From this, the proportion of the total pixel values that corresponded to a marking was calculated. This value shows the overall proportion of the egg that is covered with markings (Stoddard & Stevens, 2010; Spottiswoode & Stevens, 2010). We also calculated these metrics for three regions of the egg surface, corresponding approximately to the top (narrow), middle, and base (wider) thirds of the egg. The standard deviation of pattern coverage among the bottom, middle, and top regions of an egg is a measure of pattern dispersion (referred as "dispersion" in analyses), which indicates how concentrated the pattern is at the bottom of the egg. As we revealed differences among egg regions (see Results), we therefore considered the egg regions separately in our linear models, and called this division as "topology". Previous studies failed to find differences among the markings of eggshell regions and averaged measurements for the overall egg surfaces (e.g. Stoddard & Stevens, 2010). Other studies looking for markings of the eggshell in cuckoo hosts revealed the importance of the bottom region (blunt pole) over the top region (sharp pole), as hosts primarily used the information content of the blunt pole for recognition (Polaciková et al., 2007, 2010, 2011; Polaciková & Grim, 2010). While there are other potential methods that could be used to quantify egg pattern mimicry, including a recent computer science approach of Stoddard et al. (2014), the metrics we use here have been shown to effectively predict egg

200	rejection behaviour in experimental studies in other parasite-host systems and are broadly
201	based on principles of early spatial vision processing (Spottiswoode & Stevens, 2010, 2011;
202	Stevens, Troscianko & Spottiswoode, 2013).
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204	Data analysis
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206	Comparison of spottiness
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208	Response variables (see below) were tested among years (factor with 4 levels: 1900, 1930,
209	1960 and 2014), species (factor with 2 levels: GRW as great reed warbler, C as cuckoo) and
210	topology (i.e. egg regions, factor with 3 levels: bottom, middle and top). We also included
211	their interactions, and the estimation was done using the ordinary least square mean method.
212	The following response variables were used (and their abbreviations): (1) proportion energy,
213	(2) filter size, (3) the common logarithm of filter size (log filter size), (4) coverage, (5) total
214	energy and (6) dispersion. These responses were tested by a single-argument ANOVA
215	(permutational ANOVA), where the number of iterations was taken to generate exact P values
216	(Box, 1988). The description of identification of overall trends by time series analyses in the
217	eggshell's pattern is available in the ESM section no. 1. We used two datasets in the analyses,
218	in the first case all parameters were considered according to the species (great reed warbler,
219	'GRW' and cuckoo, 'C'), while in the second case the egg parameters were defined as a
220	differences between the species, thus the effectiveness of the mimicry can be tested. We refer
221	to this dataset as 'mimicry data' in the text below.
222	All analyses were carried out in R 3.1. 2 (R Core Team, 2014) using the package
223	lmPerm for permutational ANOVA (Wheeler, 2010) and TTR for time series analyses (Ulrich,
224	2013).
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227	RESULTS
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229	COMPARISON OF SPOTTINESS
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231	In the first step of our analyses we compared egg parameters among the top, middle, and
232	bottom regions of the eggs. As we found differences among them (c.f. topology effects in

Table 2), we could not combine all regions of the eggs into one operation unit for subsequent analyses. Permutational ANOVA revealed different effects for all five variables measured (Table 2). All variables, except filter size (even when log transformed), showed a year effect. Topology effect (the effects of egg regions) was revealed for coverage and contrast. Compound effect ('species x topology') was significant for proportion energy and contrast. The variable proportion energy slightly increased over time with consistently higher values for great reed warblers than cuckoos, while dispersion showed the opposite trend (Table 2 and Fig. 2).

We found increasing trends for the variables coverage and total energy. However, the values of these parameters were consistently higher for great reed warblers than cuckoos (Table 2 and Fig.2). The metrics coverage and total energy showed similar trends for both of the two species (Fig. 2). Based on the above mentioned issues for egg's topology, we suggest that the source of differences is the species itself rather than any topological differences between egg's regions.

