

THE APPLICATION OF MAGNETIC RESONANCE IMAGING (MRI) TO THE EXAMINATION OF PLANT TISSUES AND WATER BARRIERS

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The aim of the present study is to extend the applicability of MRI measurements similar to those used in human diagnostics to the examination of water barriers in living plants, thus broadening their use in natural sciences. The cucumber, *Cucumis sativus*, and *Phillyrea angustifolia*, or false olive, were chosen as test plants. The MRI measurements were carried out on three samples of each plant in the same position vis-a-vis the MRI apparatus using a Siemens Avanto MRI scanner. Two different relaxation times were employed, T_1 , capable of histological mapping, and T_2 , used for the examination of water content. In the course of the analysis, it was found that certain histological formations and branching cause modifications to the intensity detected with relaxation time T_2 . Furthermore, these positions can also be found in T_1 measurements. A monotonic correlation (cucumber: $\rho = 0.829$; false olive: $\rho = -0.84$) was observed between the T_1 and T_2 measurements. In the course of the statistical analysis of the signal intensities of the xylems it was concluded that they cannot be regarded as independent in a statistical sense; these changes rather depend on the anatomic structure of the plant, as the intensity profile is modified by nodes, leaves and branches. This serves as a demonstration of the applicability of MRI to the measurement of well known plant physiological processes. The special parametrization required for this equipment, which is usually used in human diagnostics, is also documented in the present study.

Keywords: Magnetic Resonance Imaging (MRI) – nodes – water barrier – water content – xylem

INTRODUCTION

The technique known as Magnetic Resonance Imaging (MRI) is mostly associated with its use in human diagnostics. Its increasingly widespread use in areas of non-human research presents researchers with new opportunities, and it must be conceded that there is still “much ground to cover”. The application of nuclear magnetic resonance imaging (NMR) to plant studies is of great interest, as it allows for repeated, non-invasive investigation at the microscopic scale of living plants under well-defined environmental conditions [14]. It is also true that the non-destructive study of

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physiological processes in agricultural products requires increasingly higher spatial and temporal resolutions [25]. In this connection, Van As and van Duynhoven [45] give a detailed overview of MRI application in plant sciences and of the possibilities of application in food industry.

MRI – From human diagnostics to plant physiological research

The advantage of the application of MRI is that it visualizes the tissues of the examined plant in real time without causing anatomic dysfunction [30]. As in human diagnostics, the method is non-destructive, and measurements can therefore be repeated several times on the same sample. Besides their destructiveness, the other problem with classical methods of making physiological measurements of plants is accuracy [32]. This is because classical methods make it hard to detect changes on account of the effects of the direct environment on the test subjects. The use of MRI in research into plant physiology may well be the solution to this problem. MRI measures the quantity and the distribution of the protons, making it suitable for the determination of the inner structure of the plants and the examination of the microdynamics of their water content [18]. Both direct and indirect MRI methods exist for this [31]. Such NMR and n-MRI techniques have been described and applied by Van As and Windt [44]. These non-destructive, non-invasive methods have also been employed to study the dynamics of plant water relations, e.g. changes under environmental stress [44].

Application of MRI in plant studies

There have been numerous studies exploring the possibilities of the application of MRI in investigations in the physical-, chemical-, life-, and clinical sciences. For example, a study by Yang et al. [51] yielded important thermodynamic and kinetic data on the formation of CO₂ hydrate in flow systems, while Musse et al. [31] used MRI to quantify the apparent microporosity of apples and tomatoes. Quantitative MRI measurements have been used in several studies to investigate vegetal tissues [2, 6, 18]. Borisjuk et al. [7] provide a review of technological developments in MRI and the creation of opportunities for the three dimensional visualization and quantification of lipids in plant materials. In that study, the authors explain the importance of the imaging of lipids in living plants and further prospects for its use. NMR has also been used by Van der Weerd et al. [46, 47] to study the cell water balance in maize and pearl millet under osmotic stress. Mazhar et al. [27] give a detailed review of the application of MRI to the examination of fruit and vegetables, in which proton magnetic resonance imaging (¹H-MRI) has been applied since the 1980s to the non-destructive internal quality study of horticultural products. ¹H-MRI has also been employed in the determination of the state of maturity in, for instance, avocados [10]. It has also been used to study disorders in fresh produce, including e.g. heat injury in mangoes [22] and mechanical injury in avocados [37]. Hills and Clark [18]

