

## Amended synthesis of vitamin B<sub>6</sub> causes altered antioxidant responses to supplemental UV-B in *Arabidopsis thaliana*

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We previously showed that UV-B can increase metabolic hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) concentrations in leaves and that it is able to convert H<sub>2</sub>O<sub>2</sub> to hydroxyl radicals (\*OH) (Czégény et al. 2014). Thus, efficient scavenging of H<sub>2</sub>O<sub>2</sub> and \*OH are expected to be important aspects in a successful acclimation to UV-B. Vitamin B<sub>6</sub> has an essential role both in plant development and stress tolerance (Raschke et al. 2011). In addition to their coenzyme function in several biosynthetic pathways (Drewke and Leistner 2001), B<sub>6</sub> derivatives are potent quenchers of ROS (Havaux et al. 2009, Matxain et al. 2009).

In this study, we used *Arabidopsis thaliana* mutants (*rsr4-1*) reduced in B<sub>6</sub> biosynthesis (Wagner et al. 2006) to investigate how vitamin B<sub>6</sub> derivatives contribute to the plants' acclimation to supplemental UV-B in growth chambers. In response to UV-B both mutant and wild type (C24) leaves altered their antioxidant profiles – including increases in B<sub>6</sub> derivatives. Wild type plants avoided oxidative stress via increasing peroxidase activities. Mutants, however, showed elevated catalase and markedly decreased SOD activities, although these were not sufficient to maintain leaf photochemistry. Responses are also discussed in terms of changes in leaf B<sub>6</sub> profiles and ROS reactivities of these compounds.

### References

- Czégény, G., Wu M., Dér A., Eriksson L.A., Strid, Å., Hideg É (2014) FEBS Lett 588, 2255-2261.
- Raschke M., Boycheva S., Crèvecoeur M., Nunes-Nesi A., Witt S., Fernie A.R., Amrhein N., Fitzpatrick T.B. (2011) Plant J 66, 414-432.
- Drewke C, Leistner E (2001) Vitam Horm 61, 121-155.
- Havaux M., Ksas B., Szewczyck A., Rumeau D., Franck F., Caffarri S., Triantaphylidès C. (2009) BMC Plant Biol 9, 130.
- Matxain, J.M., Padro, D., Ristilä, M., Strid, Å., & Eriksson, L.A. (2009) J Phys Chem B 113, 9629-9632.
- Wagner, S., Bernhardt A., Leuendorf J.E., Drewke C., Lytovchenko A., Mujahed N., Gurgui C., Frommer W.B., Leistner E, Fernie A.R., Hellmann H. (2006) Plant Cell 18, 1722-1735.

### Acknowledgements

The work was supported by the Hungarian Scientific Grant Agency (grant number OTKA K112309). Gy. Cz. acknowledges the support of the ÚNKP-17-3-III-PTE-229 New National Excellence Program of the Ministry of Human Capacities. Å.S. was supported by grants from the Knowledge Foundation and the Swedish Research Council FORMAS. This work was also supported by the János Bolyai Research Scholarship of the Hungarian Academy of Sciences.