The Biology of *Sesamia inferens* (Walker) (Lepidoptera: Noctuidae) on Maize in the North Western Plains of India

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(Received: 17 August 2018; accepted: 7 November 2018)

The biology of *Sesamia inferens* (Walker) was studied at 25 ± 1 °C and 70 ± 5 per cent relative humidity on PMH 1 maize hybrid. The incubation period was 6.82 ± 0.05 days. There were six larval instars and the larval development was completed in 29.95 ± 0.16 days. The duration of instars I-VI was 4.17 ± 0.09 , 3.60 ± 0.08 , 4.47 ± 0.02 , 4.40 ± 0.03 , 6.18 ± 0.06 and 7.13 ± 0.05 days, respectively. The pre-pupal period was 2.83 ± 0.02 days. The male and female pupa had duration of 8.05 ± 0.12 days and 10.33 ± 0.16 days, respectively. The adult emergence was 93.02 ± 0.01 per cent, with a sex ratio of 1:1.05. The males had shorter longevity of 3.92 ± 0.23 days, while females had 5.05 ± 0.28 days. The pre-oviposition, oviposition and post-oviposition period was 1.11 ± 0.10 , 2.96 ± 0.19 and 0.79 ± 0.10 days, respectively. The fecundity was 211.92 ± 11.92 eggs with 53.69 ± 10.78 eggs per cluster. The egg hatchability was 92.19 ± 0.01 per cent. The total life cycle was completed in 47.65 ± 0.24 days (Male) and 49.93 ± 0.21 days (female). The observations on the biology will help in developing efficient strategies to manage *S. inferens* on maize in the north western plains of India.

Keywords: Biology, instar, maize, pink stem borer.

Maize (*Zea mays* L.; Poaceae) is a versatile crop grown over a wide range of agro climatic zones. Advancement of agricultural technologies enabled the cultivation of maize throughout the year in different parts of India. This coupled with improper pesticide usage has led to the wide spread occurrence of various pests in the non conventional areas. Pink stem borer, *Sesamia inferens* (Walker) is a polyphagous pest attacking various graminaceous crops. The species was originally described in the genus *Leucania* by Walker in 1856, which got later transferred to the genus *Sesamia* (Hampson et al., 1912). It is a key pest of winter maize in the peninsular India (Sekhar et al., 2005) causing dead heart in young and stem tunneling in grown up plants. The losses due to pink stem borer in maize in India vary from 25-80 per cent (Rao et al., 1983). It was reported in the borer complex in Kharif maize in Punjab (Singh et al., 2014). The pest has assumed major pest status in recent time in cereals (Baladhiya et al., 2018). It may pose a serious threat to maize in the rice-wheat/maize based cropping system in Punjab (Singh, 2013). Understanding the detailed biology of the pest on maize is a key to its successful management and sustained maize production in this region.

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Materials and Methods

The detailed aspects of life cycle of *S. inferens* were studied in 2015 at 25 ± 1 °C and 70 ± 5 per cent relative humidity under Maize Entomology Laboratory, Punjab Agricultural University (PAU), Ludhiana.

Insect culture

The larvae were collected from maize fields and were reared in artificial diet as per Kumar and Siddiqui (1993). The adults emerged were made to oviposit on 4–5-day maize seedlings. The seedling pots containing eggs were replaced regularly with fresh pots. The neonates emerged from the eggs (F_1 population) were reared on artificial diet. The adults emerged were released on maize seedlings to collect the eggs. This F_2 population was used for studies on biological parameters. There were 12 replications each having five individuals. The seeds of hybrid PMH 1 were sown in the field as well as in small earthen pots at weekly intervals and maintained to ensure the availability of maize plants throughout the rearing process.

Egg

The number of eggs in each cluster was recorded. The dimensions of the egg (mm) were measured with ocular and stage micrometer.