review the general postharvest applications of ^1H -MRI, including for internal quality evaluation in other fruit and vegetables including blueberry, cherry, durian, melon, mangosteen, nectarine, papaya, pineapple, strawberry, tangerine, water melon, and zucchini.

Being non-invasive, MRI gives high resolution information on the quantity of hydrogen in tissue without loss of or damage to that tissue [3]. As such, it has proved highly adaptable in physiological research, not only in the case of fruit [35] and vegetables [29], but also trees [41]. MRI has been used to examine the maturing process of apples by Wang et al. [49], tomatoes by Ishida et al. [20] and Saltveit [36]. Several pieces of research conducted using MRI have had the aim of determining the nature of water transport and water distribution in the plant. Kuchenbrod et al. [25] conducted research on maize to examine the variation in flow conditions connected to transpiration at rates of between 1 and 10 mm s⁻¹. This research was intended to enable the visualization of the water content/condition in the plant [39]. Research has also been carried out on cucumber plants using portable NMR to measure the water transport in situ in xylem [38, 41–43].

Environmental aspects

Water plays an essential role in plant development [40]. According to the Fifth Report of Intergovernmental Panel on Climate Change (IPCC), changes in the global water cycle in the 21st century will not be uniform the world over. The contrast in precipitation between wet and dry regions and between wet and dry seasons will increase, although there may be regional exceptions [19]. Prista et al. [34] worked out chronostratigraphic analogies for IPCC scenarios, and came to the conclusion that the Pliocene (Mid Piacenzian Warm Period) offers the best analogue for a warming climate in Europe nowadays. The regional climate modelling results of the PRUDENCE project [4] showed that a change in precipitation may be expected in the Carpathian Basin, both in terms of quantity and distribution in the first decades of 21st century. Apart from the fact that simulation results suggest that total annual precipitation has not changed to any great degree, changes significant in their size and opposite in their trends are to be expected in different seasons. The amount of seasonal precipitation is expected to increase in winter and decrease in summer. This may well culminate in the total restructuring of the annual distribution of precipitation [4]. Such regional precipitation changes as those already expected may cause permanent water stress to plants, in addition to any increase in heat stress that will likely be part of any such scenario.

Several abiotic stress factors have negative effects on the plant-water relationship [13, 48]. Therefore, if the growth of plants is to be facilitated through scientific efforts, the detailed investigation of the relationship between the plant and its water uptake is a priority. Plant responses to changing environmental conditions are discussed in Brüggemann et al. [8], while the impact of anoxic conditions on phloem and xylem transport processes are discussed by Peuke et al. [33], as measured using MRI

flowmetry. In plants, the transport and accumulation of water and dry matter in different organs and tissues is a major determinant on their growth and final composition [16, 23]. For this reason, it is important to review the functions of the recurring elements on the plant's stem. These are the following: nodes, internodes, leaf fundaments and axillary buds [28]. Of these, the internodes play a key role in plant growth [17]. Beyond this, several other factors that cannot be disregarded influence plant development. One is the minerals supplied by the soil. In this regard, the zinc content of the plant is important [9] in processing the auxin hormone [12]. Another factor worth mentioning is the plant's exposure to light. [26].

