

1 **The original published PDF available in this website:**

2 <https://www.sciencedirect.com/science/article/pii/S0304377018303024?via%3Dihub>

3

4 Short communication

5 **Is *Nymphaea lotus* var. *thermalis* a Tertiary relict in Europe?**

6 Levente Laczkó<sup>a,1</sup>, Balázs András Lukács<sup>b,1\*</sup>, Attila Mesterházy<sup>c</sup>, Attila Molnár V.<sup>a</sup>, Gábor Sramkó<sup>a,d</sup>

7 <sup>a</sup> Department of Botany, University of Debrecen, H-4032, Debrecen, Egyetem tér 1., Hungary

8 <sup>b</sup> Department of Tisza-River research, MTA Centre for Ecological Research, H-4026 Debrecen,  
9 Bem tér. 18/C, Hungary

10 <sup>c</sup> Hortobágy National Park Directorate, H-4026 Debrecen Sumen u. 2., Hungary

11 <sup>d</sup> MTA-DE “Lendület” Evolutionary Phylogenomics Research Group, H-4032 Debrecen, Egyetem  
12 tér 1, Hungary

13

14 \* Corresponding author.

15 E-mail address: lukacs.balazs@okologia.mta.hu

16 <sup>1</sup> Contributed equally to this work.

## 17 **Abstract**

18 *Nymphaea lotus* L. is a tropical and subtropical species of waterlilies with an African distribution. A  
19 conspicuous satellite occurrence can be found in Europe in a thermal spring in NW Romania. This  
20 population (treated as var. *thermalis* (DC.) Tuzson) received much attention as a potential Tertiary  
21 relict in the flora of Europe, although its relict vs. planted nature has been part of great debate  
22 among scholars during the last 200 years. We revisit this question by using molecular phylogenetic  
23 methods to estimate the timing of divergence of this species, and put this population into a  
24 phylogeographic context by comparing it to samples coming from the whole area of the species. By  
25 using sequences of the nuclear and the plastid genome, we reconstructed the phylogeographic  
26 relationships within *N. lotus* with a haplotype network building approach, that identified a  
27 genetically distant lineage in western Africa, which we relate to *N. zenkeri* Gilg, the sister species of  
28 *N. lotus*. All the populations of *N. lotus* s.str. displayed genetic distance between each other except  
29 for the Egyptian and Romanian populations. On our dated phylogeny of the subgenus *Lotus* the  
30 separation between *N. lotus* and its sister species was found to be younger than the end of the  
31 Tertiary (1.26 Mya). These results clearly refute the Tertiary relict status of the European population  
32 of *N. lotus*.

33

## 34 **Keywords:**

35 cultural relict; *Nymphaea* subg. *Lotos*; *N. zenkeri*; phylogeography; satellite population; Natura  
36 2000, Transylvanian hot-spring lotus beds (31A0); thermal refugium

