

Activity of the lysosomal cysteine proteinases (cathepsin B,H,L) and a metalloproteinase (MMP-7-ase) in the serum of cystic fibrosis homozygous children

Aranka LÁSZLÓ, I SOHÁR*, K GYURKOVITS

Department of Pediatrics, and *Department of Biochemistry, University Medical School, Szeged, Hungary

Received 4 March 1987

Lysosomal cysteine proteinase (cathepsin B, H, L) and metalloproteinase (MMP-7-ase) activities were measured from serum of 19 cystic fibrosis (CF) homozygotes and of 13 healthy children, as control group.

The activity of cathepsin B and H significantly increased in the CF-group.

INTRODUCTION

The plasma lysosomal hydrolases, arginine esterase and IRT (immuno reactive trypsin) had been investigated in patients suffering from cystic fibrosis (CF); the latter was recommended for the screening of CF [1-6].

The diminished activity of the pancreatic enzymes is known as a leading laboratory finding in CF. Koheil et al [7] have published diminished carboxipeptidase-B-like activity among the intracellular proteases.

The cysteine proteinases (CB, CH, CL) and the metalloproteinase (MMP-7-ase) belong to the intracellular proteinases. These cysteine proteinases are lysosomal enzymes (Table I) [8-9], while MMP-7-ase is a cytosol one [10].

Earlier the elevated activity of the cysteine proteinases was investigated

in the muscle tissue of dystrophic animals or of human myopathic patients [10-12].

There are no data available about the cysteine proteinases or metalloproteinase of CF patients, that was the reason of our investigations.

MATERIAL AND METHODS

The activity of cathepsin B, H and L (CB, CH, CL) as lysosomal cysteine proteinase, MMP-7-ase as a metalloproteinase was measured in the serum of 19 CF homozygotes and in 13 healthy control children (4 months - 15 y.). Cysteine proteinase activities were measured by the method of Kirschke et al [12] and MMP-7-ase activity was determined by the method of Nakajima et al [13] for measurement of cathepsin G activity by Sohár et al [10] modified to MMP-7-ase activity using Suc-Ala-Ala-Pro-Phe-AMC (AMC means 7-methyl-4-amino coumaryl) as substrate. The conditions of assays are summarized

TABLE I

Nomenclature of the intracellular proteases (according to Barrett, modified)

<i>Earlier names</i>	<i>Up-to-date names</i>
cathepsin (1929)	pepsin
cathepsin (1937)	cathepsin E (1962)
	cathepsin D (1960)
	cathepsin S (1975)
cathepsin II — cathepsin B (1941) (1952)	cathepsin B (1972)
	cathepsin B' — cathepsin B1 (1969) (1971)
	cathepsin L (1976)
	cathepsin N (1972)
	cathepsin H (1976)
	cathepsin B3
	cathepsin B2
cathepsin IV (1941)	lysosomal carboxy peptidase B (1975)
	lysosomal carboxy peptidase A (1975)
	lysosomal carboxy- peptidase C (1975)
	lysosomal aminopeptidase (1977)
	dipeptidyl-peptidase I (1972)
	dipeptidyl-arylamidase — dipeptidyl-aminopeptidase I (1965) (1968)
	peptidyl-dipeptidase (1972)
cathepsin I (1941)	lysosomal carboxy- peptidase A (1975)
cathepsin-like carboxypeptidase C (1972)	lysosomal carboxy- peptidase C (1975)
cathepsin III (1941)	lysosomal aminopeptidase (1977)
cathepsin Va (1951)	dipeptidyl-peptidase I (1972)
	dipeptidyl-arylamidase — dipeptidyl-aminopeptidase I (1965) (1968)
	peptidyl-dipeptidase (1972)
	carboxy-cathepsin (1971)

BARRETT A.J. (ed): *Proteinases in Mammalian Cells and Tissues*. North-Holland Publ. Co., Amsterdam (1977)

TABLE II

Biochemical conditions of assays of examined enzymes

Name	EC. No.	Substrate	Concentration in assay, μ M	pH	Buffer	Inhibitor	Catalytic group
Cathepsin B	3.4.22.1	Z-Arg-Arg-MCA	5.0	6.0	phosphate	chloroquine	-SH 2 mM
Cathepsin H	3.4.22.16	Arg-MCA	20.0	6.8	phosphate	Leu-CH ₂ -Cl	-SH 2 mM
Cathepsin L	3.4.22.15	Z-Phe-Arg-MCA	5.0	5.5	acetyte	Z-Phe-Phe-CHN ₂	-SH 2 mM
MMP-7-ase	3.4.24	Suc-Ala-Ala-Pro-Phe-MCA	5.0	7.5	Tris-HCl	EDTA	metallo ion

MCA: 7-methyl-coumarynil-4-amide, Arg: arginine, Phe: phenylalanine, Ala: alanine, Pro: proline
S: benzoyloxycarbonyl, Leu: leucine,
Suc: succinyl

TABLE III

The values of the activities of the serum cathepsin B, H, L and of the metalloproteinase (MMP-7-ase) in the group of CF homozygotes and of the control group

	Cathepsin = C mU/ml			MMP-7-ase
	CB	CH	CL	
CF homozygotes				
n = 19				
X =	17.39	123.65	1.66	7.67
S.D. ±	6.51	25.75	1.66	2.39
	p < 0.01	p < 0.01	p = 0.36	p = 0.67
	T = 3.32	T = 3.64	U* = 64.0	T = -0.82
Control group				
n = 13				
X =	10.26	89.6	1.3	8.4
S.D. ±	3.7	33.6	0.83	2.61

U = $\mu\text{mol AMC/min}$

U* = -Wilcoxon test

in Table II. The liberated AMC was measured fluorimetrically (Ex.: 360 nm, Em.: 460 nm) with a Hitachi 650-10 (Tokyo, Japan) spectrofluorimeter. Substrates were supplied from Enzyme System (Livermore, CA).

