



Rethinking the nature of fibrolamellar bone: An integrative biological revision of sauropod plexiform bone formation

Journal:	<i>Biological Reviews</i>
Manuscript ID:	BRV-08-2012-0123.R1
Manuscript Type:	Original Article
Date Submitted by the Author:	n/a
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Keywords:	sauropod dinosaur, fibrolamellar bone, woven bone, plexiform bone, palaeohistology, osteocyte lacuna, section planes, growth rate, collagen fibre orientation

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2 **Rethinking the nature of fibrolamellar bone: An integrative biological revision of sauropod**
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4 **plexiform bone formation**
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49
50
51 Word count:

52
53 12,794 excluding title page, abstract, table of contents, references, figure captions and tables
54

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56 19,308 for the entire manuscript file
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ABSTRACT

We present novel findings on sauropod bone histology that cast doubt on general palaeohistological concepts concerning the true nature of woven bone in primary cortical bone and its role in the rapid growth and giant body sizes of sauropod dinosaurs. By preparing and investigating longitudinal thin sections of sauropod long bones, of which transverse thin sections were previously published, we found that the amount of woven bone in the primary complex has been largely overestimated. Using comparative cellular and light extinction characteristics in the two section planes, we revealed that the majority of the bony lamina consists of longitudinally organized primary bone, whereas woven bone is usually represented only by a few cells thin layer in the laminae. Previous arguments on sauropod biology, which have been based on the overestimated amount, misinterpreted formation process and misjudged role of woven bone in the plexiform bone formation of sauropod dinosaurs, are thereby confuted.

To explain the observed pattern in fossil bones, we review the most recent advances in bone biology concerning bone formation processes at cellular and tissue levels. Differentiation between static and dynamic osteogenesis (SO and DO) and the revealed characteristics of SO- vs. DO-derived bone tissues shed light on several questions raised by our palaeohistological results and permit identification of these bone tissues in fossils with high confidence. By presenting the methods generally used for investigating fossil bones, we show that the major cause of overestimation of the amount of woven bone in previous palaeohistological studies is the almost exclusive usage of transverse sections. In these sections, cells and crystallites of the longitudinally organized primary bone are cut transversally, thus cells appear rounded and crystallites remain dark under crossed plane polarizers, thereby giving the false impression of woven bone. In order to avoid further confusions in palaeohistological studies, we introduce new osteohistological terms as well as revise widely used but incorrect terminology.

1
2 To infer the role of woven bone in the bone formation of fast growing tetrapods, we review
3
4 some aspects of the interrelationships between vascularity of bone tissues, basal metabolic rate,
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6 body size and growth rate. By putting our findings into the context of osteogenesis, we provide a
7
8 new model for the diametrical limb bone growth of sauropods and present new implications for the
9
10 evolution of fast growth in vertebrates. Since biomechanical studies of bone tissues suggest that
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12 predominant collagen fibre orientation (CFO) is controlled by endogenous, functional and maybe
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14 phylogenetic factors, the relationship between CFO and bone growth rate as defined by Amprino's
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16 rule, which has been the basis for the biological interpretation of several osteohistological features,
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18 must be revised.
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22 Our findings draw attention to the urgent need for revising widely accepted basic concepts
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24 of palaeohistological studies, and for a more integrative look at bone formation, biomechanics and
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26 bone microstructural features of extant and extinct vertebrates to infer life history traits of long
27
28 extinct, iconic animals like dinosaurs.
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30

31 32 **Key words**

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34 sauropod dinosaur; fibrolamellar bone; woven bone; plexiform bone, palaeohistology; osteocyte
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36 lacuna; section planes, growth rate; collagen fibre orientation
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(1) Observed structural composition of sauropod primary bone

(2) Woven bone and HOPB in extant and fossil vertebrates

(a) Primary bone formation: SO vs. DO

(b) SO- vs. DO-derived osteocytes and lacunae

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V. ACKNOWLEDGMENTS

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I. INTRODUCTION

(1) General introduction

Dinosaurs grew at rates rivaling those of extant birds and mammals. High growth and metabolic rates have been assessed by modern paleohistological studies using fossilized bone tissues of extinct dinosaur and other archosaurian taxa. The main aim of paleohistology is to understand life histories of these extinct groups, and also to extrapolate these results into a phylogenetic context to infer evolutionary patterns of growth strategies (e.g. Chinsamy-Turan, 2005; Cubo et al. 2005, 2008, 2012; Erickson, Rogers & Yerby, 2001; Erickson, 2005; Horner, de Ricqlès & Padian, 2000; Horner, Padian & de Ricqlès 2001; Horner & Padian, 2004; Sander et al. 2004; de Ricqlès, Padian & Horner, 2003; de Ricqlès, 2008).

(2) Sauropod dinosaurs as prime example of high growth rates

Sauropod and sauropodomorph bone microstructure has been widely studied over the past decade (e.g. Sander, 2000; Sander & Tückmantel, 2003; Sander et al. 2004, 2006; Lehman & Woodward, 2008; Woodward & Lehman, 2009; Klein & Sander, 2008; Klein, Sander & Suteethorn, 2009; Stein et al. 2010; Company, 2011; Sander et al. 2011). These studies mostly report long bones with a primary cortex consisting of laminar to plexiform fibrolamellar bone. Fibrolamellar bone is defined as a composite tissue comprising a rapidly growing, woven fibred bone matrix in which osteonal lamellar bone later infills the space between woven bone and primary vascular canals (de Ricqlès, 1974). A main characteristic of woven bone (often synonymized with fibrous bone, Francillon-Vieillot et al. 1990) is the random orientation of the collagen fibrils in the matrix, the deposition of which is thought to require less time than that of a highly organized bone matrix (Amprino, 1947; Curry-Rogers & Erickson, 2005). In contrast, in the lamellar bone of the primary osteons, the collagen fibrils have a highly structured parallel spatial organization, and thus a lower depositional rate. In cross section, laminarity refers to the mainly

1
2 circumferential orientation of the vascular canals with a few radial anastomoses, which hence
3
4 divide the bony constituent into “brick-like” structures. Based on its general isotropic appearance
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6 under cross polarized light (Francillon-Vieillot et al. 1990; Castanet et al. 1993), the majority of the
7
8 laminae in transverse long bone sections of sauropod dinosaurs has been identified as woven bone.
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10 Rapid osteogenesis of fibrolamellar bone is thus inferred from the presumed random orientation of
11
12 the crystallites of the bony laminae, which reflects the original haphazard collagen fibril orientation
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14 of the living bone. It is widely accepted that this form of fast osteogenesis, probably already present
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16 in basal archosaurs (de Ricqlès et al. 2003, 2008; Cubo et al. 2012), was an exaptation that allowed
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18 sauropods to rapidly grow to enormous sizes (Curry-Rogers & Erickson, 2005; Sander et al. 2004,
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20 2011).
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24 However, our findings based on longitudinal sections of sauropod long bones question the
25
26 woven nature of the non-lamellar component of fibrolamellar bone, thereby contesting previous
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28 descriptions. These results have led the current study to review as well as revise former ideas about
29
30 fibrolamellar bone in fossil and extant vertebrates.
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33 34 35 36 37 **II. NEW RESULTS CONTESTING GENERAL CONCEPTS**

38 39 40 41 **(1) Acquiring new insight: Materials and methods**

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43 Samples of sauropod long bones (listed in Table 1.) were obtained by histological core
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45 drilling (Sander 2000; Stein & Sander, 2009). Transverse sections of these samples had been made
46
47 for previous studies and were described as laminar fibrolamellar bone (Sander & Tückmantel, 2004;
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49 Klein & Sander, 2008; Stein et al., 2010; Sander et al., 2011; Klein et al., 2012). Remaining halves
50
51 of the drill cores were cut along the long axis of the sampled long bone (perpendicular to the
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53 original sections), and thin sectioned with standard paleohistological methods (Stein & Sander
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55 2009). The sections were then investigated with linearly polarized light (LPL) microscopy with a
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1
2 Leica DM 4500 LP microscope (Leica Microsystems, Wetzlar, Germany), and compared with
3
4 transverse sections of the same drill core. Images were obtained with a Leica firecam (DFC 450)
5
6 and processed and measured with Imagic ImageAccess software. Additional specimens of two
7
8 undescribed ornithomimid theropods were sectioned and investigated to provide a broader
9
10 phylogenetic comparison. Further details of the investigated specimens are provided in Table 1.
11
12 Basic statistical evaluation of measured histological features was performed in SOFA (Statistics
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14 Open For All v.1.1.5, Paton-Simpson & Associates Ltd, Auckland, New Zealand).
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19 (2) Novel findings

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21 All of the longitudinal sections of relatively well known sauropods (*Camarasaurus*,
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23 *Apatosaurus*, *Alamosaurus* and an indeterminate diplodocid from the Morrison Fm.; detailed in
24
25 Table 1) confirm the laminar to plexiform vascular architecture previously reported for these
26
27 animals. However, strong anisotropy characterizes almost the entire primary cortical bone (Figs 1,
28
29 2). The osteocyte lacunae in this anisotropic matrix show a typical spindle shape, indicating that
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31 they were cut along their longitudinal axis (Fig. 3), and the long axis of most of them adheres to the
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33 long axis of the bone element. In transverse section, the general appearance of most of the osteocyte
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35 lacunae is rounded (sometimes referred to as 'plump') (Fig. 3). This preferential cell orientation
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37 suggests a mostly longitudinal structural organization of the primary bone (from now on referred to
38
39 as highly organized primary bone, HOPB) in all investigated specimens (Figs 2, 3). These features
40
41 were also observed in the undescribed theropod specimens.
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46 A very thin layer of bone, usually central, but often offset from the centre of the bony strut
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48 (the bony constituent of laminae *sensu* Sander [2000]) shows general light isotropical features on
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50 tissue scale in longitudinal section (Fig. 2). In the sauropod samples, this layer is one to maximum
51
52 four osteocyte lacunae wide (thickness of 20 - 40 μm), and has been described as hypermineralized
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54 'bright line' in transverse sections by other authors (Currey 1962; Francillon-Vieillot et al. 1990;
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56 Kerschnitzki et al. 2011). In this region, the osteocyte lacunae do not show any kind of preferential
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1 orientation in longitudinal nor in transverse sections. Generally, the lacunae have a random shape,
2
3 are more densely spaced and mostly appear larger than the osteocyte lacunae in HOPB. Using the
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5 definition of woven bone provided by Francillon-Vicillot et al. (1990), this 1-4 cells thin layer is the
6
7 only true woven bone in the primary cortex. Locally in this layer, two to three cells may have the
8
9 same spatial orientation. On cellular scale, the matrix surrounding these cells shows anisotropic
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11 features under cross polarized light suggesting the presence of small patches of highly organized
12
13 fibres. The average thickness of the woven bone layer ranges from 14 to 22% of the width of the
14
15 entire bony lamina and both absolute and relative thickness (Table 2) varied across taxa (ANOVA,
16
17 $p < 0.001$). The average values of absolute thickness of the woven bone layer increase with
18
19 increasing long bone shaft circumference (linear regression, Pearson $R = 0.783$; $p = 0.007$), but
20
21 there is no correlation between the average fraction of woven bone and shaft circumference (linear
22
23 regression, Pearson $R = -0.128$; $p = 0.725$). The correlation between shaft circumference and
24
25 average thickness of the bony lamina (Pearson $R = 0.642$) was statistically not significant (linear
26
27 regression, $p = 0.045$). These results suggest that the proportional thickness of the woven bone layer
28
29 is variable within the bony lamina across different taxa. For further details on statistical output,
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31 please see Table 2 and 3.
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37 The highly organized primary bone (HOPB), which is characterized by the long range order
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39 of matrix and cell lacuna orientation, may consist of lamellated and non-lamellated structural
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41 subunits in the investigated specimens (Fig. 4). Lamellation is easily detectable in transverse and
42
43 longitudinal sections if the successive lamellae appear alternating dark and bright under cross
44
45 polarized light. Such a pattern of alternating dark and bright bands adjacent to the vascular space
46
47 was observed in *Apatosaurus* (two in femur SMA 'Chris'; up to five in humerus CM3378, Fig. 4A).
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49 In these specimens, usually the lamellation starts with a dark lamella in longitudinal section, and the
50
51 same lamella will appear bright in transverse section, hence exhibiting the complementary pattern
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53 of interference. Wherever osteocyte lacunae are embedded in the lamellae, the appearance of them
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55 in the dark bands is generally rounded, whereas in the bright bands they have an elongated spindle
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1 shape in either section plane. These features imply a plywood structure with changing fibre
2 orientation in successive lamellae in the living bone (Francillon-Viellot et al. 1990 and references
3 therein). Fine lines visible under plane polarized light are mostly associated with these lamellated
4 perivascular regions (Fig. 4B). However, the borders of the individual lamellae defined by their
5 extinction pattern do not always match these lines. Vascular canals surrounded by such lamellated
6 areas have been referred to as primary osteons by other authors (e.g. de Ricqlès 1968; Currey, 2002;
7 Chinsamy-Turan, 2005). If adjoining lamellae have a uniform extinction pattern rather than an
8 alternating dark and bright appearance, distinguishing them may be more difficult. This is the case
9 in *Alamosaurus*, where 7 to 10 thin lamellar bands are detectable under single plane polarizer based
10 on the presence of fine lines, but these appear uniformly dark under cross polarized light in
11 transverse section and uniformly bright in longitudinal section (Fig. 4B). In transverse section, these
12 dark lamellae are sometimes lined with one bright lamella positioned on the internal surface of the
13 vascular space. The dark lamellar bands have longitudinally oriented osteocyte lacunae (rounded in
14 transverse section; strongly elongated in longitudinal section), whereas the long axes of the
15 osteocyte lacunae in the bright lamellar band are oriented in the transverse plane.

