

# Road-killed mammals provide insight into tick-borne bacterial pathogen communities within urban habitats

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## Funding information

Hungarian Eötvös Scholarship; EurNegVec Cost Action, Grant/Award Number: TD1303; János Bolyai Research Scholarship of the Hungarian Academy of Sciences; Hungarian Ministry of Human Resources, Grant/Award Number: 12190/2017/FEKUTSTRAT, 11475-4/2016/FEKUT; In the light of evolution: theories and solutions, Grant/Award Number: GINOP-2.3.2-15-2016-00057

## Abstract

Small- and medium-sized mammals play an important role in the life cycle of tick-borne pathogens in urban habitats. Our aim was to apply the general protocol, DAMA (documentation–assessment–monitoring–action), which is an integrated proposal to build a proactive capacity to understand, anticipate, and respond to the outcomes of accelerating environmental change. Here we tested whether road-killed carcasses in urban areas are useful sources of tissue and parasite samples to investigate these species' contribution to the epidemiology of vector-borne diseases. We collected 29 road-killed and 6 carcasses with different causes of mortality (23 northern white-breasted hedgehogs and 12 from seven other mammal species) mainly from Budapest, Hungary. We used quantitative and conventional PCRs to determine pathogens in 90 collected tissues (52 from hedgehogs; 38 from other species) and 417 ticks that were only found on hedgehogs. Tissue samples revealed a wide range of bacteria including human zoonotic pathogens identified as *Anaplasma phagocytophilum* ecotype I, *Borrelia afzelii*, *B. spielmanii*, *Borrelia miyamotoi*, *Rickettsia helvetica*, and *Bartonella* species. Among the 23 collected hedgehog carcasses, 17 (74%) were infected with *A. phagocytophilum*, 6 (26%) with *Borrelia burgdorferi* s.l., 12 (52%) with *R. helvetica*, and 15 (65%) with *Rickettsia* sp. Furthermore, we report the first detection of *Rickettsia* sp. infection in European moles and lesser weasel and *R. helvetica* in stone marten. Through sequencing *B. afzelii*, *R. helvetica*, *R. monacensis* and *A. phagocytophilum* ecotype I were identified in the ticks removed from the carcasses. We showed that road-killed urban mammal species are exposed to multiple tick-borne pathogens but further studies have to clarify whether they, in fact, also have a role in their maintenance and spread. Our study also demonstrates that road-kill can be used in the risk assessment of potential human infection and in the implementation of the DAMA protocol.

## KEYWORDS

DAMA protocol, eco-epidemiology, roadkill, tick-borne pathogens, urban mammals

## 1 | BACKGROUND

Many of the approximately 700 known hard tick species (Acari, Ixodidae) act as vectors of pathogens with veterinary and medical

relevance. Wild and urban animals may play an important role in the epidemiology of tick-borne diseases (TBDs) and in the spread (to new regions) and maintenance of tick populations. In Hungary, several wildlife species have been found to harbour genetic

material of various tick-borne pathogens potentially affecting both domestic animals and humans (Földvári et al., 2014; Hornok et al., 2014, 2015; Szekeres et al., 2015a,b; Tolnai, Sréter-Lancz, & Sréter, 2015). Within Budapest, the capital of Hungary, 48 different mammal species from bats (Chiroptera) to wild boar (*Sus scrofa*) have been recorded since 1990 (Tóth-Ronkay et al., 2015). Hedgehogs and squirrels can reach higher densities in urbanized areas than in wild habitats (Reeve, 1994; Tóth-Ronkay et al., 2015). They are living in surrounding forests, bigger city parks, cemeteries and also close to human dwellings, e.g., hedgehogs and squirrels in the backyards or martens in the lofts. The trend to increase the urban green areas for recreation, reduce air and noise pollution, and mitigate the frequent fluctuations of humidity and heat is improving the well-being of the citizens. On the other hand, these green areas can serve as important habitats for several wildlife species (e.g., in winter time or when food is scarce). These animals often disperse when looking for new territories and can spread ticks from natural habitats to the heart of the cities and can help to maintain a stable tick and tick-borne pathogen population. As an example hedgehogs can maintain the whole cycle of *Ixodes ricinus* and several tick-borne pathogens as well (Földvári et al., 2011, 2014; Krawczyk et al., 2015). For urban animals, additional hazards of the city life such as vehicles on the roads are frequent cause of death. The resulting carcasses may provide a convenient, noninvasive, and useful source of samples for research projects targeting tick-borne pathogens in urban habitats. Exploring the spectrum of pathogens in roadkill could help to assess possible risk factors of human infection.

Our aim was to apply the general protocol, DAMA (documentation–assessment–monitoring–action), which is an integrated proposal to build a proactive capacity to understand, anticipate, and respond to the outcomes of accelerating environmental change (Brooks et al., 2014). In this article, we focused on documentation and assessment of communities of tick-borne bacterial pathogens in ectoparasites and tissue samples collected from mammalian roadkills in urban areas.

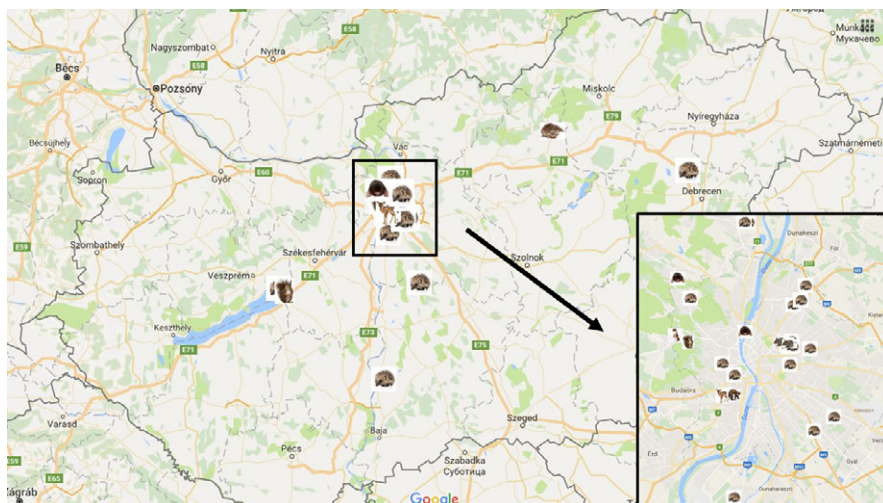


**FIGURE 2** Partly engorged *Ixodes ricinus* female attached to the abdomen of a road-killed hedgehog [Colour figure can be viewed at wileyonlinelibrary.com]