We also analyzed mimicry (the difference between the two species' values for each variable) for only those pattern metrics where significant effects were revealed above. Interestingly, most variables did not show a year effect – only total energy showed a significant year effect, a decreasing trend (p = 0.004), and differences among egg parts (topology: p = 0.008; Table 3 and Fig. 3). As the difference decreased between cuckoo and host eggs regarding this variable, this means that mimicry improved with regards to total energy.

ESTIMATING TRENDS IN TIME

We found a clear and distinctive seasonal pattern in the egg parameters based on the decomposition of time series between 1900 and 2014. The optimum ARIMA model proved to be the most useful, with 2, 2, and 5 parameters for the eggs' data. The residuals indicated small variation around the mean zero; none of them was greater than the double of the standard deviations, thus this model provided the best fit. The autocorrelation of the residuals was not significantly different from zero as a set, and had a constant variance, thus confirming the adequacy of the model for all egg parameters. The decomposition of time series data (summarized in ESM as SFigs. 1-4) revealed that (those parameters which were affected significantly by the year) the parameter proportion energy showed consistent changes in its pattern over the period studied, while total energy showed a decreasing trend in time for both

species. For the mimicry data, this latter variable also showed a similar decreasing trend (Fig. 4). As we defined mimicry as the difference between host and parasitic eggs, this suggests that mimicry was improving with time.

DISCUSSION

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Here, we compared egg pattern components of cuckoo-host egg mimicry at four stages over a period of more than 100 years. Such data are scarce in studies of the evolutionary biology between hosts and brood parasites. We evaluated five parameters of eggshell spottiness of great reed warblers, cuckoos, and calculated their resemblance (indicative of 'mimicry') over time. Although great reed warbler and cuckoo eggshell patterns are widely thought to have coevolved, each of the five characters showed different trends, except filter size (equating to the main egg marking size), which did not show any changing pattern with time. In general, the magnitude of these changes was greater for great reed warblers than cuckoos, which could be a consequence of their more intensive and variable spottiness (see similar results in Spottiswoode & Stevens, 2012 for another host-brood parasite relationship). These results could be explained by stronger selection pressures on hosts to evolve signatures by which they can recognise their eggs than on cuckoos to match their eggs (M.I. Cherry, pers. comm.). Interestingly, a recent study on cuckoos and great reed warblers in Hungary pointed to the stability of this host-brood parasitic relationship regarding parasitism rate and hosts' responses to parasitism in the last 70 years (Zölei et al., 2015). In the present study, most variables showed no difference between species, with the exception of coverage and total energy. For the latter two characters, our results provided support for the assumption that brood parasitism is a selective force in cuckoo-host coevolution and egg phenotypes. Hosts of brood parasites only use some aspects of egg appearance in guiding their egg rejection behaviour at any one time, and those features used might be the ones that differ most between parasites and hosts (e.g. Spottiswoode & Stevens, 2010, 2011). Therefore, it may be that at different stages (time periods) of the arms race different aspects of egg appearance are used in rejection and therefore under selection. Other traits may not be under selection at the same time.

Although we do not know exactly when this parasitic relationship started, our host-brood parasite system seems to be a relatively well-stabilized one as the main characteristics, such as high parasitism rate, high rate of multiple parasitism, and mid-level rejection rate,