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35 In large areas of the highly organized primary bone, no trace of lamellation can be revealed
36 under single or crossed plane polarizers. This non-lamellated HOPB is usually sandwiched between
37 the thin woven bone layer and the lamellae bordering the vascular space (e.g. *Apatosaurus*,
38 *Camarasaurus*), but it may also form the only structural subunit of the non-woven primary bone
39 (e.g. diplodocid indet. NHUB Ki2, Fig. 4C) if no lamellae had been deposited at the time of death.
40 The transition from the woven bone layer to this non-lamellated HOPB is marked by the appearance
41 of osteocyte lacunae with gradually more longitudinal orientation (parallel to the longitudinal axis
42 of bone), but no particular growth marks or any other distinct features indicating an abrupt change
43 in the deposited tissue type are present.

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55 The relative amount of lamellated and non-lamellated components in the HOPB is variable
56 in different specimens, even though the exact border between the two subunits is usually hard to
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1
2 define (Figs 4, 5). This ratio does not only differ in different areas of the same section and between
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4 different ontogenetic stages of the same species but may vary among species representing similar
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6 histologic ontogenetic stages (HOS). For example, in most areas of the section of *Apatosaurus*
7
8 CM3378 (HOS 11), the ratio of lamellated and non-lamellated HOPB is approximately 1/1, whereas
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10 in *Alamosaurus* TMM46002 (HOS 7), the lamellae are frequently deposited directly on the woven
11
12 bone, thus the highly organized primary bone consists solely of lamellae in these regions. In
13
14 “*Barosaurus*” Ki2 (HOS 10.5), the lamellated tissue is usually completely absent. Figure 5 shows
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16 further details on the average proportional distribution of different constituents of the laminae
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18 (*sensu* Sander, 2000) in the investigated specimens.
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26 III. REVIEW AND REVISION PROMOTED BY THE NEW RESULTS

27 (1) Observed structural composition of sauropod primary bone

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32 Strong polarized light anisotropy in longitudinal sections of sauropod long bone cortices
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34 indicates a mostly longitudinal crystallite orientation in the non-lamellar primary bone as well (Fig.
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36 1). These crystallite orientations correspond to the original crystallite and matrix orientation in the
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38 bone tissues of the living animals (Kolodny et al. 1996; Hubert et al. 1996; Trueman & Tuross,
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40 2002). Truly woven bone, that appears generally darker in longitudinal sections too, is represented
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42 only by a thin layer centrally positioned in the bony laminae (Fig. 2). This thin layer corresponds to
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44 the highly mineralized ‘bright line’ (Francillon-Viellet et al. 1990), and is the oldest part of the
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46 bony lamina. The remainder of the non-lamellar primary bone is highly organized with a
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48 longitudinal preferential organization. This extensive longitudinal structural arrangement in the
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50 non-lamellar HOPB is also reflected by the long range spatial order of the osteocyte lacunae being
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52 uniformly cut along their long axis in longitudinal sections. These results are inconsistent with
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54 widely reported claims that the bulk of the non-lamellar primary bone in sauropod plexiform bone
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1 (often synonymized with laminar fibrolamellar bone, see also de Ricqlès 1968a; 1974) consists
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3 entirely of woven bone with originally randomly oriented collagen fibrils and apatite crystallites. In
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5 contrast, our results indicate that the microstructural arrangement of sauropod long bones
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7 corresponds to the strong longitudinal organization also observed in ovine and bovine plexiform
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9 (fibrolamellar) primary bone (Kerschnitzki et al. 2011). Kerschnitzki and coauthors (2011) have
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11 shown that the amount of woven bone in ovine and bovine plexiform bone is limited to a layer only
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13 a few cells thin, similar to our findings in sauropod primary bone.
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17 In the sauropod samples presented here, highly organized primary bone may appear in the
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19 form of lamellated and/or non-lamellated subunits, both of which show a long range uniform
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21 extinction pattern and parallel orientation of the osteocyte lacunae. The histological distinction of
22
23 these two HOPB subunits is often ambiguous for several reasons. First of all, the true nature of
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25 lamellation is still under debate. Lamellated appearance has been interpreted as the result of
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27 changing fibre orientations (Gebhardt, 1906; Giraud-Guille, 1988) or changing fibre and mineral
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29 densities (Boyde & Hordell, 1969; Marotti, 1993; Marotti, Palazzini & Palumbo, 1993) in
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31 subsequent lamellae both of which eventuate in an alternating extinction pattern under cross
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33 polarized light. Lamellation has also been defined by the presence of interlamellar zones that
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35 separate the successive lamellae, consist only of less organized collagen fibres and appear as darker
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37 stripes between the individual lamellae under crossed plane polarizers (Ascenzi & Bonucci, 1968;
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39 Bromage et al, 2009). We do not support the exclusive validity of the first definition of lamellation,
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41 because we could still detect lamellation based on the perivascular fine lines visible under plane
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43 polarized light in specimens with uniform fibre orientation in successive lamellae (e.g.
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45 *Alamosaurus*, Fig. 4B). However, the structure of these fine lines is unclear, because they do not
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47 always correspond to the border of lamellae with alternating extinction pattern or to the dark stripes
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49 that might represent interlamellar zones either. We hypothesize that the fine lines represent
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51 temporal interruptions in matrix deposition that might be followed by synchronized spatial
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53 reorganization of the osteoblasts and thus uniform change in fibre orientation. In this concept, all
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1
2 previously proposed definitions of lamellation may be valid representing a considerable variability
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4 in the formation, structure and composition of lamellated bone. Furthermore, the thickness of the
5
6 lamellae can vary considerably (Bromage et al, 2009; 2011), thus local changes in fibre orientation
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8 in the non-lamellated HOPB may appear as lamellae. Finally, the border between the first lamella
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10 and the non-lamellated HOPB is not always distinct which is probably due to the same principles of
11
12 formation of the two subunits on a cellular level. Hence, we prefer to rely on the combination of the
13
14 presence of fine lines and extinction pattern when defining HOPB lamellar patterns. Even though
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16 their image of plexiform bovine bone shows lamellated and non-lamellated HOPB, Kerschnitzki et
17
18 al. (2011) did not differentiate these two structural appearances but rather referred to all HOPB
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20 deposited on the woven bone layer as lamellar bone. Similarly, other authors consider any type of
21
22 primary bone as lamellar if it reveals high degree of parallel fibre and crystallite orientation. We
23
24 propose a consistent usage of lamellar bone for HOPB tissues which show clear characteristics of
25
26 lamellation. Since HOPB, be it lamellated or non-lamellated, always exhibits high degree of parallel
27
28 organization, the term 'parallel-fibred bone' is more adequate to describe both types of HOPB
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30 rather than an intermediate degree of structural order as it was suggested by other authors (e.g.
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32 Francillon-Viellet et al. 1990; Currey, 2002). Thus, HOPB should be synonymized with primary
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34 parallel-fibred bone. However, with the unification of the two forms of appearance (lamellar and
35
36 non-lamellar) of parallel-fibred bone by introducing the new term HOPB, we can avoid confusion
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38 originating from the inconsistent or even incorrect usage of the term parallel-fibred bone.
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43 Furthermore, based on the common formation principles (see below) and because of the difficulties
44
45 related to the distinction of lamellated and non-lamellated HOPB, we suggest that any form of
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47 HOPB deposited on the initial woven bone layer be considered as the bony constituent of primary
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49 osteons (cf. Ferretti et al. 2002).
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55 **(2) Woven bone and HOPB in extant and fossil vertebrates**

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2 Our results clearly indicate that the amount of woven bone in the long bones of sauropods
3 and probably a number of other dinosaurs has been largely overestimated. In a classical transverse
4 section under crossed plane polarizers, the dark part of the lamina that was previously referred to as
5 woven bone is up to three to four times thicker than the actual layer of woven bone (Fig. 2). This
6 overestimation has led to a number of conclusions and generally accepted interpretations that, in the
7 light of the current observations, call for urgent revision and modification. To overcome confusion
8 and avoid further misconceptions when working with fossil bones, the true nature of woven bone
9 and HOPB needs to be considered on cellular level. Understanding the formation process of these
10 bone tissues is the key to understanding their microstructure, cellular composition, spatial
11 arrangement, growth, and developmental role. This background knowledge is essential to draw any
12 biological inferences based on the presence of woven bone and HOPB in extinct animals.
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28 *(a) Primary bone formation: SO vs. DO*

29 Recent histological studies of extant vertebrates revealed two major types of primary bone
30 formation: stationary and dynamic osteogenesis (Marotti et al. 1999; Ferretti et al. 2002; Palumbo,
31 Ferretti & Marotti, 2004; Marotti, 2010).
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37 In stationary or static osteogenesis (SO), the ossification center is composed of highly
38 vascularized mesenchymal tissue in which plump cells of diverse shapes start to differentiate into
39 osteoblasts half way between two adjacent primary vascular spaces. In this layer, the thickness of
40 which does not exceed 2-3 cells, the osteoblasts are randomly arranged with respect to each other;
41 however, each of them is functionally polarized with definite secretory territory (Ferretti et al. 2002;
42 Palumbo et al. 2004; Marotti, 2010). The secretory territory is a membrane surface area
43 characterized by finger-like processes at the base of which collagen fibrils secreted in exocytotic
44 vesicles assemble to form extracellular fibril bundles (Palumbo et al. 1990; Pazzaglia et al. 2010).
45
46 As these osteoblasts start to secrete the proosseus matrix, they do not form a movable sheet of cells
47 such as a monostratified osteogenic lamina, but stay at the same place in this spatially unorganized
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2 manner “trapped” in the matrix they produce. Because their orientation is random with respect to
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4 the adjacent osteoblasts, their secretory territories are differently directed resulting in a highly
5
6 porous matrix. Nevertheless, fibres still form highly organized bundles around the individual cells
7
8 that secrete them. This short range (cellular scale) order of fibres has been referred to as
9
10 ‘microlamellar bone’ by Kerschnitzki et al. (2011). The osteoblasts soon begin to differentiate into
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12 large, globous osteocytes *in situ* forming the initial bony trabeculae that are positioned at constant
13
14 distance from the adjacent primary vascular spaces. Thus, the term stationary or static osteogenesis
15
16 refers to the immobile nature of these osteoblasts and consequently to their transformation into
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18 osteocytes at the same place where they differentiated from mesenchymal stem cells. Shortly after
19
20 osteocyte transformation, the preosseous matrix undergoes considerable mineralization. Some of the
21
22 osteocytes are enclosed in groups within the same large, confluent lacunae (Ferretti et al. 2002;
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24 Palumbo et al. 2004; Marotti, 2010). This porous, irregular spatial construction is probably
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26 responsible for the fast volume increase i.e. high formation rate of this *de novo* bone tissue. As SO
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28 proceeds, it encloses the capillary network, thereby significantly expanding the volume of the
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30 forming bone in a relatively short amount of time. Thus, SO results in a connective tissue type that
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32 clearly corresponds on each hierarchical level to what has been defined as woven bone. This
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34 equivalence has also been pointed out by Palumbo et al. (2004) and Marotti (2010).
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40 In contrast to static osteogenesis, dynamic osteogenesis (DO) involves a movable set of
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42 osteoblasts organized in a monolayer osteogenic lamina (Ferretti et al. 2002; Palumbo et al. 2004;
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44 Marotti, 2010). Within this lamina, the osteoblasts are all polarized in the same direction and
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46 produce matrix in a highly organized manner (Marotti, 2010; Pazzaglia et al. 2010). The
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48 designation ‘moveable’ implies that the lamina adheres to the preosseous matrix it secretes,
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50 continuously thickening the deposited layer from which the lamina seems to “move away”. In a
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52 regular pattern, those osteoblasts that are destined for differentiating into osteocytes, stop producing
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54 matrix, whereby the neighboring, matrix-secreting osteoblasts bury them one by one into the
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56 deposited layers of bone (Marotti et al. 1992; Franz-Odenaal, Hall & Eckhard Witten, 2006).
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2 However, the formation of an osteogenic lamina with synchronized secretion activity requires the
3 presence of an initial substrate, on the surface of which the lamina can differentiate (Marotti, 2010;
4 Kerschnitzki et al. 2011). This substrate can be provided by the cord of woven bone trabeculae
5 formed *de novo* by SO or on the surface of cartilage precursors in endochondral ossification
6 (Marotti, 2010). Thus, DO corresponds to accretional or appositional bone growth and always
7 results in the formation of structurally highly organized bone tissue with preferential fibre and cell
8 orientation. DO is responsible for the thickening of the 2-3 cells thin SO-trabeculae, and thereby for
9 the compaction of bone through infilling of the primary vascular spaces using SO-trabeculae as the
10 primer substrate.
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21 On the basis of the above review on primary bone formation, it becomes evident that *de*
22 *novo* i.e. woven bone is formed by static osteogenesis, whereas HOPB of parallel structural
23 arrangement is the product of dynamic osteogenesis. Although the well-known classification of
24 intramembranous ossification (e.g. formation of most cranial bones) *versus* endochondral
25 ossification (e.g. formation of long bones) is also defined on tissue level, it is clear that bone
26 formation in both involves the combination of static and dynamic osteogenesis. This has significant
27 bearings on the evolutionary considerations of different bone tissue types (see below).
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44 **(b)** *SO- vs. DO-derived osteocytes and lacunae*