## 2 | METHODS

### 2.1 | Sample collection

Between April and August of 2015, we collected 29 road-killed mammals, mainly hedgehogs with the help of volunteers mainly from Budapest and some other locations around Hungary. In addition, we also collected six animals that died for other reasons (e.g., caught by cats, for details see Supporting Information Table S1). Volunteers were asked personally or through social media to inform us about road-hit carcasses in urban areas and we collected the samples. The study was carried out with the official permission from the Middle-Danube-Valley Inspectorate for Environmental Protection, Nature Conservation and Water Management, Hungary (KTF:4018-4/2015). We bagged carcasses on site and dissected them under laboratory conditions. We collected samples from various identifiable tissues (minimum: skin, maximum: five different tissues) paying attention to use sterilized or unused devices for every tissue sample. Blood was taken from the heart of fresh carcasses, and coagulated blood was



**FIGURE 1** Locations of the studied road-killed urban mammals in Hungary. An online zoomable version of the map with details about each sample is available at: <https://goo.gl/9eeZm7>. Symbols represent one mammal specimen each from the following species: hedgehog: *Erinaceus roumanicus*; mole: *Talpa europaea*; squirrel: *Sciurus vulgaris*; mouse: *Mus musculus*; marten: *Martes foina*; weasel: *Mustela nivalis*; shrew: *Crocodyra suaveolens*; roe deer: *Capreolus capreolus* [Colour figure can be viewed at wileyonlinelibrary.com]

**TABLE 1** Number, stage, and species of ticks removed from the collected hedgehog carcasses

Host code	<i>Ixodes ricinus</i>				<i>Ixodes hexagonus</i>				Sum
	Larva	Nymph	Female	Male	Larva	Nymph	Female	Male	
H1	-	1	-	-	-	-	-	-	1
H4	35	156	11	5	7	5	-	-	219
H7	-	87	10	4	-	-	-	-	101
H9	-	-	1	1	-	-	-	-	2
H11	2	4	2	3	-	-	-	-	11
H12	-	10	1	1	-	-	-	-	12
H15	-	5	4	-	-	-	-	-	9
H16	-	15	1	1	-	41	2	-	60
H21	-	-	1	1	-	-	-	-	2
Sum	37	278	31	16	7	46	2	-	417

**TABLE 2** Prevalence of tick-borne pathogens in tissue samples of small- and medium-sized mammals detected with real-time PCR. *Can. N. mikurensis* was not detected in any of the tissue samples

Mammal/Pathogen species	<i>Borrelia burgdorferi</i> s.l.	<i>Borrelia miyamotoi</i>	<i>Anaplasma phagocytophilum</i>	<i>Rickettsia helvetica</i>	<i>Rickettsia</i> sp.	<i>Bartonella</i> sp.
	No. positive tissue samples/No. tested tissue samples/prevalence (%)					
White-breasted hedgehog ( <i>Erinaceus roumanicus</i> )	6/52/12%	0/52/0%	27/52/52%	15/52/29%	19/52/37%	4/52/8%
European mole ( <i>Talpa europaea</i> )	0/15/0%	0/15/0%	0/15/0%	0/15/0%	1/15/7%	9/15/60%
House mouse ( <i>Mus musculus</i> )	0/5/0%	0/5/0%	0/5/0%	1/5/20%	1/5/20%	1/5/20%
European red squirrel ( <i>Sciurus vulgaris</i> )	1/6/17%	1/6/17%	0/6/0%	0/6/0%	0/6/0%	0/6/0%
Roe deer ( <i>Capreolus capreolus</i> )	0/3/0%	0/3/0%	1/3/33%	0/3/0%	0/3/0%	0/3/0%
Lesser shrew ( <i>Crocidura suaveolens</i> )	0/3/0%	0/3/0%	1/3/33%	0/3/0%	0/3/0%	0/3/0%
Stone marten ( <i>Martes foina</i> )	0/2/0%	0/2/0%	0/2/0%	1/2/50%	0/2/0%	0/2/0%
Lesser weasel ( <i>Mustela nivalis</i> )	0/4/0%	0/4/0%	0/4/0%	0/4/0%	1/4/25%	2/4/50%
Sum	7/90/8%	1/90/1%	29/90/32%	17/90/19%	22/90/24%	16/90/18%

Bold values indicate prevalence data.

taken from the body cavity in case of older carcasses if organs were not smashed. The species, date of collection, location, and the degree of body degradation (see categories in Table 3) were recorded. Before dissection, we thoroughly inspected the bag and the body surface at room temperature and collected all the ectoparasites and stored them in 70% ethanol at 4°C until further molecular analysis. Ticks were identified using standard identification keys (Hillyard, 1996; Nosek & Sixl, 1972). Carcasses and the collected tissue samples were stored at -20°C.

## 2.2 | DNA extraction

DNA was extracted from the ticks individually (adults) or in pools (larvae by up to 10; nymphs by up to 5) with alkaline hydrolysis (Guy & Stanek, 1991). All tick samples were tested. In pooled samples, pools comprised of ticks of the same species and from the same host individual. We used ISOLATE II Genomic DNA Kit (Bioline Reagents Ltd, London, UK) to isolate the nucleic acid from the tissue samples. A pea-sized sample was taken from all available organs and

lysed without any mechanical disruption according to the manufacturer's instruction. The final elution volume was 100 µl. We stored the DNA samples at -20°C until further molecular analysis.

## 2.3 | Polymerase chain reaction

To determine whether the tissue or tick samples contained any pathogens, we used species or genus specific quantitative PCR (qPCR) assays. When a sample was positive with genus-specific reactions, we used conventional PCR and sequencing to specify the pathogen species. In the qPCR reactions, we used primers targeting the specific gene regions for *Borrelia burgdorferi* s.l. (*ospA*: outer surface protein A gene, *flab*: flagellin gene) (Heylen, Tijssse, Fonville, Matthysen, & Sprong, 2013), *Borrelia miyamotoi* (*flaB*) (Hovius et al., 2013), *Anaplasma phagocytophilum* (*msp2*: major surface protein 2 gene) (Courtney, Kostelnik, Zeidner, & Massung, 2004), *Candidatus Neohrlichia mikurensis* (*GroEL*: heat shock protein gene) (Jahfari et al., 2012), *Rickettsia helvetica* (*gltA*: citrate synthase gene) (de Bruin et al., 2015), and *Rickettsia* sp. (*gltA*) (Stenos, Graves, & Unsworth, 2005).