seem to be consistent over time (see Zölei et al., 2015). The study by Zölei et al. (2015) also suggests that despite the unusually and permanently high parasitism rate found in our study area for at least the last 70-80 years, our host-brood parasite system is in dynamic evolutionary equilibrium, where the ratio of hosts accepting or rejecting parasitic eggs is in equilibrium (Lotem & Nakamura, 1998), and the recognition error rate is low (Stokke et al. 2016). This could explain the lack of one universal trend in the changes of eggshell spotting characters revealed in the present study. Newly established cuckoo-host relationships are expected to be rather unstable, as both empirical and modeling studies have revealed. For example, they may show well-documentable changes in time, either in parasitism rate (Takasu et al., 1993; Barabás et al., 2004), egg appearance (Avilés et al., 2006), or egg discrimination of hosts (Røskaft et al., 2002, 2006). Sometimes, phenotypic changes can be detectable within a shorter period when strong selection pressures affect coevolutionary changes (Spottiswoode & Stevens, 2012), and phenotypic traits may oscillate around fixed points in the phenotypic space and result in discrete categories of egg phenotypes (Takasu, 2003, 2005).

Our results revealed multidirectional changes in coevolutionary adaptations in eggshell spottiness, which is in accordance with previous results that eggshell pigments are likely to have several functions (Maurer, Portugal & Cassey, 2011), suggesting multidirectional evolution of different eggshell components (Gosler, Higham & Reynolds, 2005). For example, eggshell pigments may protect embryos from solar radiation (Lahti, 2008). Furthermore, while some eggshell characteristics are heritable, others may also be affected by the temporary and local availability of chemical and dietary components necessary for pigment synthesis (Hubbard et al., 2010). Rainfall (Avilés et al., 2007), temperature (Honza, Procházka & Pozgayová, 2012; Hargitai et al., 2016), food type (Moreno et al., 2006; Hargitai, Herényi & Török, 2008), and light exposure (Navarro & Lahti, 2014) also could be factors leading to changes in population trends of egg appearance. Berkowic et al. (2015) revealed that cuckoo and host egg sizes also changed during a longer period. Their study pointed to climatic factors influencing host-brood parasite egg morphs primarily being driven by coevolution. The complexity of coevolution of host and brood parasitic eggs is also demonstrated by Cherry, Bennett & Moskát (2007a). They revealed a weak matching process of host and cuckoo eggs by cuckoos when they lay; i.e. cuckoos less frequently selected host nests where host eggs differed from their own egg type. In this way cuckoos were selective for clutch appearance when they laid, and consequently, they slightly increased the matching efficiency of their eggs (see also Honza et al., 2014 for a similar result in the same host species). However, regarding the unusually high parasitism rate in our study area (frequently

over 50%, Zölei et al., 2015, instead of the normal 5-10%, Davies, 2000), this weak selection process cannot always prevent host clutches from receiving poorly-matched cuckoo eggs (see Fig. 1).

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Although the eggs of some of the host species and their corresponding cuckoo eggs are immaculate blue, pale blue, or white (e.g. Yang et al., 2014), the eggs of most species are speckled with dark spots (Moksnes & Roskaft, 1995). Such eggshell pigments have a role for coding information regarding the ownership of the eggs (Stevens, 2013), as signatures for individual recognition of eggs by nest owners (Stoddard et al., 2014), or the lack of spotting may also be indicative on the parasitic status of the eggs (de la Colina et al., 2012). Selection may favour the development of these egg signatures (Swynnerton, 1918), which may be of help in the discrimination of own eggs (Moskát & Hauber, 2007). Typically, enhanced egg discrimination is observed in hosts of avian brood parasites and brood parasitism is an important factor in the evolution of such traits (Stokke, Moksnes & Røskaft, 2002; Kilner, 2006; Igic et al., 2012; Thorogood & Davies, 2013). For example, in an experimental study on great reed warblers' egg discrimination, Moskát et al. (2008a) revealed that density of small spots (speckling) affected recognition of parasitic eggs. However, large spots, either in low or high density, only moderately enhanced egg rejection, except when the whole eggshell surface is covered with these spots, at which point rejection rates abruptly increased (Moskát et al., 2008b; see also Hauber et al., 2014 for similar results on two thrush species, Turdus spp.). Interestingly, cuckoo eggs have smaller spot sizes than great reed warbler eggs, something that is clearly visible to humans at first sight and which was also supported by our analyses. This may increase the deception of the parasitic eggs because small spots have increased importance in egg recognition by hosts (see above). As such, the higher importance of small speckling mimicry over that of larger spots suggests a role of functional mimicry, which is related to great reed warblers' pattern recognition mechanisms (Moskát et al., 2008a,b). Stoddard and Stevens (2010) also suggested that cuckoos may develop more general characteristics (which is illustrated by high consistency of egg parameter values for cuckoos) in egg mimicry rather than highly specialized characters. Good potential examples for these jack-of-all-trade eggs are the garden warbler (Sylvia borin)-type cuckoos in Europe, which appears in nests of several host species (Lovászi & Moskát, 2004), and the Horsfield's bronze-cuckoo Chalcites basalis in Australia (Feeney et al., 2014). However, in Japan, cuckoo eggs with highly specialized complex lining pattern characters were also found (Nakamura, Kubota & Suzuki, 1998; Takasu et al., 2009; Moskát et al., 2012).