Aims

The aim of the present study is to highlight the applicability of MRI measurements of the type used in human diagnostics to the examination of water barriers in living plants, as part of the expansion of a new frontier in the natural sciences. It should be emphasized that the method employed in this study is not the usual one used in plant physiological studies. Rather, it is an MRI technique used in human diagnostics specifically tuned (that is, parametrized in a series of trials) for the purposes of this specific piece of plant physiological research

MATERIALS AND METHODS

The basis of MRI measurements is the interaction between the external magnetic field, the electromagnetic waves and the hydrogen atoms of the material. MRI measures the quantity and distribution of the protons. Water is among the materials with relatively high proton content, making it suitable for the examination of the plant-water relationship using MRI. It should be noted that MRI does not detect the particular anatomic structure, but the quantity and distribution of water [21, 50] on the basis of the spin system in living plants. This, in turn, engages in just a weak interaction with the examined biological system's macroscopic parameters, that is, those which influence its biological and chemical behavior. Therefore, such measurements make it possible to draw conclusions concerning the behavior of the examined biological system [5, 9, 15].

The measurements were carried out at the Kaposvár University Institute of Diagnostic Imaging and Radiation Oncology using a Siemens Avanto MRI apparatus of the kind used in human diagnostics, and capable of producing a 1.5 T magnetic field (Fig. 1).

The applied sequence is a medley of the sequences used in human diagnostics, specifically, those performed on the cranium, ankle and knee [21], each with its particular setting (Table 1). These were obtained from a set of trials in which the factory settings corresponding to human diagnostics were adjusted on the basis of the optimization of the localized images. In terms of their mass, the plant samples are much

Table 1

Specific settings used in the MRI measurements tuned for plant physiological purposes, the analysis of cucumber and false olive

MRI Setting		Name of the plant	
		Cucumber	False olive
T1	Repetition time (sec)	1160	1160
	Echo time (sec)	42118	3.94
	Pixel spacing (mm)	0.36	0.97
	Slice thickness (mm)	0.69	0.89
	Resolution (pixel)	0.089	0.837
T2	Repetition time (sec)	42315	42151
	Echo time (sec)	42131	2.38
	Pixel spacing (mm)	0.35	0.976
	Slice thickness (mm)	1	0.7
	Resolution (pixel)	0.122	0.666

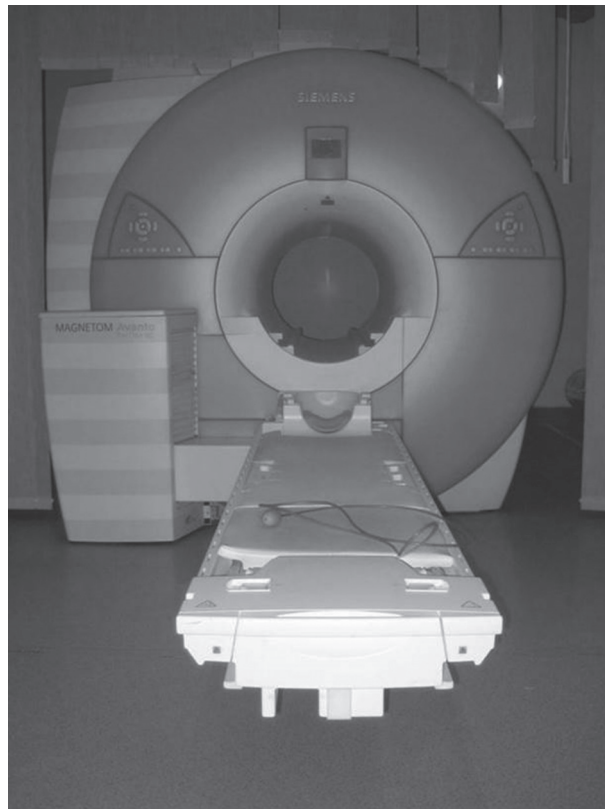


Fig. 1. Siemens Avanto MRI apparatus

smaller than the human body, with much less total water content. Therefore, a cranial coil was used, enabling investigators to focus in on the sample. This also meant that it was capable of a slice thickness of less than one millimeter (Table 1, T1: slice thickness).