**FIGURE 3** Seemingly intact road-killed red squirrel [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]



**FIGURE 4** Seemingly intact road-killed northern white-breasted hedgehog [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]

Conventional PCRs were used for *A. phagocytophilum* (GroEL) (Jahfari et al., 2014), *Borrelia burgdorferi* s.l. (IGS: intergenic spacer) (Hansford, Fonville, Jahfari, Sprong, & Medlock, 2015), *Rickettsia* (ompB) (Roux & Raoult, 2000), and *Bartonella* species (gltA) (de Bruin et al., 2015). We used distilled water for negative and sequenced known positive samples for positive control.

## 2.4 | Data analysis

To calculate the pathogen prevalence in ticks, we used minimum infection rate, by calculating the prevalence considering that at least one individual is positive per positive pool.

## 3 | RESULTS

### 3.1 | Collected samples

Twenty-three road-killed northern white-breasted hedgehogs (*Erinaceus roumanicus*) and 12 other collected mammals from seven different species (e.g., European red squirrel and European mole) were



**FIGURE 5** Dissection of a road-killed northern white-breasted hedgehog with organs in good condition (degree of degradation 1) [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]



**FIGURE 6** Flat and dry carcass of a road-killed northern white-breasted hedgehog with unrecognizable organs (degree of degradation 5) [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]

included into the study. We collected carcasses of animals struck and killed by motor vehicles on highways ( $n = 29$ ) or accidentally killed animals, e.g., killed by cat ( $n = 6$ ) from urbanized habitats, mainly from Budapest, Hungary (Figure 1; Supporting Information Table S1).

All the removed ticks were from nine hedgehogs (Figure 2); the other carcasses did not harbour any ticks. The maximum number of ticks/host was 219 (Table 1). We collected 417 ticks, 55 *Ixodes hexagonus* (7 larvae, 46 nymphs, 2 females), and 362 *I. ricinus* (37 larvae, 278 nymphs, 16 males, 31 females). Two hedgehogs were infested with

**TABLE 3** Detection of tick-borne bacterial pathogens with real-time PCR in tissue samples according to the degree of degradation of the carcasses

Sample type	Degree of degradation (1–5) <sup>a</sup>	Number of host individuals	Number of collected samples	<i>Anaplasma phagocytophilum</i>	<i>Borrelia burgdorferi</i> s.l.	<i>Borrelia miyamotoi</i>	<i>Rickettsia helvetica</i>	<i>Rickettsia</i> sp.	<i>Bartonella</i> sp.	Number of positive samples <sup>b</sup>
Hedgehogs	1	4	16	4	2	-	3	4	2	15
	2	2	4	1	1	-	2	2	1	7
	3	6	10	5	3	-	5	5	1	19
	4	5	8	3	-	-	2	2	-	7
	5	6	8	3	-	-	1	2	-	6
Other species <sup>c</sup>	1	10	33	1	1	1	1	3	4	11
	3	1	3	1	-	-	-	-	-	1
	4	1	3	-	-	-	1	-	-	1

<sup>a</sup>Degree of degradation: 1—intact perfect condition for internal organ tissue collection at most skull crashed; 2—crushed body with mostly intact internal organs; 3—partly smashed internal organs; 4—smashed internal organs, only muscle and skin tissue, semi-dry carcass; 5—flat and dry carcass with unrecognizable organs (except skin). <sup>b</sup>The number of positive samples for the pathogens is higher than the collected sample number at a given degree of degradation, because one sample can be positive for more than one pathogen. <sup>c</sup>See details in Supporting Information Table S1.

**TABLE 4** Prevalence of tick-borne pathogens in ticks removed from road-killed hedgehogs detected with real-time PCR. *Borrelia miyamotoi* and *Can. N. mikurensis* were not detected in any on-host ticks

Tick species and stage		<i>Borrelia burgdorferi</i> s.l.	<i>Anaplasma phagocytophilum</i>	<i>Rickettsia helvetica</i>	<i>Rickettsia</i> sp.
		No. positive/No. tested/prevalence (%)			
<i>Ixodes ricinus</i>	Female	10/31/32%	23/31/74%	19/31/61%	26/31/84%
	Male	6/16/38%	8/16/50%	7/16/44%	10/16/63%
	Nymph <sup>a</sup>	34/59/17%	54/59/45%	34/59/17%	44/59/26%
	Larva <sup>a</sup>	2/5/6.4%	5/5/7%	3/5/10%	3/5/10%
<i>Ixodes hexagonus</i>	Female	0/2/0%	2/2/100%	0/2/0%	0/2/0%
	Nymph <sup>a</sup>	1/10/2%	9/10/54%	3/10/7%	6/10/19%
	Larva <sup>a</sup>	0/1/0%	1/1/96%	0/1/0%	0/1/0%
Sum*		51/124/16%	101/124/24%	66/124/21%	89/124/33%

Bold values indicate prevalence data.

<sup>a</sup>Minimum infection rate (up to 5 nymphs/pool; up to 10 larvae/pool).

both tick species and seven hedgehogs were infested with only *I. ricinus* (Table 1). From the 417 removed ticks, 124 samples were created and assayed (111 *I. ricinus* and 13 *I. hexagonus* individual or pooled samples).

From the carcasses, we collected 90 tissue samples for molecular analysis (52 from hedgehogs and 38 from the other species) (Table 2). The degree of degradation of the carcasses was different; some specimens were in “perfect” condition (Figures 3–5) with no sign of degradation (degree of degradation 1) and some were flat, dry (Figure 6) and heavily damaged by vehicles (degree of degradation 5) (Table 3).

### 3.2 | Tick-borne pathogens in tissue samples

All examined pathogens were detected in the collected mammal tissue samples except *Can. N. mikurensis*. Among the 23 collected hedgehog carcasses, 17 (74%) were infected with *A. phagocytophilum*, 6 (26%) with *B. burgdorferi* s.l., 12 (52%) with *R. helvetica*,

and 15 (65%) with *Rickettsia* sp. *Borrelia burgdorferi* s.l. was detected in the muscle and skin of hedgehogs (12%) and skin sample from a squirrel (Table 2). *Borrelia miyamotoi* was only detected in a squirrel spleen sample. *Anaplasma phagocytophilum* was found in muscle, skin, liver, spleen, and coagulated blood from hedgehogs (52%); in the skin of a juvenile roe deer; and a lesser shrew muscle sample. *Rickettsia helvetica* DNA was amplified in skin, spleen, and muscle from hedgehogs (29%); skin of a house mouse; and muscle of a stone marten. We found *Rickettsia* sp. positive in skin, spleen, liver, muscle, and coagulated blood of hedgehogs (37%); skin of a house mouse and a mole; and a muscle sample of a lesser weasel. *Bartonella* species were detected in all tissue sample types of moles, skin samples of hedgehogs, muscle of a house mouse, and coagulated blood and muscle of a lesser weasel.