Spottiswoode and Stevens (2012) revealed in an African finch-cuckoo finch relationship that egg spot characteristics appeared to be evolving independently from another in a 40 years period. Interestingly, our conclusion in the common cuckoo-*Acrocephalus* host system is also similar, in that several egg characteristics showed different types of change in a greater than 110 year period. We suggest further analyses on changes of egg spottiness in a wide range of brood parasite-host relationships. Egg collections in museums offer a good basis for comparison with recent states of coevolutionary processes among hosts and brood parasites.

The present study concentrated on changes of eggshell spottiness in time. Previous studies on Hungarian great reed warblers and cuckoos investigated aspects of eggshell coloration in natural parasitism (e.g. Cherry, Bennett & Moskát, 2007b; Igic et al., 2012). A future study may reveal the long-term changes of eggshell coloration in this system. For example, colour space modeling approaches on eggshell coloration of chaffinches (*Fringilla coelebs*) and bramblings (*F. montifringilla*), being ancient or rarely parasitized hosts of the cuckoo in Fennoscandia, revealed that importance of site fidelity versus nomadity of host species, which could also lead to evolutionary changes in host eggshells, representing different evolutionary trajectories (Vikan et al., 2011). Although the basic mechanism affecting eggshell appearance seems to be coevolutionary interactions in brood parasitism, future studies should also focus on other potential effects that could influence changes in egg appearance in each party (e.g. immigration at a metapopulation scale, rainfall, solar effects, and so on). Furthermore, studies on the mechanisms of how hosts use key features of eggshell spotting (signatures) for egg recognition are also badly needed.

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Name of	Description
variables	
Proportion	Expresses the dominance of the main marking size. (It can be defined as the
energy	measure of the importance of the main marking size in relation to the overall egg
	pattern; a high value of it indicates that the egg pattern is dominated by this spot
	size.)
Filter size	The dominant marking filter size reveals which filter size has the most energy,
	i.e. what is the most important marking size.
Total	The overall amplitude of the spectrum, which is essentially how contrasting the
energy	markings are against the egg background colour.
Coverage	The proportion of the egg/region (upper, middle, and base) covered with
	markings.
Dispersion	Degree of how concentrated the markings are across the different egg regions.
	(The standard deviation of pattern coverage in the three egg regions.)

