Date: 15-12-14 12:48:59

Eurjoc of Organic Chemistry - 12:48:59 Pages: **13**

Buchwald–Hartwig Reactions of Monohaloflavones

Krisztina Kónya,^[a] Dávid Pajtás,^[a] Attila Kiss-Szikszai,^[a] and Tamás Patonay^{*[a]}

Keywords: Homogeneous catalysis / Cross-coupling / Amination / Palladium / Amino acids / Oxygen heterocycles

The article describes the amination of different monobromoor monochloroflavones with primary and secondary alkylamines and aniline derivatives by Buchwald–Hartwig reaction. The influence of the phosphine ligands used is described. The use of amino acid derivatives as a nitrogen source is also demonstrated. This latter reaction allows the synthesis of unique flavone–amino-acid conjugates.

Introduction

Benzopyran derivatives with amino functionalities are a very important class of oxygen-containing heterocycles, and many of them have remarkable biological activity. Chromone and flavone alkaloids without a direct link between the aromatic/heteoaromatic ring and the amine nitrogen are natural products, and their occurrence and chemistry have been reviewed.^[1] Rohitukine, a chromone alkaloid isolated from *Disoxylum binectifarum* and *Amoora rohituka*, inhibits cyclin-dependent kinase (CDK).^[2] Its semisynthetic analogue, flavopiridol (Alvocidib, HMR 1275, L86–8275, NSC 649890) is an antineoplastic agent, inhibiting CDK1, CDK2, CDK4, CDK5, CDK6, and CDK9 kinases; it has been studied alone and in combinations in phase I and II clinical trials (Scheme 1).^[3]

The other important subclass of amine-containing benzopyran derivatives consists of chromones and flavones in which the nitrogen is directly attached to an aromatic sp² carbon. 7-Aminoflavone was found to show antibacterial effects against, among others, *M. tuberculosis* strains.^[4,5] 6-Amino-5,7-dihydroxyflavone showed outstanding activity in the low micromolar range as an α -glucosidase inhibitor.^[6]

Many aminoflavones belonging to this second subclass have shown considerable cytotoxic, kinase inhibitory, or antiproliferative effects. 4'-Aminoflavones with one or more hydroxy groups in their A-ring have shown tyrosine-kinase inhibitory effect against p56^{*lck*}, an enzyme of 56 kDa. The most active derivative was 4'-amino-6-hydroxyflavone.^[7a] 5,7,4'-Triamino-6-hydroxyflavone selectively inhibited p56^{*lck*}, while 6,4'-diamino-7-hydroxyflavone and 5,7,3'-triamino-6-hydroxyflavone were more or less active against EGFr and p60^{*v*-src.^[7b] 5,4'-Diaminoflavone, 5-amino-4'-dimethylaminoflavone, and 6-amino-4'-(dimethylamino)flavone}

one have shown considerable antiproliferative effects against the MCF-7 breast-cancer cell line.^[8] 4'-Amino-6- or 7-fluoroisoflavones showed moderate antitumor effects against MDA-MB-468, MCF-7, HT29, and HCT-116 cell lines.^[9] The aminoflavone-based peptide NSC 710464 (AFP 464)^[10] is considered to be an anticancer prodrug. It is in phase I clinical trials, and it seems to be useful against solid tumors (Scheme 1).^[11]

3-Amino-4'-methoxyflavone and 3,3'-diamino-4'-methoxyflavone showed remarkable cytotoxicity aginst L1210 leukemia cells, and their 3-(diethylaminoethyl)amino analogues showed activities similar to those of the parent 3aminoflavones.^[12] T-614, a 3-formamidochromone derivative, showed anti-inflammatory, antiarthritic, and oral antirheumatic effects.^[13] 3-(N'-Alkylureido)chromone and 3-(N'-alkylureido)-1-thiochromone selectively inhibited the v-*abl* tyrosine kinase.^[14]

However, methods for the synthesis of aminochromones and flavones suffer from limitations, particularly for those derivatives with amine substituents in the A-ring. The most common synthetic methods are based on a Claisen-Schmidt condensation or a Baker-Venkataraman rearrangement (Scheme 2). In the first case, a base-induced aldol-type condensation gives a chalcone 2, which, in turn, can be cyclised into a nitro- or acetamidoflavone 3.^[15–21] The same intermediates (i.e., 3) are accessible by aroylation followed by Baker-Venkataraman rearrangement under strongly basic conditions.^[7b,8,22-27] The desired aminoflavones (i.e., 4) can be obtained by reduction or hydrolysis. The latter transformation usually requires harsh conditions (Scheme 2). The major problem with these methods is the synthesis of the appropriate acetophenone (i.e., 1), which is limited by the regioselectivity of the nitration of acetophenone, and by the sensitivity of the R¹ substituents under the harsh conditions.