Pathogen prevalence data of different mammal species are shown in Table 2, and all data for positive samples from the small mammals are shown in the Supporting Information Table S1. Pathogen detection was possible from even the most degraded tissues (Table 3).

**TABLE 5** Specification of tick-borne pathogens in tissues samples from small- and medium-sized mammals and ticks removed from hedgehogs

Sample type	Pathogen	Host/tick species	Tissue type/tick stage/sex	Number of samples	Accession numbers	
Tissue	<i>Borrelia afzelii</i>	Northern white-breasted hedgehog ( <i>Erinaceus roumanicus</i> )	Skin	1	MF163403	
			Muscle	2	MF163401, MF163402	
	<i>Anaplasma phagocytophilum</i> ecotype I	Northern white-breasted hedgehog ( <i>E. roumanicus</i> )	Liver	1	MF372764	
			Skin	2	MF372765, MF372766	
			Spleen	3	MF372767-MF372769	
	<i>Bartonella</i> sp.	Northern white-breasted hedgehog ( <i>E. roumanicus</i> )	Muscle	1	MF372778	
			Skin	2	MF372780, MF372781	
			House mouse ( <i>Mus musculus</i> )	Muscle	1	MF372781
			European mole ( <i>Talpa europaea</i> )	Blood	2	MF372782, MF372786
				Liver	1	MF372787
			Muscle	2	MF372785, MF372789	
			Skin	2	MF372784, MF372788	
			Spleen	1	MF372783	
			Lesser weasel ( <i>Mustela nivalis</i> )	Blood	1	MF372790
	Spleen	1	MF372791			
Tick	<i>Borrelia afzelii</i>	<i>Ixodes ricinus</i>	Female	1	MF163404	
			Male	1	MF163405	
			Nymph <sup>a</sup>	7	MF163406-MF163412	
	<i>Anaplasma phagocytophilum</i> ecotype I	<i>Ixodes ricinus</i>	Female	2	MF372771, MF372772	
			Nymph <sup>a</sup>	5	MF372773-MF372777	
	<i>Rickettsia monacensis</i>	<i>Ixodes hexagonus</i>	Female	1	MF372770	
			<i>Ixodes ricinus</i>	Nymph <sup>a</sup>	4	MF170619-MF17062
		<i>Ixodes ricinus</i>	Larva <sup>b</sup>	1	MF170623	

<sup>a</sup>Pooled sample from five nymphs. <sup>b</sup>Pooled sample from 10 larvae.

### 3.3 | Tick-borne pathogens in ticks removed from road-killed hedgehogs

*Borrelia miyamotoi* and *Can. N. mikurensis* DNA were not detected in tick samples (Table 4). *Borrelia burgdorferi* s.l. was detected in 16% (minimum infection rate) of ticks; all stages of *I. ricinus* (female: 32%, male: 38%, nymph minimum infection rate: 17%, larva minimum infection rate: 6%) and *I. hexagonus* nymphs (minimum infection rate: 2%). *Anaplasma phagocytophilum* was represented in all stages of both tick species with relatively high percentage (Table 4). *Rickettsia helvetica* was found in *I. ricinus* females, males, nymphs, and larvae and in *I. hexagonus* nymphs with an overall *R. helvetica* prevalence of 20.5% (minimum infection rate) in ticks. *Rickettsia* sp. was found in all stages of *I. ricinus* and *I. hexagonus* nymphs. Pathogen prevalence in ticks is presented in Table 4.

### 3.4 | Pathogen identification

The qPCR-positive samples were also amplified with specific conventional PCR assays and were sequenced if needed. *Borrelia afzelii* was

found in one *I. ricinus* female, one male, and seven nymphal pools. *Anaplasma phagocytophilum* ecotype I was found in two females and five *I. ricinus* nymphal pools and one *I. hexagonus* female. *Rickettsia monacensis* occurred in four *I. ricinus* nymphal pools and a larval pool. *Borrelia afzelii* and *B. spielmanii* were found only in hedgehog skin and muscle. *Anaplasma phagocytophilum* was detected in liver, skin, and spleen samples from hedgehogs. *Bartonella* species DNA was amplified in *E. roumanicus* (muscle, skin), house mouse (muscle), mole (coagulated blood, liver, muscle, skin, spleen), and lesser weasel (coagulated blood, spleen) tissues. Positive samples were sequenced and a BLAST search was performed using GenBank. The identified pathogens are shown in Table 5. The highest similarity in the case of *Bartonella* sequences was found with a *Bartonella taylorii* (98%) sequence from a plateau pika (*Ochotona curzoniae*; Tibetan plateau, China; accession numbers: KT445922; KT445921; KT445919) (Rao et al., 2015) and 97% with a *Bartonella* sp. sequence from Yunnan red-backed vole (*Eothenomys miletus*; Yunnan, China; accession number: AF391281) (Ying, Kosoy, Maupin, Tsuchiya, & Gage, 2002). The other hits with lower scores were "Uncultured" *Bartonella* sp. and *Bartonella* sp. sequences. Thus, we made a BLAST search with one of the above

three similar *B. taylorii* sequences and unfortunately, it showed similarity only with the other two but none of the more than ten *B. taylorii* gltA sequences in the GenBank submitted by others. For this reason, we considered our sequences as *Bartonella* spp.

All identified pathogens and accession numbers are shown in Table 5.

## 4 | DISCUSSION

The green areas of cities, like suburban forests, cemeteries, and city parks, are suitable habitats for several wildlife species. These mammals can serve as hosts for ticks and tick-borne pathogens with medical and veterinary importance (Rizzoli et al., 2014). Our study sheds light on the diversity and composition of tick-borne bacterial communities in accidentally and road-killed urban mammals. These baseline data can be used in further eco-epidemiological studies and thus might help in the prevention of tick-borne pathogens.