## 37 1. Introduction

38 The waterlily genus (*Nymphaea* L.) has five subgenera and includes mostly tropical aquatic plants.  
39 A subgenus of the tropical Old World is subg. *Lotos* D.C. with two species in Africa, three in Asia  
40 (Conard, 1905). An African member of the genus is *Nymphaea lotus* L., whereas its sister species  
41 (sometimes also referred to as *N. lotus* in a wide sense) is *N. pubescens* Willd. (Borsch et al., 2007),  
42 which grows in South-East Asia. There is a conspicuous satellite occurrence of *N. lotus* in Europe,  
43 in a thermal lake (Lake Pețea near to Oradea city, NW Romania), *ca* 3000 km away from the  
44 nearest location (the Nile Delta in Egypt). The European occurrence was found by Pál Kitaibel in  
45 1798 during his trip to NE Hungary (the territory belonged to the Hungarian Kingdom in that  
46 period). One century later, several influential scholars started to advocate their opinion about the  
47 autochthonous nature of this population (e.g., Pax, 1905; Staub, 1903), thus, concluded the Tertiary  
48 relict origin of this species in the thermal lake. Literally speaking, this would mean the survival of  
49 this *Nymphaea* population in lake Pețea during the last *ca* 2.5 Myr including four main glaciations.  
50 Additional evidence for the Tertiary relict nature of the plant came from fossil leaves of *Nymphaea*  
51 species found in the region (Pax, 1905) and the co-occurrence of a freshwater snail from a  
52 predominantly (sub)tropical genus, *Melanopsis parreysii* (Phillippi R.A. 1847) (Brusina, 1902),  
53 together with the waterlily. A study based on pollen analysis also supported the relict status of this  
54 species (Diaconeasa and Popa, 1964). However, some scholars expressed their doubts about the  
55 relict status of this species and advocated its origin as result of recent dispersal by water birds (e.g.,  
56 Borbás, 1894; Richter, 1897) or plantation by Turkish during the Ottoman occupation of Hungary in  
57 the 16<sup>th</sup>–17<sup>th</sup> century (Tuzson 1908). Despite of this evident uncertainty, it is generally accepted  
58 nowadays that the population in the thermal lake near Oradea, treated taxonomically as *Nymphaea*  
59 *lotus* L. var. *thermalis* (D.C.) Tuzson, is a Tertiary relict in the Romanian (Pop, 1976), and an  
60 indigenous member of the European Flora (Jalas and Suominen, 1989; Tutin and Webb, 1992). The  
61 conservation importance of the plant is reflected by the designation of an original habitat type of the  
62 Natura 2000 network, Transylvanian hot-spring lotus beds (31A0), to the preservation of the unique  
63 community at this location including the local *N. lotus* population.

64 Molecular phylogenetic and phylogeographic approaches together with the usage of the  
65 molecular clock make it possible to revisit the relict nature of this population. In light of the detailed  
66 phylogenetic information on the genus *Nymphaea* including the dated phylogeny of the genus  
67 (Borsch et al., 2007; Löhne et al., 2008), we attempt to investigate the speciation rates within subg.  
68 *Lotos* and the genetic distances within *Nymphaea lotus* using fast-evolving, neutral DNA regions  
69 from the nuclear and the plastid genome. If *Nymphaea lotus* var. *thermalis* is really a Tertiary relict  
70 taxon and the speciation rate in the subgenus is constant, a great genetic distance must exist  
71 between *Nymphaea lotus* subsp. *lotus* and the European population. Additionally, the divergence of

72 other species within the subgenus must pre-date the separation of the conspecific populations in  
73 Africa and Europe.

74

## 75 **2. Materials and Methods**

### 76 *2.1 Sampling and laboratory protocols*

77 Our field sampling represents the whole distribution area of *Nymphaea lotus* s.l. (Table S1).  
78 Samples were collected and stored in either 96% ethanol or in silica gel. These field samples were  
79 genotyped for four DNA regions: the nuclear ribosomal ITS (nrITS) and three plastid spacers  
80 (*rpl32-trnL*, *psbM-trnD*, *trnT-trnL*). Newly generated DNA-sequences were uploaded to GenBank  
81 (Table S3).