45 The morphology of the osteocytes also helps identifying whether the investigated primary
46 bone tissue is derived from SO or DO, even though there is no structural or ultrastructural
47 difference between immobile and mobile osteoblasts (Marotti, 2010).
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50 Osteocytes originating from SO mostly retain features of their incipient osteoblast-
51 morphology. They are relatively large, globous (plump), and have short dendritic processes which
52 radiate symmetrically all around the cell body and run in the canaliculi to connect to the adjacent
53 osteocytes by means of gap junctions (Palumbo et al. 2004). Since some of these osteocytes share
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2 the same confluent lacunae, generally the lacunae are also larger in size and exhibit more irregular
3 shapes (Ferretti et al. 2002). No mutual alignment of SO-cells and their lacunae and canalicular
4 system can be observed.
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8 DO-derived osteocytes, on the other hand, are smaller, more elongate and flattened with
9 more and longer cytoplasmic processes which run mainly perpendicular to the long axis of the cell.
10 The long axis of DO-osteocytes is perpendicular to the thickness of the osteoid deposited in their
11 osteoblast-stage (Currey, 2003; Franz-Odenaal et al. 2006). Due to the uniform morphology of
12 these osteocytes, their lacunae and canaliculi follow this pattern exhibiting dense canaliculi that are
13 oriented perpendicularly to the long axis of the mostly spindle-shaped lacunae (Kerschnitzki et al.
14 2011). The DO-osteocytes show a long-range uniform orientation in the deposited HOPB.
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26 *(c) Occurrence, developmental sequence and role of SO- and DO-derived primary bone*
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28 The major differences in the formation principal of SO and DO-derived primary bone
29 determine their occurrence, role and timing in skeletal development. SO is needed wherever there is
30 no initial substrate surface present upon which osteoblasts could orderly assemble (Sugawara et al.
31 2005; Mori et al. 2007; Shapiro, 2008; Kerschnitzki et al. 2011). Thus, *de novo* bone formation
32 occurs in the intramembranous ossification centers in which mesenchymal cells condensate in a 2-3
33 cells thick layer to form the woven bony strut between primary vascular spaces. The formation
34 principals of this process do not differ from the woven bone formation during the diametrical
35 growth of the ovine and bovine long bones cortices (Kerschnitzki et al. 2011). Thus, certain regions
36 of the bone collar, which is the densely vascularized perichondrial mesenchymal tissue condensated
37 around the growing long bone shaft (Karaplis, 2008), can be interpreted as potential local
38 intramembranous ossification centers. However, based on the demonstrated differences in their
39 genetic regulatory systems, intramembranous ossification and bone collar formation have been
40 suggested to be developmentally distinct (Karaplis, 2008). Nevertheless, in the perivascular
41 ossification centers of the bone collar, SO most probably provides the fast volume expansion of the
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2 growing bone through the enclosure of extensive primary cavities (Ferretti et al. 2002). SO has also
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4 been reported in the first phase of bone fracture repair producing woven bone in the callus (Marotti,
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6 2010). Since SO produces the scaffold for further bone deposition, woven bone along with cartilage
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8 (chondrification into the fracture gap, cf. Pritchard & Ruzicka, 1950) is needed in fracture repair
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10 probably because the fracture surface is an ill-defined osteogenic surface with no directional
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12 information for organized bone deposition. Alveolar bone also forms *de novo* separated from the
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14 jaw bones in embryonic stages (Osborn, 1984; Smith & Hall, 1993; Smith & Coates, 2000) and
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16 woven bone formation has also been reported adjacent to alveolar endosseous implants (e.g.
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18 Berglundh et al. 2003). In some cases, SO can be induced during endochondral ossification where
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20 hypertrophied chondrocytes are resorbed or transdifferentiate into woven bone producing
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22 osteoblasts (Franz-Odenaal et al. 2006 and references therein). In each case, SO provides the
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24 scaffold for the subsequent deposition of DO-derived, hence more organized primary bone tissue
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26 (Ferretti et al. 2002; Marotti, 2010; Kerschnitzki et al. 2011; Mori et al. 2007; Shapiro 2008;
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28 Sugawara et al. 2005). As bone matures, SO is always followed by DO, consequently the presence
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30 of HOPB is always expected around the woven bone framework in later ontogenetic stages.
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35 DO is a substrate-dependent process, and the function of the osteogenic lamina is
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37 characterized by the accretional deposition of highly structured primary bone (Ferretti et al. 2002;
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39 Marotti, 2010). The initial substrate can vary, therefore DO can occur wherever accretional bone
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41 growth thickens the cortex or compacts the primary cavities. The diametrical accretional thickening
42
43 occurs in perichondral, periosteal and endosteal bone growth on the surface of cartilage precursor or
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45 on already existing bony surface. Cavity compaction can start on the framework consisting of SO-
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47 derived woven bone (Marotti, 2010) or on the surface of the erosion cavities or cutting cones in
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49 secondary remodeling resulting in secondary osteons (Pazzaglia et al. 2011). Thus, in contrast to
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51 SO, DO deposits primary as well as secondary bone and generally reflects more mature ontogenetic
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Since the end product of SO, i.e. woven bone is a poor quality, porous and brittle bone with low resistance against various mechanical loading (Marotti 2010), we suggest that this is very likely a temporarily as well as spatially limited mode of bone formation which is evidenced by the proportionally low amount of woven bone in larger, fast growing animals. In concurrence with this assumption, SO is believed to be induced by inductive stimuli such as different growth factors, whereas DO is triggered by mechanical stimuli transmitted through the osteocytes of the mechanically less resistant SO-trabeculae (Marotti, 2010). This draws attention to the predominantly biomechanical significance of the highly organized fibre arrangement in the bone, as concluded by several previous studies investigating the relation between mechanical loading and long range fibre orientation in various skeletal elements (Ascenzi & Bonucci, 1967, 1968; Boyde & Riggs, 1990; Ascenzi et al. 1987; Martin & Ishida, 1989; Bromage, 2003; Skedros & Hunt, 2004, Skedros et al. 1996, 2006, 2007).

(d) Distinguishing SO & DO characteristics in fossil bone

After demonstrating SO and DO in extant animals, identifying SO- and DO-derived tissues in fossil bones becomes possible with much higher confidence than before. This has important bearings on functional and evolutionary interpretations.

Our notion that only a centrally positioned, few cells thin cord represents woven bone in the bony laminae of sauropod plexiform bone (Fig. 2) is supported by its SO-derived characteristics. One of these characteristics is its isotropic nature on tissue level, but its local birefringence on cellular level under cross polarized light (in accordance with the ‘microlamellar’ concept of woven bone, Kerschnitzki et al. 2011, see below). Another SO-derived feature is reflected in the irregular arrangement and morphology of the cell lacunae. In addition, osteocyte lacunae are more densely spaced in the woven bone cord than in HOPB. The absolute thickness of the woven bone trabeculae ranging from 2 to 4 cells in sauropods and other investigated dinosaurs (theropods) also fits the range of that found in the same structure of ovine and bovine bone (Kerschnitzki et al. 2011). These

1 features of SO-derived trabeculae are in sharp contrast with the rest of the non-lamellar part of the
2 bony laminae that clearly shows anisotropic features and elongate osteocytes with uniform
3 longitudinal alignment in longitudinal sections (Fig. 2). The lamellated part also reveals high
4 structural organization with or without alternating extinction pattern and osteocyte orientation in
5 subsequent lamellae (Fig. 4). This clearly implies that the bulk of the bony lamina in sauropod long
6 bones consists of DO-derived primary parallel-fibred bone (synonymized with HOPB) similarly to
7 the primary cortex of ovine and bovine long bones (Ascenzi et al. 1967; Martin & Ishida, 1989;
8 Kerschnitzki et al. 2011). The majority of HOPB exhibits structural alignment along the
9 longitudinal axis of the bone with some lamellae showing transverse organization.

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22 Fibre orientation in SO- and DO-derived fossil bones cannot be observed directly, however,
23 it can be inferred with high confidence. Until now, the optical behaviour of the thin section under
24 cross polarized light was the most frequently used method in determining original fibre orientation
25 in fossil bones. This method is based on the assumption that the apatite crystallites are aligned along
26 the longitudinal axis of the anisotropic collagen fibres in the living bone, and will faithfully show
27 the same arrangement after fossilization (Kolodny et al. 1996; Hubert et al. 1996; Trueman &
28 Tuross 2002). The anisotropy of fibres and/or crystallites is expressed by a direction-dependent
29 variation in brightness under cross polarized light. Depending on how the plane of section relates to
30 the longitudinal axis of the fibres or apatite crystallite rows, they will show different extinction
31 pattern. If the plane of cut is parallel with the long axis of fibres/crystallites, they will exhibit
32 maximum brightness, whereas if the fibres/crystallites are cut transversally, they will appear
33 completely dark under crossed plane polarizers. Any other direction of the section plane relatively
34 to the long axis of the fibres/crystallites will result in intermediate brightness values (Ascenzi &
35 Bonnucci, 1968; Bromage et al. 2003). The use of full-wave (λ) or half-wave ($\lambda/2$) plates may
36 improve the quality of the image by introducing a controlled phase shift between the two
37 polarization components of a light wave, altering its polarization and resulting in different color
38 patterns which can help identifying tissues. Quarter-wave plates ($\lambda/4$) convert linearly polarized
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2 light (LPL) to circularly polarized light (CPL) thereby eradicating the obscuring effect of extinction
3 crosses. Guidelines and methodological considerations for the use of CPL are provided by Bomage
4 et al. (2003). If standardized thickness of the thin sections ($100 \pm 5 \mu\text{m}$) is achieved, with the help
5 of a circumpolarized light and grey-level analysis, finer-scale quantification of fibre orientation
6 becomes possible (Bromage et al., 2003, 2009, 2011). However, the efficiency of this method might
7 be interfered by diagenetic extinction-alterations which usually determine how thick the section of
8 fossil bones must be to get the desired resolution of the microstructure. Hence, it is clear that there
9 is always a need for at least two section planes to reconstruct the original three-dimensional
10 structure based on the preserved optical features in a fossil bone. Nevertheless, important aspects of
11 this optical-directional coherence have largely been overlooked by paleohistological investigations
12 which mainly or exclusively relied on transverse sections when drawing inferences on fibre
13 orientation in fossil specimens. This is the most important factor that has led to mistaking
14 longitudinally oriented HOPB for woven bone in sauropods, and most probably in a number of
15 other dinosaurs too (longitudinally structured HOPB will appear completely dark and isotropic in
16 transverse section on the basis of which it has been interpreted as woven bone, see below).
17 However, even if the bone sample is cut in multiple planes, relying only on optical features revealed
18 using either LPL or CPL techniques may be misleading, since diagenetic alterations may after all
19 have modified the original extinction pattern of the fossil bone tissue. Based on the findings of
20 Kerschnitzki et al. (2011), lacunocanalicular features of osteocytes help decrease the uncertainties
21 related to determining original fibre orientation in a fossil bone. Kerschnitzki et al. (2011)
22 visualized the osteocytic network in ovine, bovine and murine primary bone using staining and
23 different microscopic methods. They found a strong organization of the lacunocanalicular network
24 (LCN) in DO-derived, thus parallel-fibred primary bone with the long axis of the lacunae aligned
25 parallel to the predominant orientation of collagen fibres, and the canaliculi running perpendicular
26 to the fibres as well as to the long axis of the lacunae. The same correlation between the LCN and
27 fibre orientation exists in secondary bone, but there is no regular pattern of LCN in the SO-derived
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1 woven bone (Kerschnitzki et al. 2011). This finding is in accordance with the irregular spatial
2 arrangement and thus randomly oriented interconnections of the immobile SO-osteoblasts. Hence,
3 the structural organization of LCN is also highly indicative of large scale collagen fibre
4 arrangement, and thereby SO- vs. DO-derived bone tissues (i.e. woven vs. parallel-fibred bone) can
5 clearly be distinguished in fossil tissues as well.
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12 Because the spatial organization of collagen fibril bundles produced by the individual SO-
13 osteoblasts does not exceed the canalicular radius of a few cells, Kerschnitzki et al. (2011) referred
14 to woven bone as 'microlamellar bone', indicating the high organization of the collagenous matrix
15 on a cellular level. However, since we restrict the term 'lamellar' to structures described in the
16 above sections, we do not adopt this terminology for woven bone. On the other hand, it follows that
17 at cellular level, woven bone may not appear completely dark and isotropic but locally can also
18 reveal light extinction under crossed plane polarizers (also evident in murine woven bone,
19 Kerschnitzki et al. 2011). Indeed, in some cases we found more expressed extinction in the woven
20 trabeculae than in the HOPB which remained all dark in the transverse sections of sauropod long
21 bones. Such a pattern is to be expected if HOPB is longitudinally organized and the
22 fibres/crystallites are cut perpendicular to their longitudinal axis. This strengthens the identification
23 and structural interpretation of woven bone and HOPB in the sauropod long bones investigated in
24 the current study (Fig. 2).
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41 In the light of our results it is evident that future paleohistological studies will have to
42 consider the combination of optical features and lacunocanalicular morphology in multiple section
43 planes to draw inferences on the original micro- and ultrastructure of fossil bones. Figure 6
44 demonstrates a revised general model of the macro- and microstructure (A-C) and optical behavior
45 (D-F) of the plexiform bone of sauropods representing the new interpretations of this study. The
46 model in Figure 6 was based on the investigated specimen *Apatosaurus* CM 3378 and thereby
47 represents only one of the various possible structural organizations of sauropod primary bone. In
48 order to sum up the effects of the plane of sectioning on the appearance of histological features, this
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2 model indicates the cellular (B,C) and optical appearance (E,F) of the thin section in two section
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8 9 **(3) Causes of misinterpretations and erroneous reasoning in paleohistological studies**