Larva

The larvae were reared in 2–2.5 inch long soft stem of 20–25-day-old maize plants individually in Petri dishes and developmental changes were recorded. The body length and breadth were measured using measuring scale. The weight of each instar was recorded using electronic weighing machine (Axis® – AGN 204 PO). The length (vertical) and width (horizontal) of the molted head capsule were taken using ocular micrometer.

Рира

The pupae formed were transferred into battery jars lined with moist butter paper having a thin layer of sterile moist sand at the bottom. The jars were covered with cotton cloth. The pupae were sexed and their length, breadth, weight and duration were recorded and kept for adult emergence.

Adult

The adults emerged were sexed and the length and breadth of the body were taken. Adult pairs were released on seedlings in ovipositional cages. The adults were observed for mating and egg laying. The mean and standard error of different biological parameters were calculated.

Results

Egg

The eggs were semi globular dorsoventrally flattened, with radial ridges (Fig. 1). The eggs have an average length of 0.616 ± 0.018 mm and breadth of 0.597 ± 0.012 mm (Table 1) (Fig. 2). The freshly laid eggs were creamy white, after 48 hours, head of the embryo became visible. After 72 hours, slight pinkish colour developed with the greyish head visible prominently (Fig. 3). The pinkish colour got relatively darker after 96 hours. After 120 hours of incubation, the eggs changed into greyish black and pink with the black head of the embryo visible clearly. Further, the neonates emerged out of the eggs. The incubation period averaged to 6.82 ± 0.05 days. The eggs recorded 92.19 ± 0.01 per cent hatchability.



Fig. 1. Egg cluster of *Sesamia inferens* on maize $(\times 6)$

Table 1

Characters of eggs of S. inferens on maize

Egg parameters	Range	Mean ± SE	
Length (mm)	0.574-0.660	0.616 ± 0.018	
Breadth (mm)	0.502-0.646	0.597 ± 0.012	
Incubation period (days)	5–7	6.82 ± 0.05	
Eggs per cluster (Nos)	10-102	53.69 ± 10.78	
Hatchability (%)	90–96	92.19 ± 0.01	



Fig. 2. Dimensions of eggs of Sesamia inferens (×8) (A random representative sample)

Eggs were laid in batches, in 2-4 long rows. The adult females laid eggs in the inner side of the first three leaf sheaths, preferably in the first leaf sheath. Number of eggs per cluster varied from 10 to 102, with an average of 53.69 ± 10.78 eggs (Table 1).

Larva

The larva passed through six larval instars. The newly hatched larva was cream in colour with black head. The pink colour started developing on the larval body at the second instar and by fourth instar the colour became prominent. The molting pattern of the larvae is described (Fig. 4). The head capsule of the newly molted larvae was light yellow-ish cream, which gradually changed to light brown, dark brown and finally greyish black.

The duration of the larval instars I–VI was 4.17 ± 0.09 , 3.60 ± 0.08 , 4.47 ± 0.02 , 4.40 ± 0.03 , 6.18 ± 0.06 and 7.13 ± 0.05 days, respectively (Table 2). The total larval development was completed in 29.95 ± 0.16 days. The length of larval instars I–VI averaged to 2.10 ± 0.06 , 4.34 ± 0.25 , 8.83 ± 0.31 , 14.50 ± 0.42 , 20.83 ± 0.55 and 27.17 ± 0.49 mm, respectively. The breadth of larval instars I–VI were 0.49 ± 0.02 , 0.67 ± 0.03 , 2.00 ± 0.13 , 2.35 ± 0.10 , 2.92 ± 0.90 and 3.48 ± 0.05 mm, respectively. The different larval instars had an average weight of 0.18 ± 0.02 , 4.20 ± 0.68 , 34.48 ± 5.01 , 70.99 ± 2.89 , 123.68 ± 5.80 and 223.58 ± 6.61 mg, respectively. The molted head capsule of the instars I–VI measured 0.429 ± 0.005 , 0.488 ± 0.008 , 0.849 ± 0.010 , 0.997 ± 0.013 , 1.518 ± 0.014 and 1.944 ± 0.008 mm length (vertical). The width (horizontal) of the molted head capsule of six larval instars was 0.38 ± 0.006 , 0.46 ± 0.004 , 0.78 ± 0.015 , 0.88 ± 0.005 , 1.31 ± 0.031 and 1.79 ± 0.015 mm, respectively (Fig. 5). The larvae stopped feeding and a few white