Using road-killed animals as sources of tissue samples and ectoparasites provides both advantages and disadvantages. It is possible to collect samples not only from common species but also from internal organs of animals under protection and thus neglected in eco-epidemiological studies. A further advantage of using roadkill versus surveying wildlife through live capture or hunting is that both organization and logistics requires much less resources. In contrast, the main disadvantage is the varying condition of the carcasses. It is possible to collect dead animals in very good conditions, but carcasses are often dried samples with unidentifiable organs (except of skin) and bodies fully flattened by cars. The PCR reaction could be false-negative because of the high degree of degradation (inhibitors and DNA degradation) and the rise of decomposing bacteria. However, despite the highest degree of degradation (4 or 5) it was possible to amplify DNA of *A. phagocytophilum*, *R. helvetica*, and *Rickettsia* sp. in dried skin and muscle samples from road- and accidentally killed urban animals with real-time PCR. Therefore, even badly damaged carcasses might provide important baseline information on the presence and prevalence of a pathogen in urban habitats. The opportunistic manner of this sample collection does not allow, however, for the collection of all species of interest and, might also bias results. Since the number of specimens from each species collected cannot be determined in advance, species with limited number of samples allow no or only limited conclusions regarding the role of the host in the circulation of particular pathogen. We are also aware that some ectoparasites had already left the host by the time they were sampled, thus only a fraction of them will be found and processed.

We found six different pathogens in 90 collected tissue samples from eight mammal species (Table 2). Besides several single infections ( $n = 7$ ), double ( $n = 10$ ), triple ( $n = 5$ ), quadruple ( $n = 5$ ) infections, and one quintuple infection occurred in the collected animal tissues. Most of the multiple infections (28 tissue samples) were found in hedgehogs but double infections also occurred in the other collected mammal species (lesser weasel, European red squirrel, and house mouse) (Supporting Information Table S1).

Some road-killed mammals also proved to be a good source of ixodid ticks: 417 ticks were removed from nine carcasses, all of them were hedgehogs (362 *I. ricinus* and 55 *I. hexagonus*). The sample size in other species was small thus further sampling has to clarify whether hedgehogs tend to harbour more ticks compared to the other small mammal species in urban habitats. Four different pathogens were identified with real-time PCR within these ectoparasites (Table 4). While the pathogens detected from feeding ticks might only be part of their blood meal, the lack of *Can. N. mikurensis* and *B. miyamotoi* in them can mean that *E. roumanicus* has probably no (*B. miyamotoi*) or limited (*Can. N. mikurensis*) role in the maintenance of these bacteria as suggested by the present and previous (Földvári et al., 2014) findings regarding hedgehog tissues.

*Anaplasma phagocytophilum* ecotype I, which is the most prevalent human pathogenic ecotype, was found in northern white-breasted hedgehog tissue samples and in *I. ricinus* and *I. hexagonus* ticks. Some of the removed ticks from the *A. phagocytophilum*-positive hedgehogs were also infected with *A. phagocytophilum*. In addition, some hedgehogs were positive for *A. phagocytophilum* and their ticks were negative, and there were some positive tick samples from negative hedgehogs as well. These findings are not surprising, since transmission efficacy during feeding might be less than 100% and *A. phagocytophilum* has transstadial transmission (Sonenshine & Roe, 2014). These two ectoparasite species are the most prevalent ticks feeding on hedgehogs and they also have an important role in the eco-epidemiology of Lyme borreliosis (Rizzoli et al., 2014; Szekeres, Majláthová, Majláth, & Földvári, 2016). In a previous study about urban hedgehogs from a park in Budapest, Hungary *A. phagocytophilum* prevalence was similarly high (76%) as in the present study (74%) (Földvári et al., 2014). The data that the human pathogenic *A. phagocytophilum* ecotype I was detected in hedgehogs, and with high prevalence in both tick species removed from them (mean minimum infection rate: 56%) strongly suggest the maintenance role of *E. roumanicus* in the *A. phagocytophilum* cycles (Földvári et al., 2014; Jahfari et al., 2017). This is the first report of *A. phagocytophilum* in tissues from lesser shrew. Xenodiagnostic laboratory experiments could ascertain the reservoir role of these hosts (Kahl, Gern, Eisen, & Lane, 2002).

One roe deer fawn skin sample was positive with *A. phagocytophilum* specific real-time PCR. In roe deer *A. phagocytophilum* infection was reported in several European countries (Jahfari et al., 2014). Our finding is interesting, however, because the fawn hit by a car was at an early age (approximately 2 weeks), it had a short time-period to acquire infection via tick bite. During the dissection, no ectoparasite was found on this carcass. These circumstances suggest that transplacental transmission might be possible not just in sheep (Reppert et al., 2013) but also in wild ruminants like roe deer as well.

*Borrelia afzelii* was found in hedgehog skin and *I. ricinus* samples (female, male, nymph) and *B. spielmanii* in muscle samples from hedgehogs. Both *Borrelia* species cause erythema migrans in humans (Földvári, Farkas, & Lakos, 2005; Szekeres et al., 2015a,b). Similar with *E. europaeus*, *E. roumanicus* probably has reservoir role for Lyme borreliosis spirochaetes, which has not yet been demonstrated with xenodiagnostic experiments. A recent study from the Czech Republic

also identified *B. afzelii* as the most common genospecies in questing *I. ricinus* also within urban habitats (Kybicová, Baštová, & Malý, 2017).

*Rickettsia monacensis* was found in *I. ricinus* nymphs and one larval pool, and *R. helvetica* was detected in all stages of *I. ricinus* and *I. hexagonus* nymph samples with relatively high prevalence. These two bacteria are human pathogens (Fournier, Grunnenberger, Jaulhac, Gastinger, & Raout, 2000; Jado et al., 2007), belonging to the Mediterranean spotted fever group rickettsiae causing elevated fever, eschar, and maculopapular rash. In hedgehogs, moles, a mouse, a lesser weasel, and a stone marten, rickettsiae were detected. This is the first report about *Rickettsia* sp. infection in European moles and lesser weasel and about *R. helvetica* in stone marten. Xenodiagnostic experiments are required to determine whether these hosts play a reservoir role in the urban cycle of these pathogens.