Alternatively, a previously constructed flavone moiety can also be nitrated and then submitted to reduction, but the reactivity and selectivity patterns of the nitration step are also limiting factors.^[7b,28,29] The Wessely–Moser re-

 [[]a] Department of Organic Chemistry, University of Debrecen, 4032 Debrecen, Egyetem tér 1, Hungary E-mail: patonay.tamas@science.unideb.hu http://szerves.scienc.unideb.hu

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/ejoc.201403108.

FULL PAPER

Pages: 13



2

Ô

3

Scheme 1.



arrangement is a special route to aminoflavones.^[6,28,30] The synthesis of alkyl/arylamino or disubstituted amino derivatives represents a much greater challenge. Direct alkylation suffers from a lack of selectivity, which leads to low yields.^[8,15] Alkylation of an acetamido or sulfonylamido group followed by the cleavage of the protecting group has been reported.^[9,28] Reductive alkylation,^[31,32] and direct aromatic nucleophilic substitution of an aromatic triflyloxy group under microwave conditions^[33] have also been described.

 $X = NO_2$, NHAc

ö ö

5

The synthesis of 3-aminoflavones 7 can be carried out by the reduction of 3-nitro 8^[12,16,34–37] or 3-oximino derivatives 9, or from 3-azidoflavanones 10 under basic conditions.^[38-40] Reduction of the nitro group has been carried out by catalytic hydrogenation.^[12,34,41,42] 3-Oximinoflavanones 9 gave also 3-aminoflavones 7 upon treatment with tin(II) chloride (Scheme 3).^[43,44]



 \cap

C

4

1. nitration 2. reduction

6

H₂N

reduction or hydrolysis



Sasaki, Miyake, and their coworkers published the synthesis of various 3-alkylaminoflavones from 3-bromoflavones,^[45a] 3-tosyloxyflavones, or 3-mesyloxyflavones,^[45b] but

the structural assignment of their products proved to be wrong (vide infra).

In recent decades, Buchwald-Hartwig amination of halosubstituted aromatic/heteroaromatic systems has become the method of choice for the synthesis of alkyl- and arylamino substituted arenes.^[46] Surprisingly, only sporadic examples have been published in the field of oxygen heterocycles. There are only two published examples of the preparation of amino derivatives: the synthesis of 7-amino-5hydroxyflavone started from a triflate,^[47] while 4'-aminoisoflavone was prepared from the corresponding bromide.^[12] Caddick and his coworkers reported the Buchwald-Hartwig reaction of bromo- or triflyloxy-substituted flavones under microwave activation, but they only used hexylamine as the nitrogen source.^[48] This astonishing gap in the amination of oxygen heterocycles prompted us to start a systematic investigation of the scope and limitations of this method. We present our first results in this paper.

Results and Discussion

We started our studies using the conditions of Caddick's group,^[48] but a much wider variety of primary and secondary aliphatic and aromatic amines were tested. It turned out that microwave activation has no beneficial effect; reactions using conventional thermal heating show the same efficiency. Reactions of 6-bromoflavone (11) with various amines in the presence of $Pd_2(dba)_3$ (dba = dibenzylideneacetone) as a palladium source resulted in the formation of the desired 6-aminoflavones (i.e., 12a-12p; Scheme 4, Table 1). The only limiting factor is steric hindrance in the reacting amine. Increasing the number of α -branchings clearly decreases the yield (Table 1, entries 1-4). A similar effect was also observed in the reactions of 7-bromoflavone (17; vide infra). The lower yield for the reaction with dibutylamine in comparison with the cyclic secondary amines (Table 1, entry 10 vs. entries 11-14) also supports the role of bulkiness.