10 The term 'fibrolamellar' was devised to describe the composite nature of this bone tissue
11 type consisting of 'fibrous' and lamellar bone. 'Fibrous bone' is the direct translation of
12 'Faserknochen', a term that had been applied by Gross (1934) to all types of periosteally deposited
13 'non-osteonal' primary bone, which he therefore synonymized with 'Periostknochen'. Thus, Gross'
14 category 'Faserknochen' did not only refer to true woven bone with randomly distributed cells in
15 the unorganized matrix (which he termed 'restlicher Faserknochen'), but also to the structurally
16 organized non-lamellar primary bone (which he called 'zonarer Faser- oder Periostknochen'), and
17 even to the acellular periosteal bone (Gross, 1934). With this term, he aimed to express that 'fibrous
18 bone' mostly shows clear signs of 'being fibred' ("Der Faser- oder Periostknochen...ist meist sehr
19 deutlich gefasert."); a feature that is much less apparent in woven bone than in primary bone with
20 highly organized fibre arrangement. In spite of that, subsequent authors have synonymized fibrous
21 bone with woven bone (e.g. de Ricqlès, 1974; Francillon-Viellet et al. 1990), and in the
22 'fibrolamellar' concept it has been systematically used this way ever since. Possibly Gross' (1934)
23 description of 'Faserknochen' was misinterpreted and mistranslated from German, resulting in a
24 suite of confusing bone histological terminology. Fibrous bone (*sensu* Gross, 1934) and woven
25 bone are thus not the same. Because the formation principals of the SO-derived woven bone and the
26 DO-derived non-lamellar HOPB are so different, we do not support the unification of these
27 structures under the common term 'fibrous bone' as Gross (1934) suggested. Moreover, infilling of
28 the primary vascular spaces does not necessarily involve formation of lamellae (e.g. diplodocid
29 indet. NHUB Ki2). Thus, the term 'fibrolamellar bone' reflects neither the developmental origin of
30 the different bony constituents nor the observed diversity in the microstructures of sauropod
31 primary bone, and therefore should be abandoned. Instead, here we suggest that fast growing, thus
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1 highly vascularized composite bone tissue types that consist of the combination of woven bone and
2 HOPB, be categorized only based on their vascular organization. Hence, in the case of sauropod
3 long bones, primary bone tissue should simply be referred to as laminar to plexiform primary bone.
4 This terminology is also consistent with the new concept of DO-defined primary osteons.
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10 Another misleading conception originates from the superficial observation of the optical
11 behavior of woven bone. Due to the well-known haphazard orientation of the fibres and crystallites
12 in woven bone (Chinsamy-Turan, 2005; Currey, 2002), it has generally been claimed to remain dark
13 under crossed plane polarizers, which is in sharp contrast to the alternating bright and dark patterns
14 exhibited by the surrounding lamellar bone when the stage is rotated (e.g. Francillon-Vieillot et al.
15 1990). This statement became so dogmatic in paleohistology that almost any kind of primary matrix
16 type that stays dark under crossed plane polarizers has been referred to as woven bone. However, it
17 is clear that those areas of the matrix which steadily appear the darkest in the sections are always
18 the ones that contain fibres/crystallites which are cut exactly transversally (Ascenzi & Bonnucci,
19 1968; Bromage et al. 2003, 2009). Hence, the darkest matrix areas that are rather extensive
20 (measurable on millimeter scale) certainly have a long range order of collagen and/or crystallite
21 organization, i.e. they show a uniform preferred orientation and run perpendicular to the plane of
22 sectioning. In contrast, woven bone does show extinction due to the presence of locally organized
23 fibre bundles at the cellular level (Kerschnitzki et al. 2011). It follows from this that woven bone
24 will always have higher brightness values than the transversally cut HOPB under crossed plane
25 polarizers, even though this brighter appearance is the result of a random composite of darker and
26 brighter areas. The vulgar error that woven bone is completely dark under crossed plane polarizers
27 combined with the almost exclusive usage of transverse sections in paleohistological studies have
28 likely led to mistaking longitudinally organized (thus transversally cut) primary bone for woven
29 bone in many cases.
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54 The effect of the plane of sectioning on the appearance of the lacunocanalicular system (Fig.
55 3) presented further source of misinterpretation. A number of studies have used aspects of shape,
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1 size, orientation and density of the lacunocanalicular system to infer large scale evolutionary
2 patterns without taking bone three-dimensional structure into account. Rensberger & Watabe (2000)
3 claimed that differences in the angle of the canaliculi sprouting from the osteocyte lacuna
4 represented differential rates of osteogenesis. According to these authors, there is a random
5 orientation of canaliculi on the lacunar surface in the long bones of non-avian theropod dinosaurs
6 and birds, which is in contrast with the strong radial orientation found in ornithischian dinosaurs
7 and mammals. Rensberger & Watabe (2000) interpreted the random canalicular orientation
8 indicative of a lower degree of structural organization (i.e. woven bone), implying higher bone
9 growth rates in non-avian theropods and birds than in ornithischians and mammals with a more
10 organized LCN. Here we want to point out that the random and radial canalicular patterns described
11 by Rensberger & Watabe (2000) at the level of the osteon are actually radial and perpendicular to
12 the long axis of the lacuna respectively on a cellular scale. As prime evidence for differences in the
13 degree of structural organization, the authors provided images (Rensberger & Watabe, 2000, figure
14 2) of the LCN in secondary osteons. However, all secondary bone is DO-derived, and hence
15 characterized by a strongly organized nature. This disputes the irregular structural organization
16 hypothesized by Rensberger & Watabe (2000), and rather points to local differences in cell
17 orientation in the highly organized matrix. Depending on the plane of sectioning of the cell lacuna,
18 transverse or longitudinal, any DO-derived lacuna will exhibit a rounded or flattened shape with
19 radial or perpendicular canalicular pattern respectively. Thus, we hypothesize that Rensberger &
20 Watabe (2000) most likely documented different patterns of loading in the limb bones they studied
21 rather than real differences in the degree of structural organization (see biomechanical
22 considerations below). Morphological distinction between “flattened” and “stellate” osteocyte
23 morphologies based on ground sections of turtle shell bones described by Cadena & Schweitzer
24 (2012) is loaded with the same uncertainties. For instance, the alternating layers of “flattened” and
25 “stellate” osteocytes shown in the internal cortex of the costal bone of *Podocnemis expansa*
26 demonstrates a plywood like arrangement of longitudinally vs. transversally orientated osteocytes
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1 of HOPB rather than osteocytes of genuinely different 3D shapes. Flattened and stellate
2 morphotypes identified among isolated 3D osteocytes of extant species and the organic
3 lacunocanicular infilling of the fossil species most probably represent DO- vs. SO-derived
4 osteocytes, respectively (note that 'SO' in Cadena & Schweitzer, [2012] does not refer to static
5 osteogenesis but to stellate osteocytes). For the same reasons, studies that use osteocyte lacuna
6 volume are prone to similar mistakes. Lacuna volume has been used in paleogenomic studies to
7 reconstruct evolutionary changes in the genome size of birds compared to other large tetrapod
8 clades (Organ et al., 2007; Organ, Brusatte & Stein, 2010). Lacuna volumes in these studies were
9 calculated with measurements of lacuna long and short axis taken in classical transverse sections.
10 Because of uncertainties related to the plane of sectioning at the cellular level, at least in some
11 species with longitudinally organized bone, the longitudinal axes of the lacunae must have been
12 significantly underestimated. Montanari et al. (2011) provided further criticism by documenting
13 strong differences in estimated genome size of a single individual, depending on which skeletal
14 element was studied. Pending further investigation, we hypothesize that differences in estimated
15 genome size may lie in differences in the plane of sectioning of the lacunae.

16 Studies focusing on sauropod bone histology suffer from the same drawbacks (cf. Organ et
17 al., 2009). Transversally cut osteocyte lacunae of the longitudinal HOPB appear round or oval in
18 cross section, thus they have often been identified as the typical rounded, "plump" lacunae of
19 woven bone. Since a considerable proportion of the primary plexiform bone of sauropods is
20 composed of longitudinally organized bone, most of the non-lamellated bony laminae show up dark
21 in transverse sections and as such have been referred to as woven bone. These observations and the
22 erroneous conclusions have resulted in a considerable overestimation of the actual amount of woven
23 bone. De Ricqlès (1968a,b, 1974, 1975) provided detailed three dimensional illustrations of
24 sauropod bone, and he drew the fibres of the non-lamellated primary bone with a longitudinal
25 orientation. However, he did not specifically describe the longitudinal arrangement, and this notion
26 has largely been ignored in the following decades. The results of the current study unequivocally
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1 support de Ricqlès' original conception, and suggest furthermore that this structural arrangement is
2 probably much more widespread among dinosaurs and large mammals (Ascenzi et al. 1967, Martin
3 & Ishida, 1989, Kerschnitzki et al. 2011, and references therein) than has been demonstrated so far.
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8 Building upon the overestimated amount of woven bone, paleohistological studies
9 concluded that one of the reasons why sauropods could reach their enormous adult sizes very fast is
10 because the bulk of the bony laminae in their long bones is composed of fast forming woven bone
11 (Sander et al. 2011 and references therein). Although several characteristics of sauropods indicate
12 that in general they must have grown very fast, our study demonstrated that the amount of woven
13 bone in itself cannot account for the assumed high growth rates; hence this reasoning does not hold
14 anymore. So, instead of focusing on the amount of woven bone, it seems much more important to
15 understand the meaning of its *presence* in the long bones of these animals.
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28 **(4) Biological inferences based on SO- and DO-derived tissues in fossils**

29 *(a) Woven bone: evolutionary implications for high growth rate*

30 The presence of non-pathologic and non-embryonic woven bone in the form of
31 “fibrolamellar bone” has been reported for the long bones of several extant and extinct tetrapods
32 (e.g. Enlow & Brown, 1957, 1958; de Ricqlès et al. 2003, 2008; Ray, Botha & Chinsamy-Turan,
33 2004; Klein, 2010; Mukherjee, Ray & Sengupta, 2010), however, in the light of our results, some of
34 these studies may have to be revised. Nevertheless, woven bone has been observed during the scale
35 formation of polypteriform and lepisosteiform fish; two extant but basal groups of actinopterygians
36 (Sire, Donoghue & Vickaryous, 2009). It is important to note that the woven fibered connective
37 tissues described in the scales of these fish are real bony tissues and not dentin-like structures, i.e.
38 they are derivatives of osteogenic and not odontogenic processes (Sire et al. 2009). The
39 undoubtedly basal position of these animals on the phylogenetic tree of vertebrates clearly indicates
40 that the potential for SO had been acquired by the time of diversification of more derived
41 vertebrates such as tetrapods. Since intramembranous as well as some phases of endochondral
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2 ossification follows the SO bone formation model (Ferretti et al. 2002; Palumbo et al. 2004;
3 Marotti, 2010) and because intramembranous ossification has “older” evolutionary origin than
4 endochondral ossification (Wagner & Aspenberg, 2011), SO must have appeared at latest by the
5 time intramembranous ossification was invented.
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10 The evolutionary order of appearance of SO and DO is much more obscure. It has been
11 demonstrated in the development of dermal plates and scutes of teleost fish that the formation of
12 woven bone precedes the deposition of parallel-fibered bone (Sire et al. 2009). Furthermore, it was
13 suggested that DO can only take place in the presence of preexisting osteocytes because the orderly
14 recruitment of DO-osteocytes is controlled by the signal-transduction of the preexisting osteocyte
15 syncytium (Marotti, 1996; Palumbo et al. 2004). Both of these indicate certain developmental
16 constraints on the sequence of SO and DO and suggest that the occurrence of SO might have
17 preceded that of DO in the evolutionary scenario, as well. However, their concurrent appearance is
18 the most plausible hypothesis given the adaptive nature of DO-derived highly organized bone which
19 lies in its superior biomechanical characteristics compared to woven bone (Currey, 2002, see also
20 below). This unequivocally implies that woven bone must be widespread among all kind of
21 vertebrates, and that its presence cannot be considered as an evolutionary innovation in achieving
22 high growth rates. The question which should rather be posed is when, why and how SO-derived
23 woven bone got involved into the periosteal growth of long bones of tetrapods and how do these
24 issues relate to growth rates.
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46 *(b) Interrelationships between vascularity, BMR, body size and growth rate*
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48 Vascular canals play a crucial role in the formation of woven bone. The mostly avascular
49 long bone cortices of small extant amphibians and lepidosaurs (Foote, 1916; Enlow & Brown, 1956,
50 1957, 1958) exhibit only DO-derived lamellar or non-lamellar highly organized primary bone. This
51 exclusively accretional, slow bone growth limits the growth rate of these animals. However, from a
52 certain size, temnospondyl amphibians and large extant varanids also show various degrees of
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2 vascularization in the bone cortex (Enlow & Brown, 1958; de Ricqlès et al. 2004; Steyer et al.
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4 2004; Buffrénil, Houssaye & Böhme, 2008). In varanids, this size-specific appearance of vascular
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6 canals is independent of phylogeny and seems to reflect the absolute growth rate of the cortex
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8 (Buffrénil et al. 2008). Cubo et al. (2005) also demonstrated the positive correlation between bone
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10 size and vascular density in sauropsids. Nevertheless, it is still to be explored whether there is a
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12 critical shaft diameter (which correlates with body mass in graviportal and mediportal tetrapods,
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14 [Anderson, Hall-Martin & Russel, 1985; Alexander, 1989]) at which vascular canals start to be
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16 incorporated into the long bone cortex of tetrapods. Besides this size factor, the relatively highly
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18 vascularized cortices of extant crocodylians could be considered a reversal from a more advanced
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20 thermal and metabolic biology that characterized their archosaurian ancestors (de Ricqlès et al.
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22 2003, 2008; Seymour et al. 2004; Cubo et al. 2012). In fact, some studies concluded that extant
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24 crocodyles still possess the capability of growing fast under certain circumstances (Tumarkin-
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26 Deratzian, 2007; Owerkowicz, Eley & Hicks, 2009; Cubo et al. 2012).