82 For our field collected samples, we extracted genomic DNA using a modified CTAB method as  
83 detailed here. 800 µl of extraction buffer consisting of 2% CTAB, 0.1 M Tris-HCl (pH=7.5), 1.4 M  
84 NaCl and 0.02 M EDTA were added to the ground leaves. After 1 hour of incubation on 65°C the  
85 mixtures were washed twice with chloroform : isoamylalcohol (24 : 1). DNA was precipitated using  
86 100% isopropanol. After centrifuging pellets were washed twice with 70% ethanol, then air-dried  
87 and resuspended in 1 mM Tris-HCl (pH=7.5) buffer. Primer informations and their sequences used  
88 in the amplification and sequencing of the four loci involved in our study are listed in Table S2.  
89 Polymerase Chain Reactions (PCR) were conducted in 25 µl containing 1× Reaction Buffer  
90 (Thermo Scientific), 0.2 mM of each dNTP (Thermo Scientific), 25 mM MgCl<sub>2</sub>, 0.25 mg Bovine  
91 Serum Albumin (Invitrogen), 0.2 µM of each primer, 0.03 U of Taq Polymerase (Thermo Scientific)  
92 and 100 ng DNA template. PCR amplicons were sequenced using a commercial service (Macrogen,  
93 Korea). PCR conditions for nrITS were the following: initial 3 min denaturation at 94°C followed  
94 by 35 cycles of denaturation for 30 sec at 94°C, 30 sec annealing at 51°C, 40 sec extension on 72°C  
95 and a final extension for 7 min at 72°C. For all the plastid markers we used the same touchdown  
96 PCR profile: initial denaturation for 3 min at 94°C followed by 20 cycles of 30 sec denaturation at  
97 94°C, 30 sec annealing at 58°C at the first cycle then lowering the temperature by 0.5°C in each  
98 cycle, then extension for 50 sec at 72°C. The same cycling conditions were repeated for another 20  
99 cycles except that annealing temperature were kept at 48°C in every cycle, then final extension  
100 were conducted for 7 min at 72°C. Sequences were checked for errors and aligned by MUSCLE  
101 v.3.8.31 (Edgar, 2004). Accession numbers of sequences generated for the study can be found in  
102 Table S3.

103

### 104 *2.2 Analysis of evolutionary rates and phylogeographic patterns*

105 To assess evolutionary rates within *Nymphaea* subg. *Lotos*, we first assessed divergence times of  
106 taxa that are relatively closely related and diverged less than 25.4 Mya (Löhne et al. 2008). This  
107 analysis was based on the nrITS + *trnT-trnL* dataset and was run using \*BEAST v.2.5.0 (Bouckaert  
108 et al., 2014). Sequences of species that were not available for us as field-collected samples were  
109 obtained from GenBank (Table S3). As our phylogeographic analyses (see below) unveiled a  
110 significant genetic distance between western African samples and the rest in *N. lotus* s.l., we treated  
111 this population as a separate species (*Nymphaea zenkeri* Gilg) in the divergence time analyses.  
112 \*BEAST was run two times for one billion generations using the uncorrelated lognormal relaxed  
113 clock model with a Yule process of speciation. Bayesian Model Test was run simultaneously with  
114 the data to find the optimal *a priori* molecular evolutionary model for the analysis. Runs were only  
115 accepted if Effective Sample Size (ESS) values were >200. The topology was constrained for the  
116 outgroup topology and secondary dating was accomplished by fixing the mean values of the  
117 chronogram of Löhne et al. (2008). Two calibration points were applied. We constrained the  
118 divergence of *Nymphaea* subg. *Nymphaea* from the rest of the taxa as the root age in our analysis  
119 ( $25.4 \pm 14.3$  Mya). The second calibration point was fixed for the separation of *Nymphaea* subg.  
120 *Lotos* and *Nymphaea* subg. *Hydrocallis* ( $11.3 \pm 7.8$  Mya). Bayesian analysis of macro-evolutionary  
121 mixtures (BAMM) (Rabosky, 2014) was run for 10 million generations sampling every 10000  
122 generations on the reconstructed species tree with global sampling probability set to 0.6 (i.e., we  
123 suppose our sampling covers >60% of the species in the lineage). The first 10% was discarded as  
124 burn-in, then ESS was checked for convergence using the R package BAMMtools (Rabosky et al.,  
125 2014) which we also used for estimating evolutionary rates over time and most probable number of  
126 rate shifts.

127 To investigate the phylogeographic pattern and genetic distances within *Nymphaea lotus* s.l., we  
128 reconstructed the haplotype network of the field sampled populations. In addition to nrITS and  
129 *trnT-trnL*, this dataset was supplemented with two additional plastid DNA-regions (*rpl32-trnL* and  
130 *psbM-trnD*) to investigate phylogeographic relationships within this group in more detail.  
131 Haplotype reconstruction on the concatenated dataset (3523bp) was done in the R package PEGAS  
132 (Paradis, 2010). As an outgroup, we included the W African *Nymphaea zenkeri* population.