Table 1. Amination of 6-bromoflavone (11)

Entry	Product	R ¹	R ²	Yield [%] ^[a]
1	12a	Bu	Н	46 ^[b]
2	12b	<i>i</i> Bu	Н	46
3	12c	sBu	Н	38
4	12d	tBu	Н	0 ^[c,d]
5	12e	Bn	Н	59
6	12f	cHex	Н	57
7	12g	2-Me-C ₆ H ₄	Н	53
8	12h	$4-Cl-C_6H_4$	Н	48
9	12i	4-MeO-C ₆ H ₄	Н	75
10	12j	Bu	Bu	18
11	12k	-(CH ₂) ₄ -		40
12	121	-(CH ₂) ₅ -		58
13	12m	-(CH ₂) ₂ O(CH ₂) ₂ -		51
14	12n	$-(CH_2)_2 Me(CH_2)_2 -$		73
15	12o	Ph	Me	31
16	12p	1,2,3,4-tetrahydroquinolin-1-yl		48

[a] Yields refer to pure isolated products. [b] *N*,*N*-Bis(flavon-6-yl)butylamine (14; 11%) was also isolated. [c] Traces of the expected product could be detected by TLC. [d] Flavone (13; 7.7%) and β diketone 15 (15%) were isolated. These by-products were also detected by TLC in several other reactions.

In all cases, flavone (13) was detected or isolated as a byproduct. Competitive reductive elimination and β-hydride elimination from the organopalladium intermediate may both take place to reform the catalyst, but the latter pathway gives a dehalogenated product, i.e., flavone (13), which results in a decrease of the yield of the aminated product. In the reaction of substrate 11 with butylamine, an interesting bis-arylated product 14 was also obtained in low (11%)yield. Obviously, here the Buchwald-Hartwig product acts as the amine in a second cross-coupling reaction. Surprisingly, no similar double Buchwald-Hartwig coupling was observed with any of the other amines tested. It is noteworthy that Caddick and his coworkers^[48] also mentioned the formation of a similar diflavonoid tertiary amine derivative starting from 4'-bromo-2'-chloro-6-methylflavone and hexylamine. In some cases, we also observed the formation of 5'-bromo-2'-hydroxydibenzoylmethane (and its tautomeric form) (15). The appearance of this by-product under the strictly water-free conditions used can be explained by



Scheme 4.



Scheme 6. Structures of the ligands used.

a concurrent ring-opening of the heterocycle: the tertbutoxide anion attacks the electrophilic C-2 atom of the flavone moiety, and the intermediate enol ether (i.e., 16) hydrolyses during aqueous work-up and column chromatography on silica (Scheme 5).

Next, we investigated the role of the phosphine ligands and their effect on the yield of the cross-coupling reaction. 6-Bromoflavone (11) was treated with butylamine, cyclohexylamine, or piperidine in the presence of various phosphines A-I (Scheme 6) keeping all the other reagents the same, but using microwave activation. The amount of the 6-(di)substituted aminoflavone product (i.e., 12a, 12f, or 12l) was determined by HPLC. The results are summarised in Table 2, and they clearly show that there is no "magic" ligand with outstanding efficiency for every model amine. The best yields were achieved using electron-rich monodentate biaryl ligands, particularly ligands **B**, **C**, and **E**.

The average HPLC yields were >45%, but there were large deviations. For example, ligand I gave good results with primary amines, but only a low HPLC yield with the secondary amine piperidine. We can conclude that many of these ligands can be used successfully, but a detailed optimisation is needed in each case.

We chose 7-bromoflavone (17) as our next substrate to allow a comparison of the reactivity of the 6- and 7-positions. We have previously demonstrated the higher reactivity of the 7-position in Heck-Mizoroki reactions.[49] Bromide 17 was treated with various amines under the same conditions as described above (Scheme 7, Table 3). The reactions were successful with all the tested acyclic and cyclic aliphatic amines as well as aniline derivatives, and the yields

Table 2. Efficiency of the amination of 6-bromoflavone (11) in the presence of different phosphine ligands.^[a]

Ligand	BuNH ₂	HPLC area [%] ^[b] cHexNH ₂	Piperidine
A	1	3	15
B	45	51	79
С	43	35	61
D	28	15	6
E	41	43	60
F	29	30	28
G	31	29	55
Н	38	27	57
I	75	43	24

[a] Reaction conditions: Pd₂(dba)₃ (5 mol-%), ligand (7.5 mol-%), NaOtBu (1.4 equiv.), amine (1.2 equiv.), dry toluene, MW (110 °C), 15 min. [b] Product ratios were determined by HPLC; Pinnacle II C18 column, 150×4.6 mm, MeOH/H₂O (gradient).

were either similar to or better than those obtained with 6bromoflavone (11). This observation was consistent with our expectations: the electronic effect of a carbonyl group in the *para* position usually speeds up palladium-catalysed cross-coupling reactions. We can assume that the rate-de-



Scheme 7.

Buchwald-Hartwig Reactions of Monohaloflavones

termining step of this amination is the oxidative addition, i.e., the insertion of palladium(0) into the C–Br bond.

Table 3. Amination of 7-bromoflavone (17).