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31 Size-dependent vascularity, i.e. the trend that larger-bodied forms show more densely
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33 vascularized long bone cortices, also applies to extant homeothermic amniotes with generally high
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35 basal metabolic rate (BMR) and growth rate, namely birds and mammals (Klevezal, 1996 pp. 22;
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37 Padian, de Ricqlès & Horner, 2001; Erickson et al. 2009). Nevertheless, they still possess a higher
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39 number of cortical vascular canals than a poikilotherm animal of similar size but of much lower
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41 BMR. The presence of this trend in vascularity vs. body size in both groups implies a common
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43 cause which must be irrespective of the physiological differences between them. This common
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45 cause most probably originates in biomechanics, fluid mechanics and scaling laws related to
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47 nutrient transport within the bone tissue (Mishra, 2009). Above a certain distance between the blood
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49 supply and cells (~150 μm), further fluid and solute transportation requires additional energy input
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51 because of growing hydraulic resistance. The relatively constant distance found in large bodied
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53 dinosaurs (*Iguanodon*) and mammals (cow) implies a functional and metabolic optimization of the
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55 transportation path distance in the vascular-lacunocanalicular system of bones (Mishra, 2009). On
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2 the other hand, the apparent deviations in the degree of vascularity of equally sized members of the
3 considered clades might be explained by the differences in BMR. High BMR is related to enhanced
4 aerobic capacity associated with elevated oxygen consumption of all cellular tissues (Bennett &
5 Ruben, 1979; Biewener, 2003) including bone, therefore the osteocytes in animals with high BMR
6 have higher metabolic demands than the osteocytes of similar sized animals with lower BMR.
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8 Providing the osteocytes with sufficient supply at high rates requires more extensive vascularity in
9 the bone cortex than that found in similar sized animals with low BMR (Montes, Castanet & Cubo,
10 2010; Seymour et al. 2011). Furthermore, the complexity of the vascular-lacunocanalicular
11 organization of bones also increases with increasing body mass and BMR (Mishra, 2009), and
12 BMR itself universally scales with respect to body mass (Bishop, 1999; White & Seymour 2005;
13 Speakman, 2005; West & Brown, 2005; but see Kolokotronis et al. 2010). In sum, there is a
14 positive correlation between vascularity (density and/or total porosity percentage) and body size, as
15 well as vascularity and BMR (Montes et al. 2010). Vascularity in the long bone cortices of different
16 tetrapods is most likely determined by the combination of body size and BMR with mostly
17 structural and functional components, even though a phylogenetic influence is also to be expected
18 to some degree (Cubo et al. 2005). The obvious relation between BMR and growth rate is that only
19 high BMR can motorize fast growth, since besides sustaining other essential body functions, a
20 considerable amount of energy must be invested into growth (Cubo et al. 2008; Montes et al. 2007,
21 2010). Dense vascularization of the fast growing bone characterized by elevated tissue metabolism
22 is also essential above a certain cortical thickness (i.e. body size, see above). The fact that body size
23 of the largest poikilotherm tetrapods never exceeds that of the largest endotherms in a similar
24 environment implies that growth rate limits maximum body size/body mass an animal can achieve
25 in its lifespan. Hence, high vascular densities and enormous body sizes of sauropods all point to
26 high BMR and high growth rates in these animals.

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57 *(c) The role of woven bone in fast growth: a model for bone histomorphogenesis in sauropods*
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2 Due to the characteristics of woven bone (providing *de novo* formed scaffold for DO, see
3 above), the involvement of SO into the primarily DO-based periosteal growth of the long bone shaft
4 greatly extends the potential upper limits of diametrical growth rates. In the following description,
5 we demonstrate how woven bone can contribute to the acceleration of long bone growth throughout
6 sauropod ontogeny. For the sake of simplicity, only diametrical growth of the long bone shaft will
7 be considered here.
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11 In the sauropod embryo, the cartilage precursor of limb bones is embedded in a thick layer
12 of densely vascularized mesenchyme. The outermost region of the cartilage precursor is an
13 avascular layer of condensated progenitor cells, called the perichondrium (Hall, 2005). Signaling of
14 the hypertrophied chondrocytes in the cartilage precursor stimulates the invasion of capillaries, and
15 induces the differentiation of the perichondrial cells into osteoblasts, ultimately forming the
16 periosteum (Karaplis, 2008). At this stage of development, the periosteum corresponds to the bone
17 collar, which is the precursor of the cortical region of the limb bone (Ferretti et al. 2002; Hall, 2005;
18 Karpalis, 2008). The bone collar is a thick, vascularized tissue layer surrounding the cartilaginous
19 shaft, in which further maturation of pre-osteoblasts is either controlled by SO or DO processes. As
20 SO starts to take place, randomly oriented mesenchymal cells condensate into 1-3 cells thick layers
21 half way between the adjacent capillaries of the bone collar, and start to differentiate into stationary
22 osteoblasts. At the same time, in other regions of the periosteum, DO-controlled osteoblasts are
23 being organized into osteogenic laminae. Whereas the static osteoblasts produce the first thin
24 trabeculae of porous, irregularly fibred woven bone between the capillaries, the DO-derived
25 osteogenic lamina starts to deposit highly organized bone matrix on already existing surfaces like
26 the limb precursor or the forming SO-derived trabeculae. Since the SO-derived woven framework
27 incorporates large vascular spaces into the bony substance within a short period of time, fast
28 diametrical growth can be realized by means of fast volume expansion of the forming limb bone
29 shaft. SO-osteoblasts start to mineralize their secreted matrix very early, filling up the pores with
30 apatite crystallites. As this SO process extends diametrically and more and more vascular spaces get
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1 enclosed into the shaft, the firm substrate of SO-derived trabeculae provides the initial conditions
2 for the formation of more and more osteogenic laminae on their surface. With the deposition of
3 highly organized bone on the surface of the SO-scaffold, the DO-derived laminae uniformly thicken
4 the trabeculae in all directions and fill up the entrapped vascular spaces. The onset of this process
5 launches the formation of primary osteons (redefined in this study, see above). Meantime, the SO-
6 osteoblasts in the woven trabeculae differentiate into osteocytes *in situ* retaining their unorganized
7 osteoblastic appearance. From time to time, those DO-osteoblasts of the osteogenic lamina which
8 are destined to become osteocytes, stop secreting matrix and get buried into the osteoid
9 continuously produced by the neighboring osteoblasts. As a consequence, the osteogenic lamina
10 moves away from the entrapped osteoblasts. The latter soon differentiate into osteocytes in a
11 spatially highly organized manner aligning their long axis to the long range order of the collagen
12 fibres, parallel to the long axis of the limb bone. The osteoblasts of the osteogenic lamina begin to
13 mineralize the deposited extracellular matrix during which process plate-like apatite crystallites
14 develop along and within the collagen bundles with their c-axis (long axis) following the long axis
15 of the fibres.
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35 After the sauropod embryo hatches, the diametrical expansion as well as compaction of the
36 developing limb bone performed by SO and DO processes, respectively, continue the same way in
37 the active growth phases of postembryonic ontogeny. As DO proceeds and compacts the primary
38 vascular spaces, the osteogenic lamina may at some point start to form lamellae around the
39 progressively narrowing vascular space. These lamellae might have alternating fibre orientation in
40 which case the secretion activity of the osteogenic lamina slows down or ceases periodically while
41 all the osteoblasts in the lamina uniformly and synchronously change their orientation, and soon
42 they start to produce a new sheet of highly organized matrix but with differently oriented collagen
43 bundles. In sum, the combination of SO-controlled fast volume expansion and DO-controlled bone
44 compaction characterizes the phases of active growth, and their proportional contribution to the
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2 growth process determines the diametrical growth rate of the limb bone in the different growth
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4 phases.

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6 As the animal matures and approaches its maximum size, less and less vascular canals get
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8 encased into the growing bone, whereby the high diametrical expansion rate of the limb bone
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10 drastically decreases. Since there is a well-defined bony substrate, the surface of which is much less
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12 interrupted by vascular canals by this time, DO-derived bone deposition now dominates the growth
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14 of the outermost cortical region. As soon as no SO-derived woven bone is needed anymore, the
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16 diametrical growth of the limb bone is ensured exclusively by DO processes. At this stage, the
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18 relatively thin periosteum does not exhibit any SO ossification centers, but only its innermost cell-
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20 layer is organized into a unified osteogenic lamina which still deposits bone on the surface of the
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22 bone shaft. This slow accretional growth provided by the periosteum is not perpetual but rather
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24 cyclical producing lines of arrested growth in the almost avascular outermost cortex. The onset of
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26 this process is the onset of an EFS (*sensu* Ham, 1953). In each and every subsequent cycle, the
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28 amount of bone deposited by the periosteum decreases, and as a consequence, LAGs get more
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30 densely spaced. This residual growth can go on for a long time, however, the amount of deposited
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32 bone is so negligible that by this time the animal is said to be skeletally fully grown. Finally, the
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34 osteoblasts of the periosteum differentiate into lining cells (basically inactive osteoblasts), and this
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36 membrane of lining cells covers the fully grown bone shaft.
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42 The model clearly demonstrates that woven bone has a crucial role in fast diametrical bone
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44 growth. However, the significance of woven bone does not lie in its absolute amount in the growing
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46 bone, since woven bone trabeculae make up only a low fraction (~14-25%) of the entire bony
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48 lamina. Its high formation rate is clearly of importance but not in the same sense as it would be if
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50 the majority of the primary bony substance was composed of woven bone as it has always been
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52 assumed. There is not one particular feature but rather a combination of key features of woven bone
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54 (*de novo* formation without substrate, high formation rate and fast mineralization) which allows it to
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56 enclose extensive vascular spaces with its forming network of trabeculae, thereby creating a highly
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1 porous, fast expanding composite of vascular cavities and bony substance. Subsequent deposition of
2 DO-derived highly organized bone is responsible for the infilling of the extensive vascular spaces
3 and providing the skeleton with higher resistance against mechanical loading.
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8 Although the SO-controlled volume expansion is much faster than the DO-controlled
9 compaction of the growing bone, it is important to note, that the comparison of the formation rate of
10 SO-derived woven bone with that of DO-derived highly organized bone can be very misleading
11 because they have such different formation principles. First of all, only the actual amount of bony
12 substance should be reckoned with when formation rate of SO- vs. DO-derived bone is considered,
13 since woven bone cannot take credit for the huge enclosed vascular spaces. Second, whereas the
14 irregular orientation of the secretory territories of the static osteoblasts results in a highly porous
15 matrix, the matrix deposited by the osteogenic lamina is much more compact. This means that the
16 bulk of the woven bone matrix consists of holes which only provide extra volume but do not
17 contribute to the dry organic matter of the bone tissue. If osteoid (mainly collagen) secretion rate is
18 considered to be the true measure of bone formation rate, then the secretional activity of the SO-
19 and DO-osteoblasts must be compared. The osteoid secretion rate of each osteoblast depends on the
20 amount of its protoplasm engaged on a given bone surface (Marotti, 1976; Ferretti et al. 2002).
21 Since no structural or ultrastructural differences have been observed between SO- and DO-
22 osteoblasts so far (Marotti, 2010), we hypothesize that secretion activity can be highly variable
23 depending on several exogenous and endogenous factors (e.g. actual physiological state,
24 ontogenetic stage, environmental conditions etc.) but might be irrespective of the SO or DO origin
25 of osteoblasts. This suggests that on the cellular level, the secretion activity i.e. matrix production
26 rate of SO-derived osteoblasts is not necessarily higher than that of DO-osteoblasts.
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51 Osteocyte density clearly depends on the incorporation rate of the osteoblasts into the
52 matrix, and it has been suggested by some authors that it represents overall growth rate (Bromage et
53 al. 2009; Skedros, Hunt & Bloebaum, 2004). SO-osteoblasts differentiate into osteocytes *in situ*,
54 thus the term incorporation rate has no meaning in the context of woven bone. The primary density
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2 of SO-osteoblasts, on the other hand, could be related to the growth rate of the animal; however, the
3 assumed correlation is still to be explored. The density of osteocytes in DO-derived bone tissue
4 depends on how many osteoblasts stop producing matrix in a given area within a given time period
5 (i.e. on the incorporation rate) which, similarly to the secretion activity of the cells, might also be
6 influenced by a variety of exogenous and endogenous factors. Thus, incorporation rate and thereby
7 osteocyte density of DO-derived bone tissues may indeed correlate with growth rate but this
8 relation is still largely unexplored.
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17 It must be emphasized that woven bone is always present in fast growing bones even if its
18 significance does not lie in its absolute volume. This means that its presence in long bones is indeed
19 indicative of fast growth. However, in large bodied animals, woven bone can only initiate fast
20 volume expansion if sufficient amount of blood vessels are also provided. Thus, allowing for the
21 bone growth related issues discussed above, the most reliable histological indicator of bone growth
22 rate of all candidates suggested so far (e.g. vascular architecture, fibre orientation, woven bone
23 amount, osteocyte density etc. see above) is probably the vascular density and/or total porosity
24 percentage (de Margerie, Cubo & Castanet, 2002). Nevertheless, this assumption also needs further
25 testing without neglecting possible phylogenetic, biomechanical and/or developmental constraints.
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39 *(d) Evolutionary implications on fast growth*