tion (days)		$Mean \pm SE$	4.17 ± 0.09	3.60 ± 0.08	4.47 ± 0.02	4.40 ± 0.03	6.18 ± 0.06	7.13 ± 0.05	29.95 ± 0.16
Durat		Range	3-5	3-4	4-5	4-5	5-7	6-8	25-34
Head Capsule	Breadth (mm)	Mean± SE	0.38 ± 0.006	0.46 ± 0.004	0.78 ± 0.015	0.88 ± 0.005	1.31 ± 0.031	1.79 ± 0.015	Ι
		Range	0.353-0.407	0.448 - 0.488	0.678 - 0.820	0.841 - 0.909	1.098 - 1.506	1.643 - 1.861	I
	Length (mm)	Mean± SE	0.429 ± 0.005	0.488 ± 0.008	0.849 ± 0.010	0.997 ± 0.013	1.518 ± 0.014	1.944 ± 0.008	I
		Range	0.407-0.475	0.461 - 0.529	0.773-0.881	0.949 - 1.086	1.456 - 1.587	1.890 - 1.992	I
Body	Weight (mg)	Mean± SE	0.18 ± 0.02	4.20 ± 0.68	34.48 ± 5.01	70.99 ± 2.89	123.68 ± 5.80	223.58 ± 6.61	I
		Range	0.1 - 0.3	2.7-11.6	17.0 - 68.0	50.5 - 84.4	100.0 - 165.2	182.0-254.0	I
	dth (mm)	Mean± SE	0.49 ± 0.02	0.67 ± 0.03	2.00 ± 0.13	2.35 ± 0.10	2.92 ± 0.90	3.48 ± 0.05	I
	Brea	Range	0.4 - 0.6	0.5 - 0.8	1.5 - 2.5	2.0 - 2.5	2.5 - 3.0	3.7-4.0	
	h (mm)	Mean±SE	2.10 ± 0.06	4.34 ± 0.25	8.83 ± 0.31	14.50 ± 0.42	20.83 ± 0.55	27.17 ± 0.49	I
	Lengt	Range	1.4–3.1	3.3-5.8	5.9 - 10.8	12.1–16.8	20.0-23.0	26.0-29.0	I
Instar			I	П	Ξ	\geq	>	ΙΛ	Total

Morphometrics and duration of different instars of S. inferens on maize

Table 2











Fig. 3. Sequence of changes in the eggs of *Sesamia inferens* (a) Freshly laid eggs in cluster;
(b) Eggs changed to pink colour; (c) Dark pink coloured eggs;
(d) Greyish black and pink eggs with head of embryo visible properly and neonates started emerging;
(e) Empty egg shell left after the emergence of neonates



Fig. 4. Molting pattern of larvae of *Sesamia inferens* (a)–(d) Larval molting;
(e) Molted larva with brown head capsule; (f) Brown head capsule turned to black;
(g) Molted head capsule; (h) Exuvia of *Sesamia inferens*



Fig. 5. Dimensions of larval head capsule of *Sesamia inferens* (a)–(f) Molted head capsules of 1–6 larval instars (Random representative samples)