Entry	Product	R ¹	R ²	Yield [%] ^[a]
1	18a	Bu	Н	81
2	18b	iBu	Н	79
3	18c	sBu	Н	79
4	18d	tBu	Н	28
5	18e	Bn	Н	89
6	18f	$4-Cl-C_6H_4$	Н	55
7	18g	4-MeO-C ₆ H ₄	Н	74
8	18h	Bu	Bu	16
9	18i	-(CH ₂) ₄ -		79
10	18j	-(CH ₂) ₅ -		55
11	18k	-(CH ₂) ₂ O(CH ₂) ₂ -		74
12	18 l	Ph	Me	41

[a] Yields refer to pure isolated products.

We also compared the reactions of 6- and 7-haloflavones containing different halogen atoms under our standard conditions. Consistent with our expectations, bromoflavones 11 and 17 gave much higher yields than the corresponding 6-chloro (21) and 7-chloro (22) derivatives (Table 4). This difference can be explained on the basis of the higher activation energy of the oxidative addition step of the shorter and stronger C–Cl bond, and the electronic effect of the carbonyl group in the *para* position.

Table 4. Isolated yields from the Buchwald–Hartwig reactions of 6/7-bromo- and -chloroflavones 11, 17, 21, and 22 with various amines.

	Yield [%] ^[a]				
Amine	6-Br (11)	7-Br (17)	6-Cl (21)	7-Cl (22)	
BuNH ₂	46	81	7.7	32	
$BnNH_2$	59	89	8.8	41	
Pyrrolidine	40	79	7.3	38	
4-Cl-C ₆ H ₄	48	55	6.5	36	

[a] Yields refer to pure isolated products.

Next, we tested the amination of 7-bromo-4'-methoxyflavone (19) with some model primary and secondary amines (Scheme 7). No significant differences could be observed in the yields of the coupled products. The distant substituent does not exert any significant electronic effect (see Supporting Information).

Next, we studied the possible Buchwald–Hartwig reaction of 3-bromoflavone (23) under our previously optimised conditions, since the number of methods that have been reported for the preparation of 3-alkyl/arylaminoflavones or related systems is very limited (vide supra). Unfortunately, this coupling reaction gave the expected amines (i.e., 24) in much lower yields than those obtained with the bromides studied earlier (i.e., 11, 17, and 19), particularly with secondary amines (Scheme 8, Table 5). The use of microwave activation did not change the outcome of the coupling reactions (Table 5, entry 2). Typically, reductive elimination leading to flavone (13) took place to a considerable extent. We can conclude that this method, although it seems to work, has only limited synthetic value for the preparation of mono- or alkylated/arylated 3-aminoflavone derivatives.



Scheme 8.

Pages: 13

Table 5. Synthesis of 3-aminoflavone derivatives **24** by Buchwald–Hartwig reaction.

Entry	Product	\mathbb{R}^1	\mathbb{R}^2	Yield [%][a]	Yield of 13 [%][a]
1	24a	Bu	Н	33	14
2 ^[b]	24a	Bu	Н	27	n.d.
3	24b	Hex	Η	23	n.d.
4	24c	Bn	Η	56	9
5	24d	$-(CH_2)_4-$		13	17
6	24e	-(CH ₂)5-	6	26
7	24f	4-Me-C ₆ H ₄	Н	26	n.d.

[a] Yields refer to pure isolated products; n.d.: not determined. [b] Reaction carried out under MW irradiation (110 °C, 15 min, 200 W).

Earlier both Caddick's group^[48] and Miyake's group^[45] reported the synthesis of 3-alkylaminoflavones from 3bromo-, 3-tosyloxy-, or 3-mesyloxyflavones. Caddick and his coworkers^[51] treated 3-bromoflavone (23) with hexylamine under microwave irradiation, but in the presence of Pd(OAc)₂ instead of Pd₂(dba)₃ used previously. However, our ¹H and ¹³C NMR spectroscopic data were completely different from those reported by them in their Supplementary Information. The major problem with the assignment of the right structure is that the heterocyclic product contains only quaternary carbons, which makes the ring size questionable. Miyake and his coworkers also reported the synthesis of 3-amino- or 3-(alkylamino)flavones in good or excellent yields by simply treating 3-bromo-, 3-(tosyloxy)-, or 3-(mesyloxy)flavones with ammonia or with primary amines in THF solution at room temperature, but they did not present any m.p. values or spectroscopic data.^[45] Therefore, we tested their reaction conditions by treating 3bromoflavone (23) with butylamine and hexylamine in THF without any palladium source. The isolated products had identical or similar spectral data to "3-hexylaminoflavone" reported by Caddick et al. The spectral data indicated the presence of the same groups, thus, our Buchwald–Hartwig products and the Caddick/Miyake products should be structural isomers; one of them is the expected 3-(alkvlamino)flavone, while the other is $2-[\alpha-(alkylamino)benzyl$ idene]benzofuran-3(2H)-one, a product of a ring-contraction. Significant differences could be found between the ¹H NMR data of our Caddick/Miyake products and our Buchwald-Hartwig products. The Caddick/Miyake products showed NH signals at $\delta = 10$ ppm, signals of the hydrogen in the *peri* position to the carbonyl group at $\delta = 7.84$ ppm, and NCH₂ signals at $\delta \approx 3.22$ ppm. The corresponding values for our Buchwald–Hartwig products were $\delta \approx 4.4$,