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41 The evolutionary innovation which gave way to fast skeletal growth (with special emphasis
42 on the appendicular skeleton) was the involvement of static osteogenesis into primarily dynamic
43 osteogenic processes, i.e. *de novo* bone formation into accretional bone formation in the diametrical
44 growth of limb bones. Combined with the enhanced blood supply of the developing bone, SO-
45 derived woven bone permits fast volume expansion, whereas DO-derived highly organized bone
46 accounts for the mechanical stability of the growing skeleton. In many vertebrates that exhibit this
47 combination in their limb bone development, fast skeletal growth has been experimentally proven
48 (Castanet et al. 2000; de Margerie et al. 2002; 2004; Starck and Chinsamy 2002).
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2 The incorporation of SO into predominantly DO-controlled skeletal growth processes
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4 evolved independently in several tetrapod lineages (see “fibrolamellar bone” in de Ricqlès et al.
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6 2003, 2008; Botha-Brink, Abdala & Chinsamy-Turan, 2012; Ray, Bandyopadhyay & Bhawal,
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8 2009). The independent appearance of fast skeletal growth by means of fast volume expansion
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10 through simultaneous encasement of extensive vascular spaces is further supported by the apparent
11
12 independent evolution of high BMR in birds and mammals. In an integrative biological view, all
13
14 fundamental resources (mitochondria, advanced circulatory system and lungs for acquiring high
15
16 BMR, SO- and DO-derived bone) had already evolved in the earliest tetrapods. Evolving a more
17
18 active metabolism, enabling fast growth, therefore became possible on several lineages. A
19
20 considerable evolutionary flexibility of physiological traits is exemplified by the reverse
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22 thermobiology and growth strategy of modern crocodiles (de Ricqlès et al. 2003, 2008; Cubo et al,
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24 2012; Seymour et al. 2004; Tumarkin-Deratzian et al. 2007; Woodward, Horner & Farlow 2011).
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26 Thus, the evolution of high BMR as well as fast growth is most likely motorized by long-term
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28 environmental and ecological factors and less constrained by constructional aptitude of vertebrates.
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33 Our new interpretation on the histomorphogenesis of sauropod laminar bone has significant
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35 impact on the reconstruction of the evolution of sauropod gigantism. However, our results also
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37 support the long held view that the growth rate of sauropods was very high, even if this assumption
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39 is mostly based on the extensive bone vascularity and enormous body sizes. Once this capacity was
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41 there, almost everything became possible: from reaching body masses up to 80-100 tons (e.g.
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43 *Argentinosaurus*, [Hokkanen, 1986], *Amphicoelias* [Paul, 1998]) to the complete loss of gigantic
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45 dimensions probably along with physiological reversals and becoming island dwarfs (*Europasaurus*
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47 [Sander et al. 2006] and *Magyarosaurus* [Stein et al. 2010]).
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51 52 53 *(e) Implications for biomechanics*

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55 The main function of the skeleton is to provide the body with mechanical support, thus bone
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57 is required to resist mechanical forces. Bone strength depends on the microarchitectural features,
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1 turnover rate and characteristics of the mineral and collagen phases (density, orientation) of the
2 bone tissue. In this complex system, the mineral constituent is responsible for the stiffness of the
3 bone, whereas collagen with its high tensile strength provides the bone with toughness by assuring
4 flexibility and absorbing the energy of forces acting on the bone (Viguet-Carrin, Garnero &
5 Delmas, 2006 and references therein). The mineral component is mostly integrated into the collagen
6 fibrils (Ottani et al. 2001), hence the crystallite orientation is determined by the orientation of the
7 fibres. Since the amount and orientation of these two components are clearly decisive in the
8 biomechanical properties of bones (Acsezi & Bonucci 1967, 1968; Beniash, 2011; Currey, 1987;
9 Frost, 1994), differences in their spatial arrangement as well as relative proportions reported in
10 woven bone vs. highly organized primary bone must reflect the different mechanical behaviour of
11 SO- and DO-derived bone tissues. The SO-derived woven bone has no long range preferred fibre
12 orientation, it is initially highly porous, and later becomes highly mineralized. Since apatite crystal
13 orientation is parallel to the long axis direction of adjacent collagen fibrils in both intra- and
14 extrafibrillar spaces, the long range random arrangement of fibrils and the extensive extrafibrillar
15 spaces in woven bone result in a more irregular spatial alignment of crystals (Su et al. 2003). As a
16 result, woven bone is very brittle and cannot withstand considerable mechanical impacts. In
17 contrast, in DO-derived highly organized bone, the closely packed fibril bundles with integrated
18 crystallites have a long range preferential orientation. Although it is less mineralized than woven
19 bone, it has improved mechanical properties, which indicates that collagen fibre orientation is much
20 more important than the degree of mineralization in determining the mechanical properties of bone
21 (Martin & Ishida, 1989; Marotti, 2010). Thus, predominance of woven bone in the weight bearing
22 limb elements characterizes mostly embryonic stages, in which mechanical constraints on the
23 skeleton are certainly much less than in any post-hatching stage. In extant animals, a proportionally
24 large amount of woven bone in non-embryonic developmental stages can only be found in the limb
25 bone cortex of very small species with high BMR such as mice (Kerschnitzki et al. 2011). As body
26 mass increases, the proportional amount of woven bone decreases, and DO-derived highly
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1 organized bone tissue will dominate the cortex of the limb elements. The low percentage of woven
2 bone in favour of HOPB in the primary cortex of the sauropod limb bones is therefore to be
3 expected.
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8 The origin of the predominantly longitudinal organization of DO-derived highly organized
9 bone in sauropods remains an important question. The significance of the orientation of mineralized
10 collagen fibres in bone strength has been shown by several studies (e.g. Ascenzi & Bonucci, 1967,
11 1968; Martin, 1993; Martin et al. 1996; Martin, Burr & Sharkey, 1998; Boyde & Riggs, 1990;
12 Riggs, Lanyon & Boyde, 1993; Bromage et al. 2003; Skedros et al., 2003; Skedros & Hunt 2004;
13 Skedros et al. 2007) all concluding that in a limb bone, longitudinal fibre orientation is found in the
14 cortical regions under tension, whereas transversally running fibres characterize the areas under
15 compressive loading. These results suggest that collagen fibre orientation can be used to infer main
16 loading regimes on the bone. Nevertheless, limitations of this method originate from the diverse
17 endogenous and exogenous factors which determine predominant collagen fibre orientation (CFO)
18 in the limb bones and whose proportional influence can change throughout ontogeny. In earlier
19 developmental stages, CFO seems to be controlled mainly by genetic and epigenetic factors
20 (endogenous), whereas extragenetic stimuli, such as mechanical loading or microcracks only later
21 become the prevailing inducers of fibre arrangement and/or rearrangement (Skedros et al. 2007).
22 Findings of Riggs et al. (1993) support this hypothesis, demonstrating that the consistent CFO
23 pattern found in the radius of adult horses, which unequivocally reflected the consistent pattern of
24 stress-strain distribution in the bone, did not correspond to the predominantly longitudinal CFO
25 revealed in the foals. Longitudinal CFO characterized the entire primary cortical bone, seemingly
26 irrespective of strain directions, however, remodeled areas showed tensile (longitudinal CFO) as
27 well as compressive (transverse CFO) secondary osteons (Ascenzi & Bonucci, 1967, 1968)
28 following the region-specific pattern of mechanical loading (Riggs et al. 1993). Thus,
29 predominantly longitudinal CFO in the primary bone of horses, the histology of which is strikingly
30 similar to that found in the humerus of *Alamosaurus* TMM 46300-2 (Fig. 4B), seems to be
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1 controlled by genetic and epigenetic regulatory systems rather than by mechanical stimuli. This is in
2
3 accordance with the conclusions of the study of Skedros et al. (2007) which were based on the
4
5 pattern found in ovine calcanei. Primary plexiform bone exhibits the same preferred longitudinal
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7 CFO in ovine, bovine and sauropod limb bones (Ascenzi et al. 1967; Martin & Ishida, 1989;
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9 Kerschnitzki et al. 2011; this study). This conformity in the histological composition of the limb
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11 bones of very distantly related groups (mammals and dinosaurs) suggests functional (analogous)
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13 rather than identical genetic and epigenetic (homologous) fibre organizing principles. On the other
14
15 hand, the apparent variability in predominant CFO in a monophyletic clade like dinosaurs (e.g.
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17 *Scutellosaurus* [Padian, Horner & de Ricqlès, 2004], *Jeholornis* [Erickson et al. 2009] or even
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19 *Ampelosaurus* with its “modified laminar bone” [Klein et al. 2012]) further implies that
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21 functionality may obscure the phylogenetic signal in this histological feature. The functional aspects
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23 of vascular architecture have also been demonstrated (de Margerie, 2002; Skedros & Hunt, 2004, de
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25 Margerie et al. 2005). Thus, CFO along with vascular architecture probably reflects the functional
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27 constraints present throughout ontogeny (Skedros et al. 2003). However, predominantly
28
29 longitudinal CFO in the HOPB may also reflect the ancestral condition of tetrapods. Finally, it is
30
31 important to note that, contrary to Amprino’s rule (Amprino, 1947), which is generally applied in
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33 paleohistological studies, the concept that CFO in primary bone is in itself suggestive of growth rate
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35 is highly unlikely.
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44 IV. CONCLUSIONS

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48 (1) Longitudinal thin sections of sauropod long bones revealed that in contrast to the general
49
50 reconstruction, the amount of woven bone in sauropod long bones is only a few cells thin
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52 layer in the laminae and the majority of the primary bone is longitudinally organized. This
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54 structural organization also characterizes the plexiform primary bone of large bodied
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1 mammals, and is likely to be more widespread among large tetrapods than previously
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3 thought.
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5
6 (2) The overestimation of the amount of woven bone in sauropod primary bone originates
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8 mostly from the almost exclusive usage of transverse sections examined under crossed plane
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10 polarizers, whereby the optical behaviour as well as the LCN characteristics of the different
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12 bony constituents have been misinterpreted.
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15 (3) Current biological studies, which revealed two major bone formation types, static and
16
17 dynamic osteogenesis (SO and DO) resulting in woven and highly organized bone,
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19 respectively, gave detailed explanation for the detectable differences between bone tissues
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21 of different developmental origin. Our better understanding of the formation process and
22
23 role of SO- and DO-derived bone tissues demonstrates the need for investigating optical
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25 behavior combined with LCN characteristics of thin sections of fossil bones in at least two
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27 section planes to make sure that the differentiation of the tissue types is justified.
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31 (4) In the revised palaeohistological terminology presented here, HOPB is introduced to
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33 describe all (lamellar and non-lamellar) DO-derived primary bone tissues and differentiate
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35 them from SO-derived woven bone. In accordance with the long range parallel fibre
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37 orientation in HOPB, the term primary parallel-fibred bone should be synonymized with
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39 HOPB rather than used to describe the suggested intermediate state of spatial organization
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41 of fibres which, based on the presented bone formation principals, may not even exist.
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43 Lamellation should be identified using both the characteristics of extinction pattern and the
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45 presence of fine lines. Based on the common formation principals but diverse structural
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47 organization of HOPB, we suggest that the term primary osteon include the entire DO-
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49 derived primary tissue (lamellated and non-lamellated) which has been deposited on the SO-
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51 derived scaffold of woven bone around the vascular canals. The widely used term
52
53 “fibrolamellar” should be discarded as it demonstrates neither formation principals nor
54
55 structural diversity of primary bone tissues. Instead, we recommend the usage of vascular
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1
2 architecture for structural classifications of fast growing primary composite tissue of woven
3 bone and HOPB.
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- 5
6 (5) The most important evolutionary innovation, which made fast growth possible, and which
7 seems to have happened independently on several lineages, is most probably the
8 incorporation of woven bone into the primarily DO-derived periosteal growth of the long
9 bone shaft.
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15 (6) The significance of woven bone in long bone growth rates does not lie in its amount but
16 rather in its capacity to rapidly enclose large vascular spaces thereby ensuring fast
17 diametrical growth. SO accounts for the fast volume expansion, whereas DO guarantees
18 compaction and improvement of the mechanical properties of primary bone tissues.
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24 (7) The primary indicator of high bone growth rates is most likely vascular density and/or
25 porosity, whereas woven bone is probably only a secondary requirement needed to encase
26 the extensive vascular network into the growing bone. Since sauropod long bones all exhibit
27 high vascular densities in their plexiform primary tissues, we support the long held view that
28 sauropods had similar growth rates to those of extant endotherms.
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35 (8) The brittle, hypermineralized SO-derived woven bone is mechanically reinforced by the
36 deposition of DO-derived HOPB, in which long range fibre orientation (CFO) is most likely
37 controlled by genetic and epigenetic factors in earlier ontogenetic stages, and by mechanical
38 requirements in more advanced ontogenetic stages. Since CFO in secondary osteons of long
39 bones is a more reliable indicator of loading regimes than the predominant CFO is in
40 primary osteons, functional aspects may be more reflected in secondary than in primary
41 bone tissues.
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51 (9) The mostly longitudinally organized HOPB found in the plexiform bone of sauropods and
52 large bodied mammals might imply functional convergence or alternatively can indicate the
53 ancestral tetrapod condition.
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2 (10) Preferred CFO in the primary bone of dinosaurs or any other tetrapods is not indicative of
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4 growth rate disputing this aspect of Amprino's rule which has been the basis of inferences
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6 drawn in several palaeohistological studies.
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12 13 **V. ACKNOWLEDGMENTS**