133

### 134 3. Results and Discussion

135 Our estimate of divergence times (Fig. 1) for the main lineages of *Nymphaea* and related genera fit  
136 in the 95% HPD interval described in Löhne et al. (2008). The lineage of subg. *Lotos* emerged 5.07  
137 (95% HPD: 0–10.32) Mya, while the separation between *N. pubescens* and *N. lotus* is estimated to  
138 be 2.85 Myr old (95% HPD: 0–6.49), which correspond to the late Pliocene. Divergence between *N.*

139 *lotus* and *N. zenkeri* is assessed to have happened 1.26 Mya (95% HPD: 0–3.62) in Mid-  
140 Pleistocene. According to the results of BAMM, we detected no sign of significant rate shift in the  
141 phylogeny reconstructed based on the dataset, although a continuous decrease is visible (Fig. 1).  
142 Mean speciation rate on the whole chronogram varies between 0.17–0.26 species/Myr, within subg.  
143 *Lotos* mean rate varies between 0.17–0.18 species/Myr. If the European population was a Tertiary  
144 relict isolated at least *ca* 2.5 Mya, given the approximately constant rate of speciation in the lineage,  
145 we would expect it to be separated from the rest of the samples of *N. lotus* s.s. with a significant  
146 genetic difference between them, similar to what we see between *N. lotus* s.l. and *N. pubescens*.  
147 However, this is not the case as our phylogeographic analysis of *N. lotus* s.l. samples did not find a  
148 significant genetic distance between the African and the European plants (Fig. 2). Nonetheless,  
149 samples from the western African population showed remarkable divergence: five hypothetical  
150 haplotypes separated these plants from the rest, whereas we can only see single steps between the  
151 other samples of *N. lotus*. This separation of the western African plant might correspond to the  
152 major clades associated with the main refugial areas in Africa (Hewitt, 2004), and can validate the  
153 taxonomic opinion of those authors who separated the western African populations of *N. lotus* s.l. as  
154 *N. zenkeri* (see Verdcourt, 1989). The East African (Lake Ziwai) and the South African (Nylsvley)  
155 populations are only two steps (i.e., one hypothetical haplotype) away from the North African (Nile  
156 Delta) population. Surprisingly, the European population shares an identical DNA sequence on  
157 3523bp length of rapidly evolving DNA regions with the plant from the Nile Delta. This stands in  
158 stark contrast to the alignment of nrITS sequences between *N. lotus* and *N. pubescens*, taxa that  
159 diverged before the Pleistocene (2.85 Mya), where we can find 27 polymorphic sites and two  
160 singleton indels on 645bp length (our sample ‘lotEgy’ aligned against GenBank accession FJ597743  
161 of *N. pubescens*). Similarly, the alignment of *trnT-trnL* contains two SNP-s and two minisatellite  
162 indel motifs on 1504 bp length (again, our sample ‘lotEgy’ aligned against GenBank accession  
163 AM422043 of *N. pubescens*).

164 Our finding is further corroborated by the molecular phylogenetic results of Smoleń and Falniowski  
165 (2010) who found that the other putative Tertiary relict snail species at the locality, *Melanopsis*  
166 *parreyssii*, is, in fact, closely related to a freshwater snail characteristic of central European thermal  
167 springs, *Fagotia acicularis* (Férussac, 1823). Whereas their isolation coincides with the onset of the  
168 Quaternary, the phylogenetic results clearly separate the subtropical *M. costata* Olivier, 1804 from  
169 *M. parreyssii*, and thus hint at morphological convergence between these two species. Moreover,  
170 paleontological evidence demonstrate that shell morphology of *M. parreyssii* at Lake Pețea is  
171 transforming through geological times and thus can be regarded as an ecological adaptation to warm  
172 water (Sümegei et al. 2012a). Additionally, paleoecological reconstructions uncovered the temperate  
173 refuge (or oasis) nature of the thermal lake during the last glaciation, not a subtropical one, and the

174 strata with fossils of *M. parreysii* and *F. acicularis* goes back only to the late Quaternary (Sümegei et  
175 al., 2012b).