The test plants were grown under different conditions. The cucumber was grown in a modified Hoagland nutrition solution [24], and the false olive in a container (Fig. 2). The cucumber plants grew to a height of 15–20 cm and the false olive to 30–35 cm.

The intensity of the detected signal in the xylem elements was assessed as a function of distance from the root. Stochastic connections were determined between the xylem elements inside the plant with the aim of examining the water-plant interac-

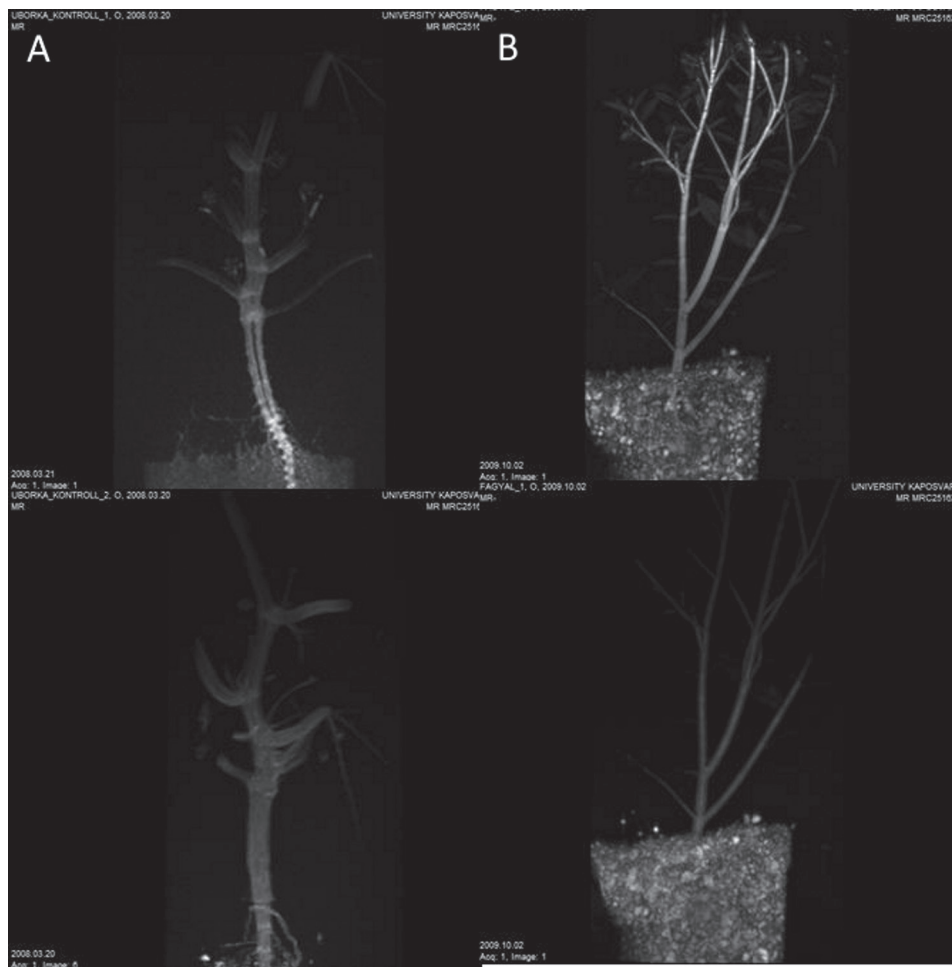


Fig. 2. MRI pictures of cucumber A) and false olive B), in which the upper panel represents the T_1 and lower the T_2 measurement

tion. Water barriers caused by nodes and differences in the intensities caused by the anatomic structure were detected using T_1 measurements; these were then correlated with the results of the T_2 measurements. Transverse relaxation time (T_2) is known to be related to the water status of cell compartments, i.e. their water content, water mobility, as well as the interactions between water and macromolecules [2, 31]. So-called T_2 -weighted ^1H MR images produce regions of higher signal intensity (hyperintense) from tissue containing mobile water, whereas water in environments with restricted mobility gives rise to a reduced signal intensity (hypointense) [1, 27].