Table 3

Parameters	Range	Mean ± SE	
Pre-pupal duration (days)	2–3	2.83 ± 0.02	
Pupal duration (days)			
Male	7–10	8.05 ± 0.12	
Female	8-12	10.33 ± 0.16	
Pupal length (mm)			
Male	11.07–13.50	12.17 ± 0.22	
Female	13.80-16.10	14.59 ± 0.28	
Pupal breadth (mm)			
Male	2.82-4.10	3.15 ± 0.10	
Female	3.50-5.00	4.01 ± 0.20	
Pupal weight (mg)			
Male	102.10-173.50	130.60 ± 4.30	
Female	153.70-260.60	194.30 ± 8.30	
Total life cycle (days)			
Male	39–54	47.65 ± 0.24	
Female	40–56	49.93 ± 0.21	

Pre-pupal and pupal characters and total life cycle of S. inferens on maize

silken threads were produced around the body. The body gradually got shrunk length wise inside the silken web, finally becoming slightly curved (Fig. 6). The pre-pupal period lasted for 2.83 ± 0.02 days (Table 3).

Pupa

The pre-pupa gradually changed to obtect pupa. The male pupa had a length of 12.17 ± 0.22 mm and a breadth of 3.15 ± 0.10 mm (Table 3) (Fig. 7). The length of female pupa was 14.59 ± 0.28 mm and the breadth was 4.01 ± 0.20 mm. The male pupa weighed 130.60 ± 4.30 mg while, the female weighed 194.30 ± 8.30 mg. The structures like, the eye, antenna, forewings and the abdomen region of the future adult were clearly visible in the pupa (Fig. 8). The pupal period of male and female were 8.05 ± 0.12 days and 10.33 ± 0.16 days, respectively. The life cycle of *S. inferens* (Fig. 10) was completed in 47.65 ± 0.24 (39–54) days in males and 49.93 ± 0.21 (40–56) days in females (Table 3).

Adult

The colour of *S. inferens* varied from light straw to medium brown (Fig. 9). Males had smaller body size than females. The males had a body length of 12.50 ± 0.41 mm and width of 3.40 ± 0.11 mm (Table 4), weighing 107.78 ± 3.47 mg. They had a wing expanse of 21.41 ± 0.19 mm with a single wing length of 12.45 ± 0.23 mm. The body length and width of female was 15.55 ± 0.18 and 4.60 ± 0.13 mm, respectively with a weight of 188.52 ± 10.53 mg. The females had a larger wing expanse of 31.00 ± 0.36 mm and a single wing length of 14.57 ± 0.19 mm. The longevity of male was 3.92 ± 0.23 days and that of female was 5.05 ± 0.28 days (Table 4). The pre-oviposition period of fe-

Table 4

Adult parameters	Ma	ale	Female		
	Range	Mean \pm SE	Range	Mean \pm SE	
Length (mm)	10.0-14.5	12.50 ± 0.41	14.5-16.5	15.55 ± 0.18	
Breadth (mm)	3-4	3.40 ± 0.11	4.0-5.5	4.60 ± 0.13	
Weight (mg)	70.30-156.46	107.78 ± 3.47	102.00-363.70	188.52 ± 10.53	
Wing expanse (mm)	20-22	21.41 ± 0.19	29-33	31.00 ± 0.36	
Single wing length (mm)	11.0-13.4	12.45 ± 0.23	13.4-15.5	14.57 ± 0.19	
Longevity (Days)	3-5	3.92 ± 0.23	4-6	5.05 ± 0.28	

Morphometrics and longevity of adults of S. inferens on maize

Table 5

Biological parameters of S. inferens on maize

Parameter	Period (Days)		
	Range	Mean ± SE	
Pre-oviposition period	1.1–1.7	1.11 ± 0.10	
Oviposition period	2.0-4.0	2.96 ± 0.19	
Post-oviposition period	0.5-1.2	0.79 ± 0.10	
Adult emergence (%)	91.67–95.65	93.02 ± 0.01	
Sex ratio (Male: Female)	1:1-1:1.8	1:1.05	
Fecundity (Nos. per female)	152–286	211.92 ± 11.92	

males was 1.11 ± 0.10 days (Table 5). The oviposition period was 2.96 ± 0.19 days and the post-oviposition period was 0.79 ± 0.10 days. Peak period of mating occurred in the night at 21.00-24.00 hours and lasted for about 1 hour. The females laid eggs on the inner side of basal leaf sheaths in 3 to 4 non-overlapping rows.