We detected one *B. miyamotoi* positive spleen sample from a European red squirrel, a similar observation was recently reported from Belgium (Ruyts, Frazer-Mendelewska, Van Den Berge, Verheyen, & Sprong, 2017). The presence of *B. miyamotoi* and the high prevalence of *B. burgdorferi* s.l. (Pisanu et al., 2014) in red squirrels suggest this species might have a role in the cycles of these *Borrelia* species (Szekeres et al., 2016). *Borrelia miyamotoi* is an emerging pathogen reported from many countries in ticks, hosts, and also humans (Wagemakers, Staarink, Sprong, & Hovius, 2015). The reported number of human disease cases is increasing in the Northern Hemisphere (USA, Russia, Japan, Germany, the Netherlands) (Boden, Lobenstein, Hermann, Margos, & Fingerle, 2016; Chowdri et al., 2013; Gugliotta, Goethert, & Berardi, 2013; Hovius et al., 2013; Krause et al., 2013; Platonov et al., 2011; Sato et al., 2014).

Northern white-breasted hedgehogs, one house mouse, two European moles, and a lesser weasel were *Bartonella* spp. specific PCR positive. The eco-epidemiology of these bacteria such as vector species and reservoir animals are unknown.

In *I. hexagonus* nymphs we found for the first time *R. helvetica* and *Rickettsia* sp. pathogens. There was a report about rickettsiae infection in *I. ricinus* and *I. hexagonus*, but unfortunately it was not clear, whether *I. hexagonus* truly carried any *Rickettsia* pathogens (Giroud, Capponi, Dumas, & Rageau, 1965). The relatively high *A. phagocytophilum* (min. prev.: 22%), in *I. hexagonus* samples might indicate that, this tick species can have a role in the endophilic pathogen cycle of tick-borne pathogens (Szekeres et al., 2015a,b). However, as all *I. hexagonus* were removed from two *A. phagocytophilum*, *R. helvetica* and *Rickettsia* sp. real-time PCR positive hedgehogs (code: H4 and H16), the pathogens might as well be only part of the tick's blood meal. Xenodiagnostic experiments are needed to clarify the vector role of these tick species in the future.

## 5 | CONCLUSIONS

Environmental change in general, but urbanization specifically leads to a change in the dynamics of hosts, vectors, and pathogens. Therefore,

monitoring potential reservoirs, vectors, and pathogens within cities is recommended. Accidentally killed and road-killed animals are simple and reasonable resources (with known limitations) that can be utilized to this end. Our molecular analyses showed that hedgehogs, moles, shrews, squirrels, mice, martens, and weasels are carriers of one or several tick-borne pathogens in urban habitats. We also highlight the importance of documentation and assessment of pathogen prevalence and diversity in carrier hosts and their ectoparasites which is a prerequisite for the risk assessment for emerging infectious diseases (see DAMA protocol in (Brooks et al., 2014). Due to the high motility of these urban mammals within and between human settlements, we expect an enhanced spread of ticks and tick-borne pathogens that represent high risk wherever these mammals are found and as long as habitats are suitable for tick survival as well. Further studies should investigate their relative contribution to the maintenance and spread of these pathogens and specify their relative roles in the incidence of human tick bites and tick-borne diseases.

## ACKNOWLEDGEMENTS

S. Sz. was supported by the Hungarian Eötvös Scholarship and Eur-NegVec Cost Action TD1303. G. F. was supported by the János Bolyai Research Scholarship of the Hungarian Academy of Sciences. This study was supported by the grants 11475-4/2016/FEKUT and 12190/2017/FEKUTSTRAT of the Hungarian Ministry of Human Resources and the grant "In the light of evolution: theories and solutions" (GINOP-2.3.2-15-2016-00057). We are grateful for the help of all the volunteers who contributed in any way to the collection of the carcasses. We are grateful to Daniel R. Brooks and Mihály Földvári for proofreading and for their valuable comments.

## CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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## REFERENCES

- Boden, K., Lobenstein, S., Hermann, B., Margos, G., & Fingerle, V. (2016). *Borrelia miyamotoi* – Associated Neuroborreliosis in Immunocompromised Person. *Emerging Infectious Diseases*, 22, 1617–1620. <https://doi.org/10.3201/eid2208.152034>
- Brooks, D. R., Hoberg, E. P., Boeger, W. A., Gardner, S. L., Galbreath, K. E., Herczeg, D., ... Dursahinhan, A. T. (2014). Finding them before they find us: Informatics, parasites, and environments in accelerating climate change. *Comparative Parasitology*, 81, 155–164. <https://doi.org/10.1654/4724b.1>
- de Bruin, A., van Leeuwen, A. D., Jahfari, S., Takken, W., Földvári, M., Dremmel, L., ... Földvári, G. (2015). Vertical transmission of *Bartonella schoenbuchensis* in *Lipoptena cervi*. *Parasites and Vectors*, 8, 176. <https://doi.org/10.1186/s13071-015-0764-y>