Statistics

Addinsoft's XLSTAT 2017's test for goodness of fit was employed to see if sample data displayed a normal distribution. The relationship was evaluated using Spearman's ρ coefficient of correlation. LOWESS (locally weighted scatterplot smoothing [11]) non-parametric regression was used to determine the relationship between the distance of the root and the T_1 and T_2 relaxation times, respectively. The coefficient of determination of non-parametric regression is used to show the percentage of the total deviation explained by distance from the root at T_1 and T_2 .

RESULTS AND DISCUSSION

Examination of the xylem of cucumber

Xylems in the hypocotyls of cucumber were examined one by one using MRI. In this part of the branch the vascular tissues may be easily seen. As the distribution of the data is significantly different from a normal distribution (p -value < 0.0001 for T_1 , and $p < 0.0174$ for T_2), a non-parametric regression method was chosen to determine the relationship between the relaxation time values of T_1 (activity of the cells) and T_2 (water content). Spearman's ρ was 0.829, indicating a significant strong positive monotonic correlation between T_1 and T_2 relaxation times (p -value < 0.0001). The coefficient of determination of the non-parametric regression was 67.9% (Fig. 3a).

Examination of the xylem of false olive

After segmenting the branch part of the false olive, since the plant has both main and lateral stems, the two sides could be differentiated in the MRI images, as the curves of the intensity signals partly diverged approaching the branch apex. Further examinations were carried out on the main branch. The intensity signals on the two sides of the branch were found to be not independent of each other. The distribution of the data is significantly different from that characteristic of a normal distribution (p -value < 0.0001 for both relaxation times). Consequently, a non-parametric regres-

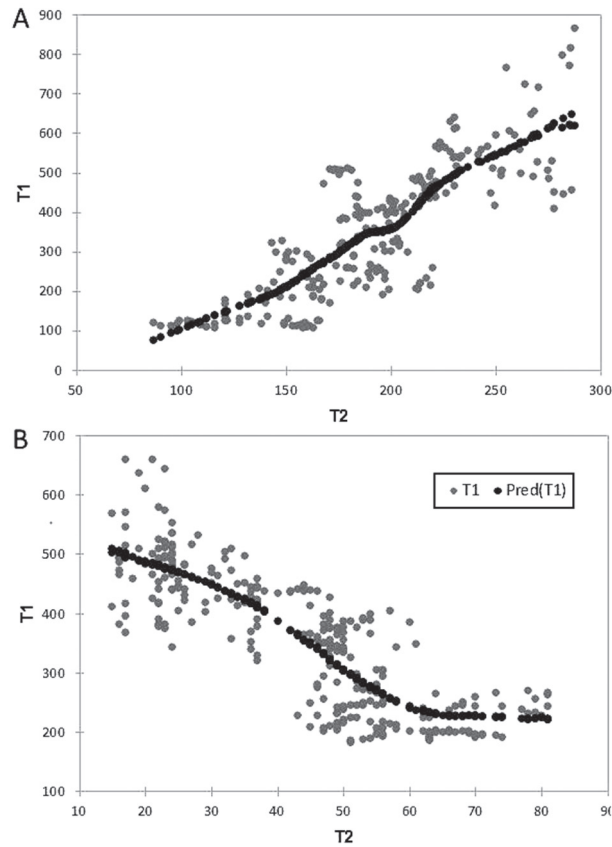


Fig. 3. Non-parametric regression between T_1 and T_2 measurements of cucumber A and false olive B

sion method was used to determine the relationship between the values of T_1 (activity of the cells) and T_2 (water content) relaxation times. Spearman's ρ was -0.84 , indicating a significant (p -value < 0.0001) and strong negative monotonic correlation between T_1 and T_2 relaxation times. The coefficient of determination given by LOWESS was 69.6%, and a negative relationship between the variables is observable (Fig. 3b).