An adult emergence of 91.67-95.65 per cent was recorded with an average of 93.02 ± 0.01 per cent (Table 5). The male to female sex ratio was found to vary from 1:1 to 1:1.18 with an average of 1:1.05. The fecundity varied from 152-286 eggs with an average of 211.92 ± 11.92 eggs. The maximum number of eggs was laid on the second day of emergence.

Discussion

The variability in biology of *S. inferens* was reported on different host plants under variable agroclimatic conditions. The morphometric features of the eggs of *S. inferens* reported by Aggarwal et al. (2004), Rajendra (1976) and Singh (2013) on different crops were in line with the present study. However, variation in the incubation period is documented. Sekhar et al. (2005) reported a comparable incubation period of 5–8 days on maize. The longer period of 6–11 days during October–November (Aggarwal et al., 2004) and 7.40 ± 0.38 days during November–January (Singh, 2013) may be attributed to the low temperature. The slower embryonic development at lower temperature (Viswajyothi et al., 2017) might have resulted in delayed development. However, egg hatchability corroborates with reports of Selvaraj et al. (2014) and Singh (2013). The



Fig. 6. Pre-pupa of Sesamia inferens



Fig. 7. Pupa of *Sesamia inferens* (a) Male pupa (×10); (b) Female pupa (×8) (A random representative sample)



Fig. 8. Body regions of pupa (a) Male; (b) Female

number of eggs per cluster showed variability. Similar to present studies, Aggarwal et al. (2004) reported 15 to 91 eggs per cluster with a mean of 58.84 ± 3.60 eggs on rice but on contrary Rajendra (1976) reported 50–150 eggs per cluster in sugarcane crop from Sri Lanka. This variation may be due to the crop, varieties and the environmental conditions on which the insect was reared. The pattern of oviposition in first leaf sheaths was similar to that reported by Kaur et al. (2015) on maize. However, Singh (2013) on wheat recorded the oviposition either at the base of the plant or on soil surface or in the stubbles of rice crop. The variation describes the ability of the insect species to select the ideal oviposition substrate.

The present studies revealed the occurrence of 6 larval instars of the pest in maize. However, 8 instars have been reported by Rajendra (1976) in sugarcane and Singh (2013) in wheat and 6-8 in rice by Aggarwal et al. (2004). The number of instars of the pest varies with host plants and it increases under stress conditions (Singh 2013). The duration of different larval instars observed is similar to that reported by Aggarwal et al. (2004) on rice. Singh (2013) reported slightly longer duration and attributed this to the difference in host, season and larval instars as wheat is grown in the winter season. The length and breadth of different larval instars observed in the present study is in line with that of Aggarwal et al. (2004) in rice crop, but was more as compared to findings of Rajendra (1976) in sugarcane. In contrast to the findings, longer duration has been reported by Aggarwal et al. (2004), Rajendra (1976) and Singh (2013).

The duration of pre-pupal period is consistently reported by various workers. However, variation is observed in pupal duration. Aggarwal et al. (2004) in rice and Singh



Fig. 9. Antenna of adults of *Sesamia inferens* (a) (\times 15) and (b) (\times 10.7) Pectinate antenna of male; (c) (\times 15) and (d) (\times 10.7) Filiform antenna of female

(2013) in wheat has reported longer duration than present studies. The length and breadth of pupa reported in the present study are in agreement with that of Aggarwal et al. (2004). The weight of male and female pupa observed in the present study was comparatively less than that reported by Singh (2013). That might be due to the accumulation of more reserves in the hibernating winter population of *S. inferens*.