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responses	explanatories	d.f.	SS	MS	iteration	P
Proportion energy						
	Year	1	0.081	0.082	5000	< 0.001
	Species	1	0.014	0.014	3167	0.031
	year:species	1	< 0.001	< 0.001	51	0.863
	Topology	2	0.005	0.002	509	0.473
	year:topology	2	0.007	0.004	936	0.405
	species:topology	2	0.017	0.008	2359	0.041
	year:species:topology	2	0.004	0.002	51	0.980
	Residuals	372	1.096	0.003		
Filter size						
	Year	1	2.56	2.555	51	1.000
	Species	1	8.17	8.167	4864	0.020
	year:species	1	0.14	0.139	51	1.000
	Topology	2	6.90	3.448	51	1.000
	year:topology	2	1.36	0.681	69	0.84
	species:topology	2	7.77	3.885	2395	0.063
	year:species:topology	2	3.52	1.761	1478	0.063
	Residuals	372	519.55	1.397		
Log filter size						
	Year	1	0.073	0.073	51	0.804
	Species	1	0.199	0.198	5000	0.017
	year:species	1	0.009	0.009	239	0.297
	Topology	2	0.071	0.036	1232	0.337
	year:topology	2	0.036	0.018	852	0.437
	species:topology	2	0.170	0.085	941	0.166
	year:species:topology	2	0.068	0.034	81	0.988
	Residuals	372	12.274	0.033		
Coverage						
	Year	1	0.110	0.109	4157	0.024
	Species	1	1.035	1.035	5000	< 0.001
	year:species	1	0.007	0.007	65	0.615

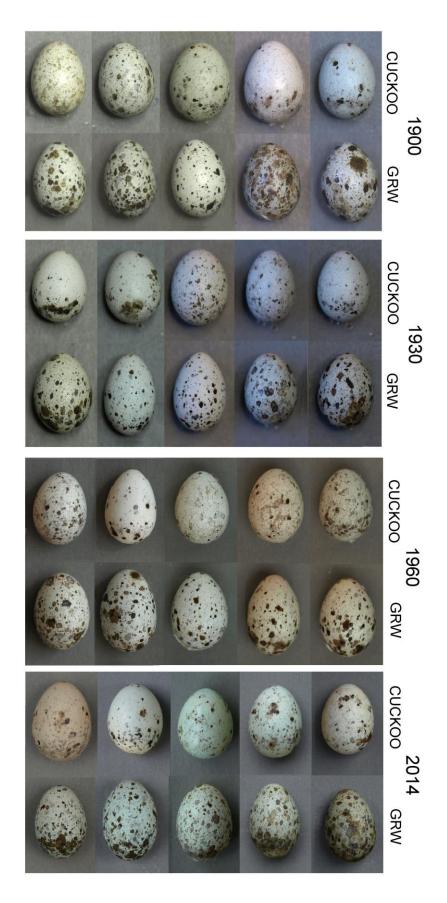
	Topology	2	1.538	0.769	5000	< 0.001
	year:topology	2	0.048	0.024	2921	0.121
	species:topology	2	0.002	0.001	51	1.000
	year:species:topology	2	0.023	0.012	154	0.870
	Residuals	372	6.332	0.017		
Total energy						
	Year	1	396596	396596	5000	< 0.001
	Species	1	4268584	4268584	5000	< 0.001
	year:species	1	131042	131042	51	1.000
	Topology	2	265843	132922	5000	< 0.001
	year:topology	2	83547	41773	2181	0.044
	species:topology	2	207637	103818	5000	< 0.001
	year:species:topology	2	5431	2716	191	0.613
	Residuals	372	6399311	17202		
Dispersion						
	Year	3	0.26	0.08	5000	< 0.001
	Species	1	0.01	0.01	342	0.22
	year:species	3	0.03	0.01	534	0.63
	Residuals	120	1.76	0.01		

Table 3. Summary table for the permutational ANOVA test for egg mimicry (the difference between the two species' values for each variable). See Table 1 for the explanation of variable names. Legends for the independent variables: year (1900s, 1930s, 1960s, and 2014), species (great reed warbler and common cuckoo), topology (the top, middle and bottom regions of eggs). Significant terms (p < 0.05) are shown in bold.