Detection of anatomic structure and its interpretation

Anatomical differences that cause local maximum and minimum points in the intensity signals of T_2 measurements could be detected by T_1 measurements. Using the two types of measurement technique, the location of the nodes on the branch causing local

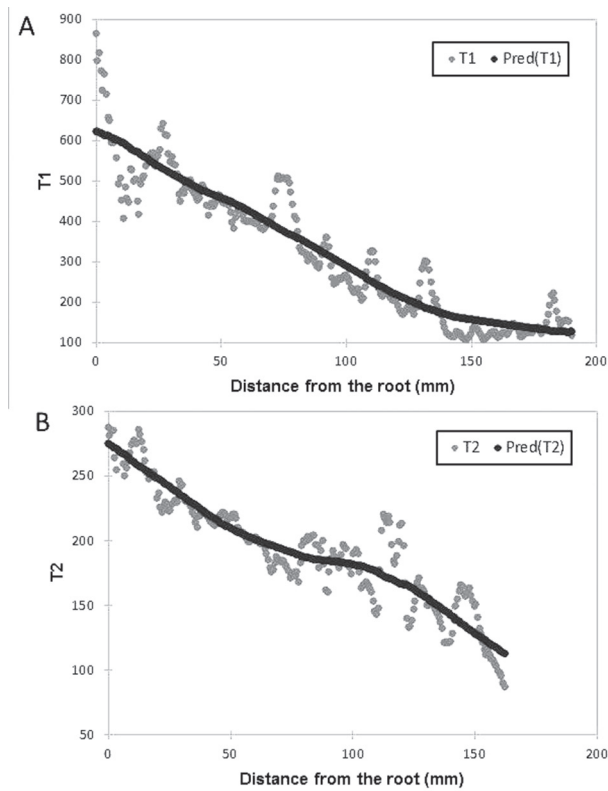


Fig. 4. The intensity signal of T_1 A and T_2 B measurements as a function of the distance from the root for cucumber

maxima was determined. Local maximum points were found at heights of 26.22 mm, 71.76 mm, 91.08 mm, 109.02 mm, and 130.41 mm, respectively (Fig. 4) in the case of cucumber. Spearman's ρ was -0.942 , indicating a significant (p -value < 0.0001) strong negative monotonic correlation between the distance from the root and T_1 values. Moreover, ρ was -0.918 in the case of the distance from the root and T_2 values, indicating a significant (p -value < 0.0001) strong negative monotonic correlation as well. A non-parametric regression was applied to describe the relationship between the distance from the root and the relaxation times (Fig. 4). The coefficient of determination using LOWESS between the distance from the root and T_1 and T_2 values was 89% and 86.6%, respectively.

Nodes were also determined for the false olive at several locations along the stem. Local maxima and minima in the signal intensity were detected using the data from the T_1 measurements. Nodes were found at heights of 10.5 mm, 25.2 mm, 39.9 mm, 60.2 mm, 72.8 mm, 86.1 mm, 102.2 mm, 118.3 mm, 128.1 mm, 154.7 mm and 186.2 mm (Fig. 5). Spearman's ρ was 0.903, indicating a significant (p -value < 0.0001)