- Chowdri, H. R., Gugliotta, J. L., Berardi, V. P., Goethert, H. K., Molloy, P. J., Sterling, S. L., & Telford, S. R. (2013). *Borrelia miyamotoi* infection presenting as human granulocytic anaplasmosis: a case report. *Annals of Internal Medicine*, 159, 21–27. <https://doi.org/10.7326/0003-4819-159-1-201307020-00005>
- Courtney, J. W., Kostelnik, L. M., Zeidner, N. S., & Massung, R. F. (2004). Multiplex real-time PCR for detection of *Anaplasma phagocytophilum* and *Borrelia burgdorferi*. *Journal of Clinical Microbiology*, 42, 3164–3168. <https://doi.org/10.1128/jcm.42.7.3164>
- Földvári, G., Farkas, R., & Lakos, A. (2005). *Borrelia spielmanii* Erythema Migrans, Hungary. *Emerging Infectious Diseases*, 11, 1794–1795. <https://doi.org/10.3201/eid1111.050542>
- Földvári, G., Jahfari, S., Rigó, K., Jablonszky, M., Szekeres, S., Majoros, G., ... Sprong, H. (2014). Candidatus *Neoehrlichia mikurensis* and *Anaplasma phagocytophilum* in Urban Hedgehogs. *Emerging Infectious Diseases*, 20, 496–498.
- Földvári, G., Rigó, K., Jablonszky, M., Biró, N., Majoros, G., Molnár, V., & Tóth, M. (2011). Ticks and the city: Ectoparasites of the Northern white-breasted hedgehog (*Erinaceus roumanicus*) in an urban park. *Ticks and Tick-borne Diseases*, 2, 231–234. <https://doi.org/10.1016/j.ttbdis.2011.09.001>
- Fournier, P.-E., Grunnenberger, F., Jaulhac, B., Gastinger, G., & Raoult, D. (2000). Evidence of *Rickettsia helvetica* infection in humans, Eastern France. *Emerging Infectious Diseases*, 6, 389–392. <https://doi.org/10.3201/eid0604.000412>
- Giroud, P., Capponi, M., Dumas, N., & Rageau, J. (1965). Les *Ixodes ricinus* et *hexagonus* de France contiennent des agents rickettsiens ou proches. *Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences*, 260, 4874–4876.
- Gugliotta, J. L., Goethert, H. K., Berardi, V. P., & S.R. 3rd Telford, (2013). Meningoencephalitis from *Borrelia miyamotoi* in an immunocompromised patient. *New England Journal of Medicine*, 368, 240–245. <https://doi.org/10.1056/NEJMoa1209039>
- Guy, E. C., & Stanek, G. (1991). Detection of *Borrelia burgdorferi* in patients with Lyme disease by the polymerase chain reaction. *Journal of Clinical Pathology*, 44, 610–611. <https://doi.org/10.1136/jcp.44.7.610>
- Hansford, K. M., Fonville, M., Jahfari, S., Sprong, H., & Medlock, J. M. (2015). *Borrelia miyamotoi* in host-seeking *Ixodes ricinus* ticks in England. *Epidemiology and Infection*, 143, 1079–1087. <https://doi.org/10.1017/S0950268814001691>
- Heylen, D., Tijssse, E., Fonville, M., Matthysen, E., & Sprong, H. (2013). Transmission dynamics of *Borrelia burgdorferi* s.l. in a bird tick community. *Environmental Microbiology*, 15, 6636–6673. <https://doi.org/10.1111/1462-2920.12059>
- Hillyard, P. (1996). *Ticks of North-West Europe*. Shrewsbury: Field Studies Council.
- Hornok, S., Földvári, G., Rigó, K., Meli, M. L., Gönczi, E., Répási, A., ... Hofmann-Lehmann, R. (2015). Synanthropic rodents and their ectoparasites as carriers of a novel haemoplasma and vector-borne, zoonotic pathogens indoors. *Parasites and Vectors*, 8, 27. <https://doi.org/10.1186/s13071-014-0630-3>
- Hornok, S., Kováts, D., Csörgő, T., Meli, M. L., Gönczi, E., Hadnagy, Z., ... Hofmann-Lehmann, R. (2014). Birds as potential reservoirs of tick-borne pathogens: First evidence of bacteraemia with *Rickettsia helvetica*. *Parasites and Vectors*, 7, 128. <https://doi.org/10.1186/1756-3305-7-128>
- Hovius, J. W. R., de Wever, B., Sohne, M., Brouwer, M. C., Coumou, J., Wagemakers, A., ... van Oers, M. H. J. (2013). A case of meningoencephalitis by the relapsing fever spirochaete *Borrelia miyamotoi* in Europe. *Lancet*, 382, 658. [https://doi.org/10.1016/S0140-6736\(13\)61644-X](https://doi.org/10.1016/S0140-6736(13)61644-X)
- Jado, I., Oteo, J. A., Aldámiz, M., Gil, H., Escudero, R., García-amil, C., ... Anda, P. (2007). *Rickettsia monacensis* and Human Disease, Spain. *Emerging Infectious Diseases*, 13, 1405–1407. <https://doi.org/10.3201/eid1309.060186>
- Jahfari, S., Coipan, E. C., Fonville, M., Docters van Leeuwen, A., Hengeveld, P., Heylen, D., ... Sprong, H. (2014). Circulation of four *Anaplasma phagocytophilum* ecotypes in Europe. *Parasites and Vectors*, 7, 365. <https://doi.org/10.1186/1756-3305-7-365>
- Jahfari, S., Fonville, M., Hengeveld, P., Reusken, C., Scholte, E. J., Takken, W., ... Sprong, H. (2012). Prevalence of *Neoehrlichia mikurensis* in ticks and rodents from North-west Europe. *Parasites and Vectors*, 5, 74. <https://doi.org/10.1186/1756-3305-5-74>
- Jahfari, S., Ruyts, S. C., Frazer-Mendelewska, E., Jaarsma, R., Verheyen, K., & Sprong, H. (2017). Melting pot of tick-borne zoonoses: The European hedgehog contributes to the maintenance of various tick-borne diseases in natural cycles urban and suburban areas. *Parasites and Vectors*, 10, 134. <https://doi.org/10.1186/s13071-017-2065-0>
- Kahl, O., Gern, L., Eisen, L., & Lane, R. S. (2002). Ecological research on *Borrelia burgdorferi* sensu lato: Terminology and some methodological pitfalls. In J. Gray, O. Kahl, R. S. Lane, & G. Stanek (Eds.), *Lyme borreliosis: Biology, Epidemiology and Control* (pp. 29–46). Bristol: CABI. <https://doi.org/10.1079/9780851996325.0000>
- Krause, P. J., Narasimhan, S., Wormser, G. P., Rollend, L., Fikrig, E., Lepore, T., ... Fish, D. (2013). Human *Borrelia miyamotoi* infection in the United States. *New England Journal of Medicine*, 368, 291–293. <https://doi.org/10.1056/NEJMc1215469>
- Krawczyk, A. I., van Leeuwen, A. D., Jacobs-Reitsma, W., Wijnands, L. M., Bouw, E., Jahfari, S., ... de Bruin, A. (2015). Presence of zoonotic agents in engorged ticks and hedgehog faeces from *Erinaceus europaeus* in (sub) urban areas. *Parasites and Vectors*, 8, 210. <https://doi.org/10.1186/s13071-015-0814-5>
- Kybicová, K., Baštová, K., & Malý, M. (2017). Detection of *Borrelia burgdorferi* sensu lato and *Anaplasma phagocytophilum* in questing ticks *Ixodes ricinus* from the Czech Republic. *Ticks and Tick-borne Diseases*, 8, 483–487. <https://doi.org/10.1016/j.ttbdis.2017.02.007>
- Nosek, J., & Sixl, W. (1972). Central-European Ticks (Ixodoidea) - Key for determination-. *Mitteilungen der Abteilung für Zoologie am Landesmuseum Joanneum*, 1, 61–92.
- Pisanu, B., Chapuis, J.-L., Dozières, A., Basset, F., Poux, V., & Vourc'h, G. (2014). High prevalence of *Borrelia burgdorferi* s.l. in the European red squirrel *Sciurus vulgaris* in France. *Ticks and Tick-borne Diseases*, 5, 1–6. <https://doi.org/10.1016/j.ttbdis.2013.07.007>
- Platonov, A. E., Karan, L. S., Kolyasnikova, N. M., Makhneva, N. A., Toporkova, M. G., Maleev, V. V., ... Krause, P. J. (2011). Humans infected with relapsing fever spirochete *Borrelia miyamotoi*, Russia. *Emerging Infectious Diseases*, 17, 1816–1823. <https://doi.org/10.3201/eid1710.101474>
- Rao, H. X., Yu, J., Guo, P., Ma, Y. C., Liu, Q. Y., Jiao, M., ... Li, D. M. (2015). Bartonella Species Detected in the Plateau Pikas (*Ochotona curzolaie*) from Qinghai Plateau in China. *Biomedical and Environmental Sciences*, 28, 674–678. <https://doi.org/10.3967/bes2015.094>. Nethe rlands
- Reeve, N. (1994). *Hedgehogs*. London: T & AD Poyser (Natural History).
- Reppert, E., Galindo, R. C., Breshears, M. A., Kocan, K. M., Blouin, E. F., & de la Fuente, J. (2013). Demonstration of transplacental transmission of a human isolate of *Anaplasma phagocytophilum* in an experimentally infected sheep. *Transboundary and Emerging Diseases*, 60, 93–96. <https://doi.org/10.1111/tbed.12120>
- Rizzoli, A., Silaghi, C., Obiegala, A., Rudolf, I., Hubalek, Z., Földvári, G., ... Kazimi-rová, M. (2014). *Ixodes ricinus* and its transmitted pathogens in urban and peri-urban areas in Europe: New hazards and relevance for public health. *Frontiers in Public Health*, 2, 251. <https://doi.org/10.3389/fpubh.2014.00251>
- Roux, V., & Raoult, D. (2000). Phylogenetic analysis of members of the genus *Rickettsia* using the gene encoding the outer-membrane protein rOmp (ompB). *International Journal of Systematic and Evolutionary Microbiology*, 50, 1449–1455. <https://doi.org/10.1099/00207713-50-4-1449>