The correlation between the clinical stages (in Schwachman score) and the investigated enzyme activities, and between activities of the cysteine proteinases and of the MMP-7-ase was determined by linear correlation coefficients.

RESULTS

The values of serum cysteine- and metallo-proteinase activities of CF homozygotes and of control group are summarized in Table III. The activity of CB and CH significantly increased in the CF homozygotes' group, while the activity of CL and MMP-7-ase did not change. The serum cysteine proteinases, the MMP-7-ase and the clinical stages did not correlate sig-

nificantly according to the determination of the linear correlation coefficients.

DISCUSSION

The cathepsin B, H, L as cysteine proteinases can be activated with SH-group containing reagents or can be inhibited by SH-blockers.

Among the cysteine proteinases the CB and CH have showed significantly elevated activity in the serum of CF patients. The latter enzymes are localized into the lysosomes.

The metalloproteinase MMP-7-ase as a cytosol enzyme and the CL belonging to the cysteine proteinases did not show any enhancement of activity. The latter fact might be explained by the effect of the serum specific protease inhibitors.

The presence of cysteine proteinase inhibitors in the human muscle tissue

[14] and of endogenic cysteine proteinase inhibitors in the spleen has been proved [15]. There are no data about the cysteine proteinase inhibitors of CF patients which prompted us to determine the capacity of the latter ones in future.

Borgström et al [2] detected increased level of the serum immune reactive trypsin (IRT), chymotrypsin activity and of pancreatic trypsin inhibitor from the umbilical venous blood of CF homozygous newborns.

Our results show that cathepsin B and H may play a role in the faster protein degradation in patients having CF.

We are going to investigate the activity of the serum cysteine proteinases in the cases of the CF obligate heterozygotes (parents), too.

We can conclude that the values of the cathepsin B and H activities in serum may be useful as complementary biochemical data of the laboratory diagnosis of CF.

REFERENCES

1. Koch FR, Schwick HG, Störko K: Alfa-Antitrypsin Bestimmungen im Serum bei Patienten mit Mucoviscidose. *Klin Wschr* 43: 1120, 1965
2. Borgström A, Sveger T, Lindberg T, Kullander S, Svanberg L: Immunoreactive trypsin, chymotrypsin and pancreatic secretory trypsin inhibitor in cord blood from infants with cystic fibrosis. *Acta Paed Scand* 70: 619-21, 1981
3. Nadler HL, Rembelski P, Mesirow KH: Prenatal detection of cystic fibrosis. *Lancet* II: 1226-27, 1981
4. Dann LG, Blan K: Amniotic fluid arginine esterases as markers for cystic fibrosis. *Lancet* I: 619, 1982
5. Hösli P, Vogt E: Detection of cystic fibrosis homozygotes and heterozygotes with plasma. *Lancet* I: 543-545, 1979
6. Walsh-Platt M, Rao GJS, Nadler HL: Protease deficiency in plasma of patients with cystic fibrosis. *Enzyme* 24: 224-29, 1979
7. Koheil A, Corey M, Forstner G: Deficiency of serum carboxy-peptidase B-like activity in sera from patients with CF. *Clin Invest Med* 2: 99-104, 1979
8. Bird JWC, Roisen FJ, Yorke G, Lee JA, McElligott MA, Triemer DF, St. John A: Lysosomes and proteolytic enzyme activities in cultured striated muscle cells. *J Histochem Cytochem* 29: 431-9, 1981
9. Roisen FJ, Kirschke H, Colella R, Wood L, St. John A, Fekete E, Li QS, Yorke G, Bird JWC: An investigation of intracellular proteinases during differentiation of cultured muscle cells. In: *Proteinase Inhibitors*. eds Katunuma N, Umezawa H, Holzer H. Springer-Verlag, New York 1983, pp 97-109
10. Sohár I, Fekete E, Yorke G, Cosentino B, Roisen FJ, Bird JWC: Proteinase activities in normal and dystrophic chicken myoblasts in culture. *Intracell. Prot Catab* 629-631, 1985
11. Jasmin G, Tautu C, Vanasse M, Brochu P, Simoneau R: Impaired muscle differentiation in explant cultures of Duchenne Muscular Dystrophy. *Lab Invest* 50: 197-207, 1984
12. Kirschke H, Wood L, Roisen FJ, Bird JWC: Activity of lysosomal cysteine proteinase during differentiation of rat skeletal muscles. *Biochem J* 214: 871-877, 1983
13. Nakajima K, Powers JC, Ashe BM, Zimmerman M: Mapping the extended substrate binding site of cathepsin G and human leukocyte elastase. *J Biol Chem* 254: 4027-32, 1979
14. Cotic V, Babnik J, Turk V: Human muscle inhibitors of cysteine proteinases. International Symposium on Adaptive Mechanisms of Muscle, Szeged, Hungary, 2-5 July 1986. (abstr.)
15. Lenarcic B, Sali A, Kotnik M, Ritonja A, Machleidt W, Turk V: Endogenous cysteine proteinase inhibitors from human spleen. International Symposium on Adaptive Mechanisms of Muscle, Szeged, Hungary, 2-5 July 1986. (abstr.)

Mrs A. LÁSZLÓ, MD

Medical University, 6701 Szeged, P.O. Box 471