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17 The authors thank P. Martin Sander for access to sauropod samples in the histotheca at the
18
19 Steinmann Institut, and for fruitful discussion on the nature of fibrolamellar bone. Olaf Dülfer is
20
21 acknowledged for technical assistance in preparing thin sections. One anonymous reviewer and
22
23 Sarah Werning provided useful comments which improved the standards of the manuscript. K.
24
25 Stein was funded by the DFG, E. Prondvai was financed by ELTE-MTA Lendület Program (Project
26
27 nr. 95102). This is contribution no. 131 of the DFG Research Unit 533 Biology of the Sauropod
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29 Dinosaurs: The Evolution of Gigantism.
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35 **VI. APPENDIX 1. INSTITUTIONAL ABBREVIATIONS**

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39 CM Carnegie Museum of Natural History, Pennsylvania, USA; NHUB Naturkundemuseum of the
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41 Humboldt-Universität Berlin, Germany; OMNH Oklahoma Museum of Natural History, Norman,
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43 Oklahoma, U.S.A.; SMA Sauriermuseum Aathal, Aathal, Canton Zürich, Switzerland; TMM Texas
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45 Memorial Museum, Austin, Texas, U.S.A.
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53 **VII. REFERENCES**

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VIII. FIGURE CAPTIONS

Figure 1. Comparative overview of transverse vs. longitudinal thin sections of the plexiform primary bone of long bones of various sauropods under crossed plane polarizers. Comparison of the paired images of transverse (above) and longitudinal (below) sections brought to the same scale clearly shows that the majority of the dark areas in the transverse section, which has been identified as woven bone in previous studies, light up brightly in the longitudinal section disputing its presumed unorganized nature. True woven bone is represented only by a thin, centrally positioned dark stripe distinct on the longitudinal slides, whereas the rest of the bony lamina is evidently composed of highly organized primary bone (HOPB). The sections presented as paired images were prepared using the same bone drill core of the considered specimen. (A) *Alamosaurus* TMM 43600-2, humerus; (B) *Apatosaurus* CM 3378, humerus; (C) diplodocid indet. NHUB Ki2 (“*Barosaurus*” sensu Janensch, 1961; *Australodocus* sensu Remes, 2007), femur; (D) *Apatosaurus* “Chris” SMA M4/10-1, femur; (E) diplodocid indet. SMA 647/87-1, femur; (F) *Camarasaurus* “ET” SMA 0002, femur. Note that the dark patches in the longitudinal section of *Camarasaurus* do not reflect original fiber orientation but represent preservational artifacts.

Figure 2. Large-scale and projected close-up images of thin sections of the bony lamina. Both, transverse and longitudinal thin sections of (A-F) *Alamosaurus* TMM 43600-2 humerus and (G-L) *Apatosaurus* CM 3378 humerus are presented under single as well as crossed plane polarizers. Large-scale images (A,D,G,J) show the overall cortical microstructure in transverse (A,G) and longitudinal (D,J) sections under plane polarized light with the white square indicating the projected and magnified area of the cortex. 40x magnified transverse sections under plane (B,H) and cross polarized light (C,I), and corresponding longitudinal sections under plane polarized light (E,K) and crossed plane polarizers (F,L). Based on the observed lacunocanalicular network (LCN) features and extinction patterns in transverse and longitudinal sections of the specimens, only a few cells

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2 thin layer of woven bone is present, and the non-lamellar part of the highly organized primary bone
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4 (HOPB) has a long range longitudinal structural arrangement. Due to the prevailing longitudinal
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6 alignment of HOPB, the LCN is characterized by transversally cut, thus rounded lacunae (tol) with
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8 radial canalicular system (rc) in the transverse sections, and by longitudinally cut, thus elongate,
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10 spindle-shaped lacunae (lol) with perpendicular canalicular system (pc) in the longitudinal sections.
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12 Osteocyte lacunae of the woven bone (olw) are larger and have genuinely irregular shapes and
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14 canalicular network irrespective of the cutting plane. The extinction of transverse and
15
16 corresponding longitudinal sections under crossed plane polarizers is complementary in HOPB,
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18 whereas woven bone retains the same, darker appearance in both section planes. Further
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20 abbreviations: colw, confluent osteocyte lacuna of woven bone; HOPB, highly organized primary
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22 bone; vc, vascular canal (primary cavity). Bony lamina is defined *sensu* Sander (2000).
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28 **Figure 3.** Interpretative drawing representing a schematic model of the effect of the different
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30 cutting planes on the general 2D appearance of the lacunocanalicular network (LCN). 3D osteocyte
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32 lacuna model cut in three different planes (transverse, longitudinal and oblique) through (A) and
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34 offset from (B) its center of symmetry results in different projected 2D shapes, sizes and canalicular
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36 arrangements. Relatively to the long axis of the osteocyte lacuna, a transversally cut lacuna (tol)
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38 appears rounded with radial canalicular system, whereas a longitudinally cut lacuna (lol) exhibits
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40 elongate, flattened shape with perpendicular canalicular system. Other sectioning planes will result
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42 in obliquely cut osteocyte lacunae (ool) with variable transitional lacunocanalicular arrangements.
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44 The lacunae will exhibit the largest 2D area when they are cut through their center of symmetry
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46 (A). Depending on the level of the offset of the cutting planes relatively to the center of symmetry,
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48 the lacunae will grossly retain their shape but will appear smaller (B) compared to any situation
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50 represented in (A).
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2 **Figure 4.** Different degrees of lamellation in the plexiform primary bone of three different sauropod
3 specimens under single and crossed plane polarizers. Paired images are taken of the same section
4 under plane polarized light (above) and crossed plane polarizers (below). The bony lamina consists
5 of woven bone (wb), and lamellated (l) and non-lamellated (nl) HOPB. (A) Longitudinal section of
6 *Apatosaurus* CM 3378 humerus with three to four distinct lamellae which can be distinguished
7 around the primary vascular space (vc) based on the fine lines (fl) under plane polarized light, and
8 on the alternating fiber orientation (al) under cross polarized light. (B) Lamellation in the transverse
9 section of *Alamosaurus* TMM 43600-2 humerus is very extensive, being deposited right on the
10 surface of the woven bone, hence non-lamellar HOPB is absent. Lamellae can be traced in form of
11 densely packed and well-defined fine lines under plane polarized light, but are hard to observe
12 under crossed plane polarizers because originally the fiber orientation did not change in the
13 subsequent lamellae. Due to this preferred longitudinal orientation in each lamella, the entire bony
14 lamina (woven bone + HOPB) appears dark in the transverse section under crossed plane polarizers.
15 (C) At the other extreme, the magnified area of the transverse section of diplodocid indet. NHUB
16 Ki2 femur shows no lamellation, but exclusively non-lamellar HOPB. Woven bone is hardly
17 discernable (wb?) and seems to occur only in scarce patches in this specimen.
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39 **Figure 5.** Relative amount of lamina constituents in percentage of total lamina thickness in different
40 sauropod long bone samples based on values given in Table 2. Constituents are measured following
41 the definition of lamina *sensu* Sander 2000, from the center of one vascular canal to the center of
42 the subsequent one. This visualisation demonstrates the low amount (max 25% of the total lamina
43 thickness in diplodocid NHUBKi2) of woven bone (wb) compared to the HOPB in all sauropod
44 samples, but also the general pattern in proportional distribution of the different lamina constituents
45 measured in the investigated specimens. The only specimen that shows considerable deviation from
46 this pattern is *Alamosaurus* TMM 43600-2, humerus exhibiting proportionally high amount of
47 lamellar subunit in the total thickness of HOPB.
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4 **Figure 6.** Summarizing model of the macro- and microstructure of sauropod plexiform primary
5 bone. Three-dimensional visualization of the vascular architecture and long range fibre arrangement
6 in a slice of the primary cortex of a sauropod long bone (A), and the theoretical overall extinction
7 pattern of the transversally and longitudinally cut surfaces of the same slice under crossed plane
8 polarizers (D). Black strips are the primary vascular spaces with surrounding lines representing
9 lamellae. Between the vascular canals, the centrally positioned, irregularly shaped grey stripes
10 represent the woven bone layer; the end product of static osteogenesis (SO). Parallel lineation on
11 the longitudinally cut surface and dots on the transversal surfaces (A) indicate the preferred
12 longitudinal fiber orientation of the non-lamellar HOPB formed by dynamic osteogenesis (DO).
13 This longitudinal structural arrangement with the very thin woven layers in the bony laminae results
14 in a predominantly dark appearance of the transversal side, and a mostly bright appearance of the
15 longitudinal side of the 3D slice under crossed plane polarizers (D). White squares on the
16 transversal and longitudinal surfaces indicate the magnified and projected areas (B,C,E,F). (B,C)
17 Schematic drawing of the lacunocanicular network (LCN) arrangement in the bony laminae in
18 transverse (B) and longitudinal (C) sections with the indication of the cutting planes and the
19 resulted 2D appearance of a 3D osteocyte lacuna model (see also Fig. 2). (E,F) Optical features of
20 (B) and (C), respectively, under cross polarized light. Note that the modeled changes in the LCN
21 and extinction in the subsequent lamellae with alternating fiber orientation and the LCN and
22 extinction in the non-lamellar HOPB all reflect that the long range fiber orientation in DO-derived
23 HOPB corresponds to the long axis of the aligned lacunae, and is perpendicular to the canalicular
24 network (in accordance with Kerschnitzki et al. (2011)). The SO-derived woven bone does not
25 show any regular pattern in LCN, and has a uniformly dark appearance under crossed plane
26 polarizers. On the basis of formation principles, primary osteon is redefined here as the primary
27 vascular space surrounded by lamellated and/or non-lamellated DO-derived HOPB. The illustration
28 is largely based on *Apatosaurus* CM 3378, and therefore represents only one of the several possible
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1 structural organizations of primary tissue in sauropod long bones (for further details see main text).

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3 Abbreviations: es, endosteal surface; lp, longitudinal plane; ps, periosteal surface; tp, transversal

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5 plane; vc, vascular canal.
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For Review Only

IX. TABLES

Table 1. Details of the studied materials providing novel results. First description refers to the source publication of HOS and first description of transverse sections of the specimen.

Abbreviations: el, element type (h, humerus; f, femur); HOS, histological ontogenetic stage (*sensu* Klein and Sander, 2008); l, element length; sc, element smallest diaphysis circumference.

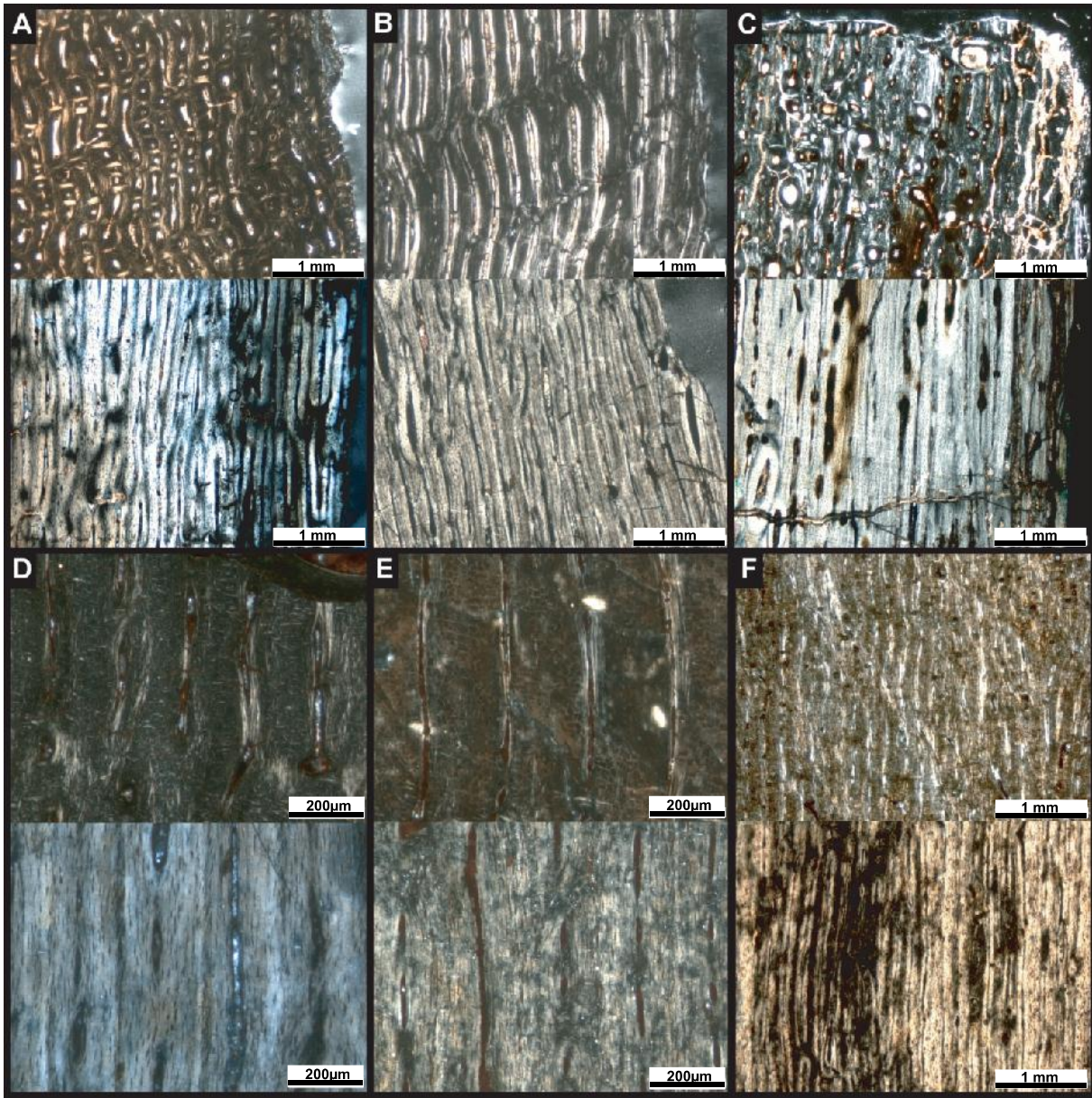
Taxon	Specimen id.	el	l (mm)	sc (mm)	HOS	first description
<i>Alamosaurus</i>	TMM 46300-2	h	915	375	7	Stein et al., 2010; Klein et al. 2012
<i>Apatosaurus</i>	OMN H1279	f	340	190	4	Klein and Sander, 2008
<i>Apatosaurus</i>	OMNH1278	h	258	144	4	Klein and Sander, 2008
<i>Apatosaurus</i>	CM 3378	h	980	492	11	Klein and Sander, 2008
<i>Apatosaurus</i>	SMA “Jaques”	f	1640	725	10	Klein and Sander, 2008
<i>Apatosaurus</i>	SMA “Chris” M4/10-1	f	1440	525	12	Klein and Sander, 2008
<i>Camarasaurus</i>	SMA “ET” 0002	f	935	422	12	Klein and Sander, 2008
diplodocid indet.	NHUB Ki2	f	1190	420	10.5	Sander, 2000; Klein and Sander, 2008
diplodocid indet.	SMA 647/87-1	f	1200	433	8	Klein and Sander, 2008

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Table 3. Linear regression parameters for element shaft circumference vs. absolute woven bone thickness, woven bone fraction and total lamina thickness in sauropod plexiform bone based on the mean values given in Table 2.