- Ruyts, S. C., Frazer-Mendelewska, E., Van Den Berge, K., Verheyen, K., & Sprong, H. (2017). Molecular detection of tick-borne pathogens *Borrelia afzelii*, *Borrelia miyamotoi* and *Anaplasma phagocytophilum* in Eurasian red squirrels (*Sciurus vulgaris*). *European Journal of Wildlife Research*, *63*, 9–12. <https://doi.org/10.1007/s10344-017-1104-7>
- Sato, K., Takano, A., Konnai, S., Nakao, M., Ito, T., Koyama, K., ... Kawabata, H. (2014). Human infections with *Borrelia miyamotoi*, Japan. *Emerging Infectious Diseases*, *20*, 1391–1393. <https://doi.org/10.3201/eid2008.131761>
- Sonenshine, D., & Roe, R. (Eds.) (2014). *Biology of Ticks*. Oxford, UK: Oxford University Press.
- Stenos, J., Graves, S. R., & Unsworth, N. B. (2005). A highly sensitive and specific real-time PCR assay for the detection of spotted fever and typhus group Rickettsiae. *American Journal of Tropical Medicine and Hygiene*, *73*, 1083–1085. <https://doi.org/10.4269/ajtmh.2005.73.1083>
- Szekeres, S., Coipan, E. C., Rigó, K., Majoros, G., Jahfari, S., Sprong, H., & Földvári, G. (2015a). Eco-epidemiology of *Borrelia miyamotoi* and Lyme borreliosis spirochetes in a popular hunting and recreational forest area in Hungary. *Parasites and Vectors*, *8*, 309. <https://doi.org/10.1186/s13071-015-0922-2>
- Szekeres, S., Coipan, E. C., Rigó, K., Majoros, G., Jahfari, S., Sprong, H., & Földvári, G. (2015b). Candidatus *Neoehrlichia mikurensis* and *Anaplasma phagocytophilum* in natural rodent and tick communities in Southern Hungary. *Ticks and Tick-borne Diseases*, *6*, 111–116. <https://doi.org/10.1016/j.ttbdis.2014.10.004>
- Szekeres, S., Majláthová, V., Majláth, I., & Földvári, G. (2016). Neglected hosts: The role of lacertid lizards and medium-sized mammals in the eco-epidemiology of Lyme borreliosis. In M. A. H. Braks, vanWieren S. E., W. Takken & H. Sprong (Eds.), *Ecology and prevention of Lyme borreliosis* (Ecology and Control of Vector-borne diseases, Vol. 4) (pp. 103–126). Wageningen: Wageningen Academic Publishers. <https://doi.org/10.3920/227-1875-0699>
- Tolnai, Z., Sréter-Lancz, Z., & Sréter, T. (2015). Spatial distribution of *Anaplasma phagocytophilum* and *Hepatozoon canis* in red foxes (*Vulpes vulpes*) in Hungary. *Ticks and Tick-borne Diseases*, *6*, 645–648. <https://doi.org/10.1016/j.ttbdis.2015.05.009>
- Tóth-Ronkay, M., Bajor, Z., Bárány, A., Földvári, G., Görföl, T., Halpern, B., ... Vörös, J. (2015). Budapest. In J. G. Kelcey (Ed.), *Vertebrates and invertebrates of European cities: Selected non-avian Fauna* (pp. 26–73). Berlin: Springer Verlag.
- Wagemakers, A., Staarink, P. J., Sprong, H., & Hovius, J. W. R. (2015). *Borrelia miyamotoi*: A widespread tick-borne relapsing fever spirochete. *Trends in Parasitology*, *31*, 260–269. <https://doi.org/10.1016/j.pt.2015.03.008>
- Ying, B., Kosoy, M. Y., Maupin, G. O., Tsuchiya, K. R., & Gage, K. L. (2002). Genetic and ecologic characteristics of Bartonella communities in rodents in Southern China. *American Journal of Tropical Medicine and Hygiene*, *66*, 622–627. <https://doi.org/10.4269/ajtmh.2002.66.622>

## SUPPORTING INFORMATION

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**How to cite this article:** Szekeres S, van Leeuwen AD, Tóth E, Majoros G, Sprong H, Földvári G. Road-killed mammals provide insight into tick-borne bacterial pathogen communities within urban habitats. *Transbound Emerg Dis*. 2019;66:277–286. <https://doi.org/10.1111/tbed.13019>