Adult morphometry features of *S. inferens* recorded presently and by earlier workers revealed minor variations. However, Singh (2013) reported higher average weight



Fig. 10. Life cycle of Sesamia inferens on maize

of 184.95 ± 10.17 mg for male and 241.55 ± 21.65 mg for female adults on wheat crop during winter season which might be due to higher larval weight of the hibernating population. The observations on adult longevity by earlier workers are in agreement with the present study. Reigada et al. (2016) also reported no difference in adult longevity in *Helicoverpa armigera* on different nutritionally suitable hosts. But, if the host plants are nutritionally different, the adult longevity differs significantly (Liu et al., 2004). The pre-oviposition (1.42 ± 0.05 days), oviposition (1.89 ± 0.12 days) and post oviposition period of 2.42 ± 0.10 days in rice reported by Aggarwal et al. (2004) was in tune with the present study. However, longer pre-oviposition (1.65 ± 0.21 days) and oviposition period (4.82 ± 0.54 days) was reported by Singh (2013) in wheat. The observation on adult emergence (95.8 %) and male to female sex ratio (1:1.04) is similar to Selvaraj et al. (2014) and Aggarwal et al. (2004) in rice, respectively.

The average fecundity in earlier findings on different hosts is in line with the present study except that of Singh (2013) on wheat in winter, who reported lower fecundity of 118.33 ± 12.04 eggs. Similarly, Khadioli et al. (2014) reported that the fecundity of *Sesamia calamistis* reared on artificial diet, reduced with decrease in temperature.

The duration and morphometrics of *S. inferens* in the present study is in general agreement with that of Sharma et al. (2017) on maize crop studied during October to December. However, their December to march study deviates much from the present study. The total life cycle of *S. inferens* in the present study is in harmony with Aggarwal et al. (2004) on rice crop. However, Sehkar et al. (2005) recorded 40–83 days on maize due to prolongation of larval period during unfavorable conditions and Singh (2013) reported 116.92 to 119.95 days in wheat during winter season. The present study deviates from these earlier reports primarily due to the influence of prevalent temperature on the duration of developmental stages (Viswajyothi et al., 2017). The larva also undergo diapauses at lower temperature which further increase the duration of total life cycle. Population enters diapauses at lower temperature where larva continue to grow and moult without pupation (Eizaguirre et al., 2002). Singh (2013) observed that the larval period got extended on wheat. Fantinou et al.

(1995) demonstrated diapause induction in *Sesamia nonagrioides*, resulting in longer generation time with decrease in temperature when reared on artificial diet.

The present finding will serve as a bench mark for studying bionomics, assessing damage potential and management of *S. inferens* on maize under Punjab conditions. However, the variation in the life cycle with respect to weather parameters, especially temperature should be taken into consideration.

Acknowledgements

We thank the Department of Entomology and Department of Plant Breeding and Genetics, Pandjab Agricultural university Ludhiana for providing the facilities for conducting the research successfully.

Conclusion

Sesamia inferens feeds on the economically important graminaceous crops including rice, wheat, maize and sugarcane. It is increasingly coming up in maize crop in the north western plains of India. Maize is being widely preferred by the farmers in the north western plains of the country, because of the wider adaptability and higher productivity of the crop. Thus, the increasing incidence of the pest in the region is of major concern. The biology of *S. inferens* was studied at 25 ± 1 °C and 70 ± 5 per cent relative humidity on PMH 1 maize hybrid. It took 47.65 days (male) and 49.93 days (female) to complete the development, passing through 6 larval instars, completing larval development in 29.95 ± 0.16 days. The adults had a sex ratio of 1:1.05 and fecundity of 211.92± 11.92 eggs. The observations are in general agreement with that of earlier workers who have conducted the experiments under comparable temperature conditions to the present study. Though variation is also observed with different host, temperature seems to be the major factor influencing the biology. The study on the detailed biology of the pest is necessary to develop successful integrated pest management strategies and to ensure sustained maize production in the region.

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