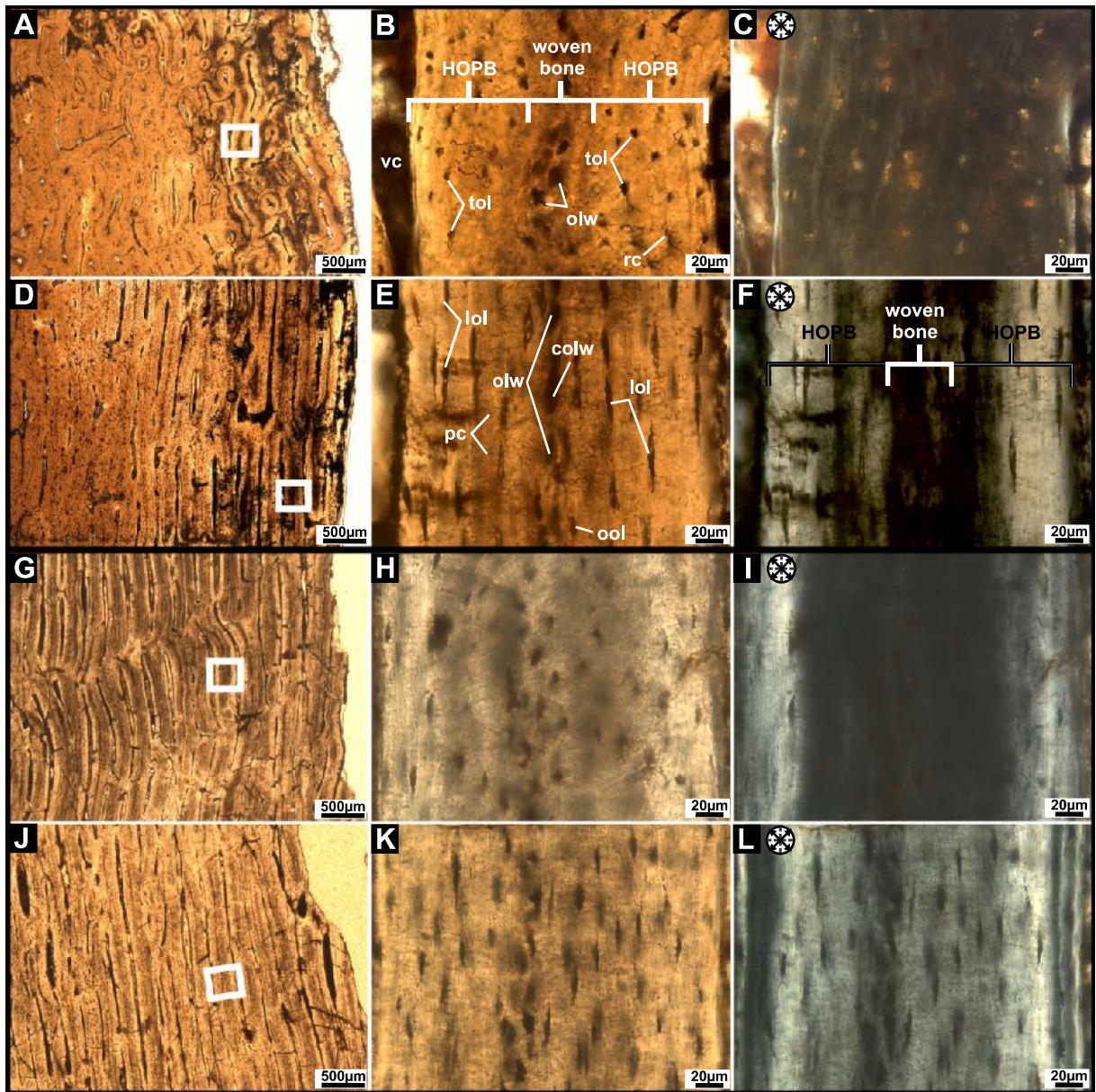
Linear regression independent	dependent	Pearson's R	df	Two tailed p-value	Intercept	Slope
shaft circumference	mean woven bone thickness	0.783	8	0.007	26.41	0.025
	mean lamina thickness	0.642	8	0.045	136.391	0.16
	mean woven bone fraction	-0.128	8	0.725	0.198	-0.0

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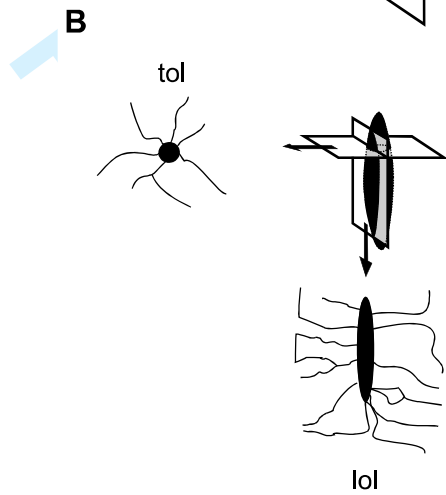
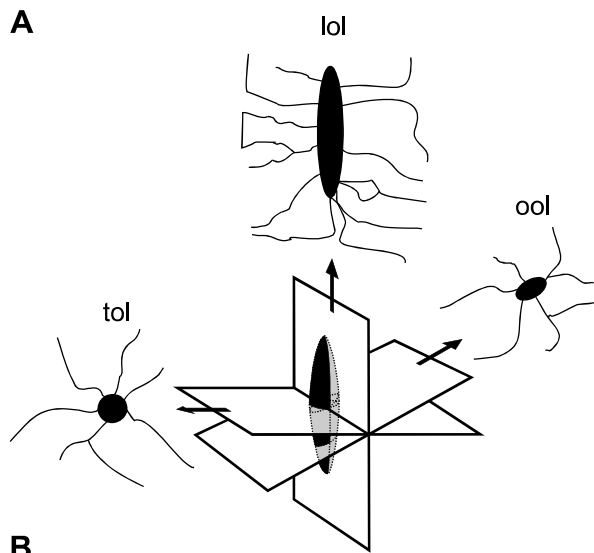


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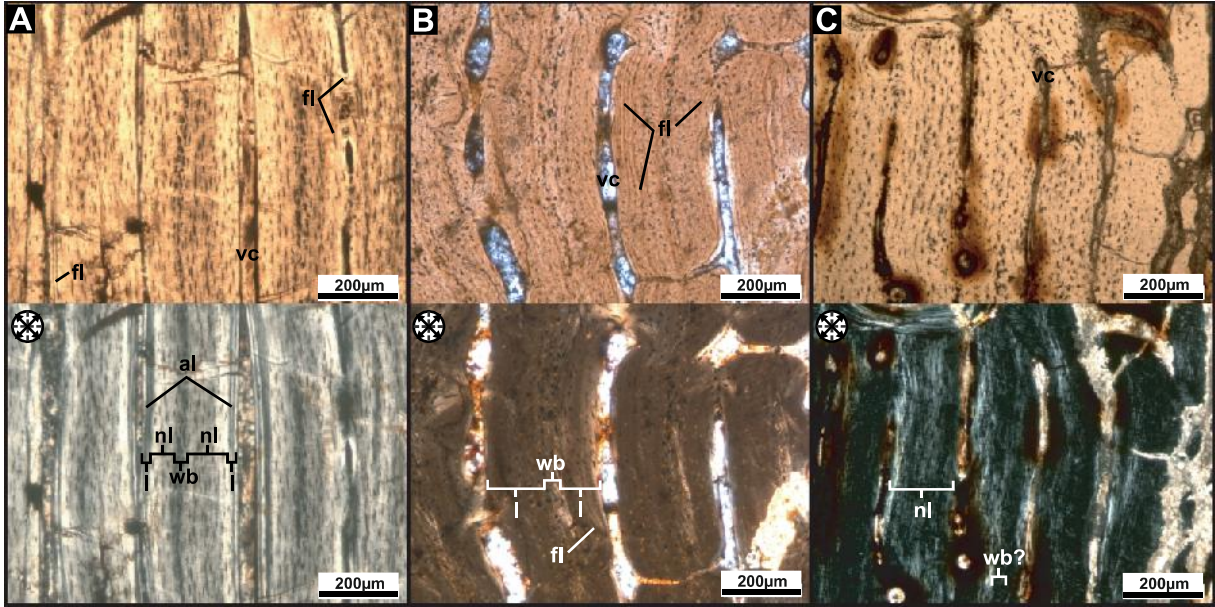


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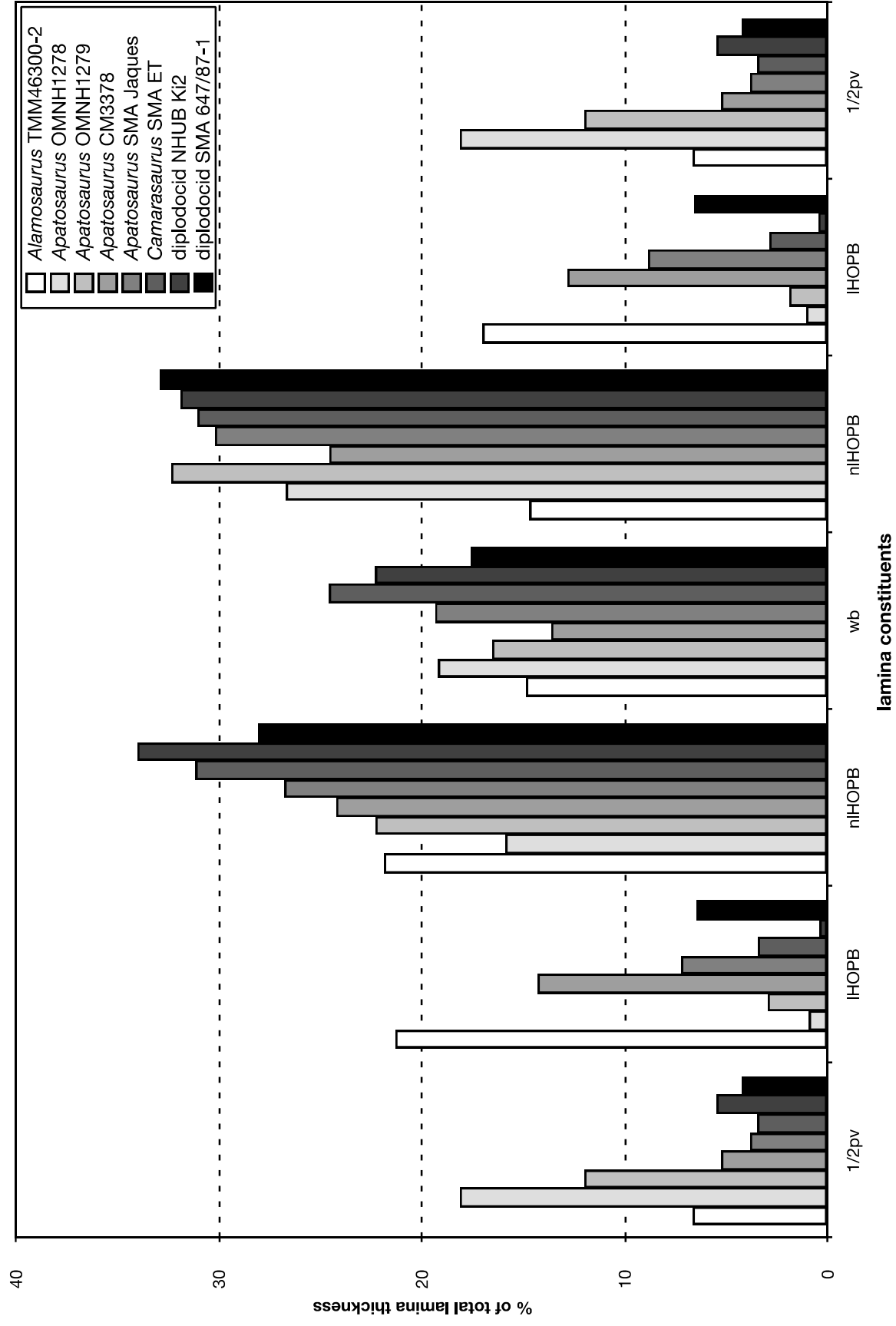


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