176 In light of the above facts, (i) the lack of genetic divergence between *Nymphaea lotus* var. *thermalis*  
177 and Egyptian *N. lotus* s.s., (ii) the evident non-Tertiary origin of its habitat, and (iii) the non-Tertiary  
178 origin of the co-occurring, presumed relict snail, we conclude the recent occurrence of this species  
179 in Lake Pețea, and rule out of its Tertiary relict status. Although our molecular data can suggest the  
180 recent occurrence of *N. lotus* in Europe on an evolutionary time scale, the exact timing of  
181 introduction is hardly assessable based on our data. According to medieval documents quoted by  
182 Negrean (2011), the lake was established as an artificial pond in the 14<sup>th</sup> century, which makes us  
183 suppose the species has a very recent history in Europe. A possible explanation can be, as already  
184 mentioned in the literature (Tuzson 1908), the plantation by Turkish during the Ottoman occupation  
185 of Hungary, when the Empire extended from North-Africa to central Europe. Ottomans had been  
186 known for their advanced horticulture and their devotion to thermal bathes, which makes it likely  
187 that *Nymphaea lotus* was planted directly to the thermal lake for ornamental purpose. If we accept  
188 this theory, it is still questionable why it was not planted anywhere else in the temperate part of the  
189 Ottoman Empire, for example in the hot springs of the Balkan Peninsula and Asia Minor? Another  
190 way of introduction could be the epizoochorous and endozoochorous dispersion by migratory birds.  
191 However, there is only a few studies that demonstrated the consumption of *Nymphaea* seeds by  
192 waterfowls, and there is no exact evidence for seed dispersal of *Nymphaea lotus* by birds at all  
193 (Green et al., 2002). We may never be able to find an unequivocal explanation for the presence of  
194 *N. lotus* in the thermal spring near Oradea, our data clearly shows its recent introduction to this  
195 habitat.

196

### 197 3.1. Conservational remark

198 Based on four rapidly mutating DNA regions we found no evidence that the only European  
199 population of *Nymphaea lotus* var. *thermalis* in Romania is a Tertiary relict. Two more plausible  
200 explanations for its origin are plantation of the population in historical times or dispersal mediated  
201 by migratory waterbirds. This may imply that we have to regard *N. lotus* var. *thermalis* as a cultural  
202 relict in the flora of Europe, and as such, we believe it still has a conservation value similar to as if  
203 it was a Tertiary relict. Consequently, we fully agree with the protection of this habitat at the EU  
204 level by maintaining the original habitat type 'Transylvanian hot-spring lotus beds' as part of the  
205 Natura 2000 network.

206

#### 207 4. Acknowledgements

208 We appreciate the plant tissue sampling in South Africa for Jacques Gerber and the laboratory help  
209 for Eszter Csoma. This work was supported by Grant TÁMOP-4.2.2/B-10/-1-2010-0024 project,  
210 implemented through the New Hungary Development Plan and co-financed by the European Social  
211 Fund and the European Regional Development Fund. BAL was supported by National Research,  
212 Development and Innovation Office – NKFIH, OTKA PD120775, grant, the Bolyai János Research  
213 Scholarship of the Hungarian Academy of Sciences and by the UNKP-18-4 New National  
214 Excellence Program of the Ministry of Human Capacities.