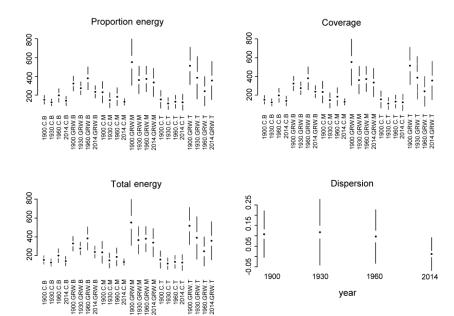
Responses	explanatories	d.f.	SS	MS	iteration 648 _P
Proportion energy					649
	Year	3	0.030	0.013	650 179_0.363
	Topology	2	0.027	0.005	651 592 0.174
	year:topology	6	0.754	0.006	541 ⁶⁵² 0.638
	Residuals				653
Coverage					654
	Year	3	0.144	0.048	21896 5% 198
	Topology	2	0.003	0.001	51 ₆₅ 6000
	year:topology	6	0.056	0.009	$251_{65}^{0.916}$
	Residuals	132	4.153	0.031	658
Total energy					
	Year	3	316611	105537	5000 ⁶⁵⁹ 0.036
	Topology	2	229311	114656	5000 ⁶⁶ 0.008
	year:topology	6	239254	39876	4060660.182
	Residuals	132	3333508	25254	662
Filter size					663
	Year	3	1.690	0.565	136 ₆₆₄ 889
	Topology	22	9.120	4.563	792 0.246 665
	year:topology	6	3.100	0.516	179 1.000
	Residuals	132	345.83	2.619	666
Log filter size					667
	Year	3	0.035	0.012	53 668943
	Topology	2	0.156	0.078	319_{66} 9241
	year:topology	6	0.071	0.012	$201_{670}^{0.935}$
	Residuals	132	8.007	0.061	671

673 Legend to figures 674 675 Fig. 1 Common cuckoo eggs found in Hungary in great reed warbler nests with a randomly-676 selected host egg from the same nest. For each study period five clutches are shown. (Scale 677 was not adjusted. Photo credit Miklós Bán and Csaba Moskát) 678 679 Fig. 2 Changes in eggshell characters over time. On the graphs, the full circles show the 680 mean, while the white space between the circles and the vertical lines represent the standard 681 error for the mean, while the vertical lines denote the standard deviation of the samples. 682 Variables: proportion energy, coverage, total energy, and dispersion, C: common cuckoo 683 eggs, GRW: great reed warbler eggs, B: bottom region, M: middle region, and T: top region 684 of eggs. (See for more details in Methods.) The two species were combined where no 685 difference was revealed. 686 687 Fig. 3 Changes in eggshell total energy for mimicry between common cuckoos and great reed 688 warblers over time. Mimicry was defined as the absolute difference between the two species 689 egg characters. On the graphs, the full circles shows the mean, while the white space between 690 the circles and the vertical lines represent the standard error for the mean, while the vertical 691 lines denote the standard deviation of the samples. (Letters B, M, and T denotes the bottom, 692 middle, and top regions of eggs, respectively) 693 694 Fig. 4 Seasonal decomposition of time series data for total energy of mimicry between 695 common cuckoos and great reed warblers between 1900 and 2014. (Mimicry was defined as 696 absolute difference between the two species egg characters.) The data denote the original 697 structure of time series data, the season indicates the within-year variation patterns, whereas 698 the trend denotes the global (between-year) patterns in the data set. The remainder shows 699 unexplained variances. 700 701 702 703

704 Fig. 1



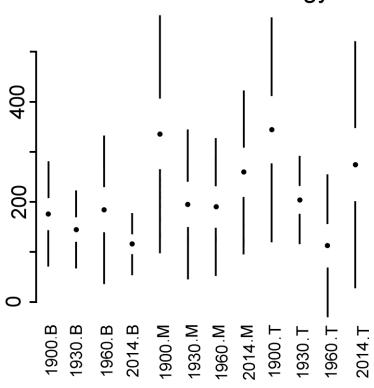
707 Fig. 2



710 Fig. 3 711

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Difference in total energy



714 Fig. 4

Total energy