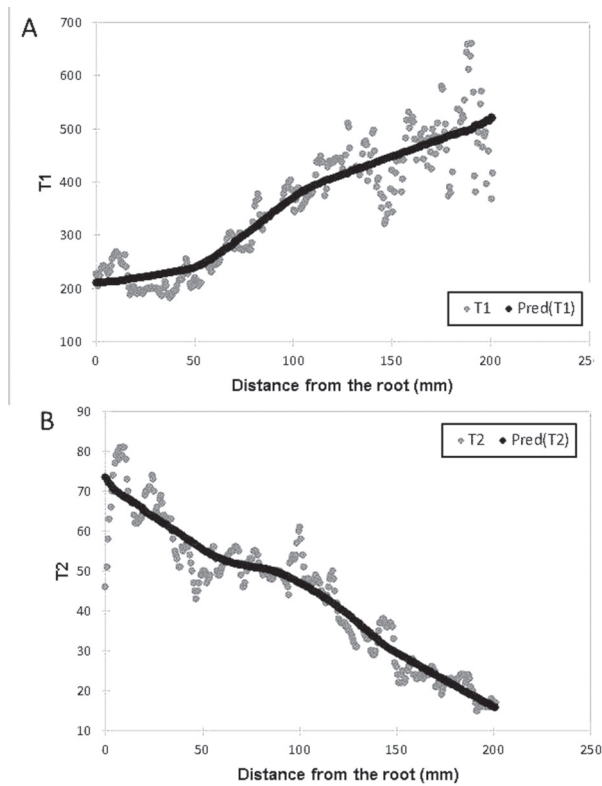


Fig. 5. The intensity signal of T_1 A and T_2 B measurements as a function of the distance from the root for false olive

strong positive monotonic correlation between the distance from the root and T_1 values. This figure was $\rho = -0.938$ between the distance from the root and T_2 values indicating a significant ($p\text{-value} < 0.0001$) strong negative monotonic correlation. A non-parametric regression method was applied to the description of the relationship between the distance from the root and the relaxation times (Fig. 5). The coefficient of determination between the distance from the root and T_1 and T_2 values was 85.2% and 91%, respectively.

In the course of the examination of the two plant species, it was demonstrated that MRI signal intensity (which is in direct relationship with the water content) decreased from the root upwards, towards the branch apex. In general, the water content of the plant is not independent of its height. The various water barriers deriving from a plant's anatomy are caused by the special structure of nodes. These anatomic elements and their effects can be detected and determined using MRI. Intensity signal data deriving from the T_1 and T_2 measurements show high correlations for both of the plant species used in the research.

Functional MRI has also been used to study transients of biophysical parameters in a cucumber plants in response to environmental changes [39]. The detailed flow imaging experiments showed the location of xylem and phloem in the stem, as well as highlighting the changes in the biophysical parameters of plants in response to environmental changes [39]. The results of the measurements made in this study concerning changes in the water content of the plants (T_2 measurements) are parallel to the findings of these investigations.

A better understanding is needed of the mechanism of phloem transport, the factors that determine the allocation of carbon to various organs, and the relation between xylem anatomy and conductivity [45]. In the T_2 measurements in the present research, the anatomical structure of xylem elements appear to affect water transport. This knowledge may in turn assist in the better understanding of phloem transport.

CONCLUSIONS

The present study demonstrates that, besides their use in human diagnostics, the utility of MRI techniques extends to the investigation of plant physiology. In this respect, it represents a novel tool in the hands of the researchers into not only plant physiology, but to the integrated examination of soil, root, stem and leaf water content, root anatomy, and (radial and axial) transport in roots, the stem, and leaves.

With this approach, accurate, high resolution measurements could be made in a non-destructive way, allowing the measurement of a given specimen several times. Moreover, it was possible to detect the exact anatomic elements of the plants at a scale of less than one millimeter. Measurements were made using complementary T_1 and T_2 relaxation time techniques. For the interpretation of the maximum and minimum values of T_2 data, T_1 measurements were also necessary. It was demonstrated that the curve of the intensity signal of xylem elements depends on the anatomic structure, and that the intensity profile was modified by the nodes, leaves and the branches.

With the application of the MRI used in human diagnostics to plant physiological studies, researchers gain the possibility of exploiting the capacity of MRI equipment in medical centers and hospitals, as in the case of the present research.

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