215

#### 216 5. Declaration of interest

217 Declarations of interest: none

218

#### 219 6. References

- 220 Borbás, V., 1894. A hévízi tündérrózsa keletkezésének analogonja. Természettudományi Közlöny  
221 26, 146–152. (In Hungarian)
- 222 Borsch, T., Hilu, K.W., Wiersema, J.H., Lohne, C., Barthlott, W., Wilde, V., 2007. Phylogeny of  
223 *Nymphaea* (Nymphaeaceae): Evidence from substitutions and microstructural changes in the  
224 chloroplast trnT-trnF region. International Journal of Plant Sciences 168, 639–671.  
225 <https://doi.org/10.1086/513476>
- 226 Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M.A., Rambaut,  
227 A., Drummond, A.J., 2014. BEAST 2: a software platform for Bayesian evolutionary analysis.  
228 PLoS Comput. Biol. 10, 1–6. <https://doi.org/10.1371/journal.pcbi.1003537>
- 229 Brusina, S., 1902. Eine subtropische Oasis in Ungarn. Mitteilungen des Naturwissenschaftlichen  
230 Vereines für Steiermark 28, 101–121. (In German)
- 231 Conard, H.S., 1905. The waterlilies: a monograph of the genus *Nymphaea*. Carnegie Institution of  
232 Washington, [Washington].
- 233 Diaconeasa, B., Popa, D. 1964. Problema relictră a Lotusului [*Nymphaea lotus* L. var. *thermalis*  
234 (DC.) Tuzs.] și a lacului termal de la Băile 1 Mai, in lumina analizelor microstratigrafice.  
235 Contribuții Botanice (1964): 135–140. (In Romanian)



236 Edgar, R.C., 2004. MUSCLE: Multiple sequence alignment with high accuracy and high  
237 throughput. *Nucleic Acids Res.* 32, 1792–1797. <https://doi.org/10.1093/nar/gkh340>

238 Green, A.J., Figuerola, J., Sánchez, M.I., 2002. Implications of waterbird ecology for the dispersal  
239 of aquatic organisms. *Acta Oecol.* 23, 177–189. [https://doi.org/https://doi.org/10.1016/S1146-](https://doi.org/https://doi.org/10.1016/S1146-609X(02)01149-9)  
240 609X(02)01149-9

241 Hewitt, G.M., 2004. The structure of biodiversity - Insights from molecular phylogeography.  
242 *Frontiers in Zoology* 1, 1–16. <https://doi.org/10.1186/1742-9994-1-4>

243 Jalas, J., Suominen, J., (Eds.) 1989. *Atlas Flora Europaeae*. Vol. 8. Nymphaeaceae to  
244 Ranunculaceae. Biological Society of Finland, Vanamo

245 Löhne, C., Yoo, M.J., Borsch, T., Wiersema, J., Wilde, V., Bell, C.D., Barthlott, W., Soltis, D.E.,  
246 Soltis, P.S., 2008. Biogeography of Nymphaeales: Extant patterns and historical events. *Taxon*  
247 57, 1123–1146. <https://doi.org/10.2307/27756769>

248 Negrean, G., 2011. Addenda to „Flora Romaniaae” volumes 1–12. Newly published plants,  
249 nomenclature, taxonomy, chorology and commentaries (Part 1). *Kanitzia* 18, 89–194.

250 Paradis, E., 2010. PEGAS: an R package for population genetics with an integrated–modular  
251 approach, *Bioinformatics* (Oxford, England). <https://doi.org/10.1093/bioinformatics/btp696>

252 Pax, F., 1905. Die fossile Flora von Gánócz bei Poprád. Beiblatt zu den *Növénytani Közlemények*  
253 4, 19–59. (In German)

254 Pop, E., 1976. Specii relictive în flora României. In: Beldie, A., Morariu, I. (Eds.): *Flora Republicae*  
255 *Socialisticae România*, Vol. 13. Editio Academiae Republicae Socialisticae Romania, pp. 106–  
256 111. (In Romanian)

257 Rabosky, D. L., Grudler, M. , Anderson, C. , Title, P. , Shi, J. J., Brown, J. W., Huang, H. , Larson,  
258 J. G. and Kembel, S., 2014. BAMMtools: an R package for the analysis of evolutionary  
259 dynamics on phylogenetic trees. *Methods in Ecology and Evolution* 5, 701–707.  
260 <https://doi.org/10.1111/2041-210X.12199>

261 Rabosky, D.L., 2014. Automatic detection of key innovations, rate shifts, and diversity-dependence  
262 on phylogenetic trees. *PLoS One* 9, e89543.

263 Richter, A., 1897. A nílusi tündérrózsa, avagy ál-lótusz a Magyar flórában. *Természetráji Füzetek*  
264 20, 204–221. (In Hungarian)

265 Staub, M., 1903. Új bizonyíték a *Nymphaea Lotus* L. magyar honossága mellett. *Növénytani*  
266 *Közlemények* 2, 1–8. (In Hungarian)

267 Smoleń, M., Falniowski, A., 2010. Molecular phylogeny and estimated time of divergence in the  
 268 central European Melanopsidae: *Melanopsis*, *Fagotia* and *Holandriana* (Mollusca:  
 269 Gastropoda: Cerithioidea). *Folia Malacol.* 17, 1–9. <https://doi.org/10.2478/v10125-009-0001-4>  
 270 Sümegi, P., Molnár, D., Sávai, S., Gulyás, S., 2012a. Malacofauna evolution of the Lake Peťa  
 271 (Püspökfürdő), Oradea region, Romania. *Nymphaea, Folia naturae Bihariae* 39, 5–29.  
 272 Sümegi, P., Molnár, D., Sávai, S., Töviskes, R.J., 2012b. Preliminary radiocarbon dated  
 273 paleontological and geological data for the Quaternary malacofauna at Püspökfürdo (Băile 1  
 274 Mai, Oradea region, Romania). *Malakológiai Tájékoztató* 30, 31–37.  
 275 Tutin, T.G., Webb, D.A., 1992. *Nymphaea* L. In: Tutin, T.G., Burges, N.A., Chater, A.O.,  
 276 Edmondson, J.R., Heywood, V.H., Moore, D.M., Valentine, D.H., Walters, S.M., Webb, D.A.  
 277 (Eds.), *Flora Europaea*. Volume 1: Psilotaceae to Platanaceae. 2<sup>nd</sup> Edition. Cambridge  
 278 University Press, pp. 246–247.  
 279 Tuzson, J., 1908. A *Nymphaea lotus* csoport morfológiája és rendszertani tagolódása. *Mathematikai*  
 280 *és Természettudományi Értesítő* 26, 101–137. (In Hungarian)  
 281 Verdcourt, B., 1989. A note on *Nymphaea zenkeri* Gilg (Nymphaeaceae). *Kew Bulletin* 44, 484.  
 282

## Figure captions

**Figure 1.** Dated phylogeny of a set of species in the genus *Nymphaea* with special focus on subgenus *Lotos* reconstructed using the dataset nrITS and *trnT-trnL* together with the plot of speciation rate through time within the lineage. Assessed diversification dates as Mya are placed next by the corresponding node together with 95% HPD bars as grey shadings. Calibration points used for secondary dating are marked with an asterisk. Mean rate of speciation is given as a black curve on the graph, the grey area refers to the 95% HPD intervals. Geographic time scale is placed below the tree at the corresponding place.

**Figure 2.** Haplotype network representing the phylogeographic structure of *Nymphaea lotus* s.l. as reconstructed using the concatenated nrITS and plastid (*trnT-trnL* + *psbM-trnD* + *rpl32-trnL*) sequences. Open circles indicate different haplotypes, whereas black dots refer to hypothetical (unsampled) haplotypes. Size of white circles are proportional to the frequency of haplotypes. Grey dashed line indicates an alternative route in the network. The inserted map shows approximate sampling location for the African samples. *Sample ID abbreviations:* lotNyl: South Africa, Nylsvley Nature Reserve; lotZiw: Ethiopia, Lake Ziwai; lotEgy: Egypt, Nile delta; thePet: Romania, Lake Pețea; zenBuss: Ivory Coast, Grand Bassam