FULL PAPER



Scheme 9.

8.25, and 2.64 ppm, respectively. The ¹³C NMR signals differed less markedly. We carried out 2D NMR spectroscopic measurements (COSY and HMBC) for both of the butylsubstituted compounds to assign the signals of the carbon and hydrogen atoms (see Supporting Information). HMBC measurements showed a strong cross-peak between the 1''-H (NCH₂) and C_a signals in the spectrum of the Caddick/ Miyake product. On the other hand, the cross-peak between the 1''-H (NCH₂) and C-3 signals in our Buchwald– Hartwig product clearly proved that Caddick/Miyake product should be 2-[α -(butylamino)benzylidene]benzofuran-3(2H)-one (**25a**).

The formation of the ring-contracted products (i.e., **25a** and **25b**) can be explained in terms of attack of the amine nucleophile on the C-2 atom of the oxygen heterocycle (Scheme 9).

To obtain additional and independent proof for the structure of 3-(butylamino)flavone (**24a**) obtained in the Buchwald–Hartwig procedure, and to rule out isomeric structure **25a**, we alkylated the parent 3-aminoflavone (i.e., **26**) with butyl bromide in aqueous solution in the presence of potassium carbonate under microwave irradiation using the protocol developed for 3-(N,N-alkylamino)flavones.^[50] The resulting product showed the same spectroscopic data as our Buchwald–Hartwig product, thus proving that the structures of Caddick's and Miyake's compounds were erroneously assigned.

We also investigated the synthesis of *N*-unsubstituted aminoflavones. The most frequently used ammonia equivalents in Buchwald–Hartwig cross-coupling are benzophenone imine^[51] and lithium silyl amides.^[52] Until now, only one example has been published in the field of flavones.^[47] We successfully prepared both 6-aminoflavone (**28a**) and 7-aminoflavone (**28b**) via the corresponding imine intermediates (i.e., **27a** and **27b**) in 48 and 38% overall yields, respectively (Scheme 10). Unfortunately, the reaction of 3-bromo-flavone (**23**) failed to give the desired 3-aminoflavone (**27**).

Next, we tested the cross-coupling reaction with α -amino acid methyl esters as the nitrogen source. This reaction could lead to hitherto almost unknown amino-acid–flavone hybrids of potential biological importance. To the best of our knowledge, only two examples of such derivatives have



Scheme 10.

been published to date, and neither of them was obtained by Buchwald-Hartwig (or Ullmann) type coupling. Korean authors^[53] prepared a conjugate by alkylation of a preformed 6-aminoflavone (28a) with an α -bromo- α -phenvlacetic acid thioester, while Georg et al.^[54] coupled 8amino-4'-chloro-5-hydroxyflavone with N-Boc-protected amino acids (Boc = *tert*-butoxycarbonyl). First, we treated 6-bromoflavone (11) with L-phenylalanine methyl ester hydrochloride (29a), but the desired coupled product could not be obtained under the previously used conditions. After extensive optimisation, we found a set of conditions: Pd₂(dba)₃ should be replaced by palladium(II) acetate, and the original base sodium tert-butoxide should be replaced by cesium carbonate. However, the expected product (i.e., 30a) was only obtained in a very low yield (5%), and, in addition, the regioisomeric 7-{[2-phenyl-1-(methoxycarbonyl)ethyl]amino}flavone (31a) was also isolated in 6%yield (Scheme 11). These structures were assigned on the basis of their NMR spectra, and the constitution of compound 31a was also supported by its independent synthesis (vide infra).

A possible explanation for this astonishing observation may be the so-called "halogen dance"^[55] under the strongly basic conditions. According to this suggested mechanism, a deprotonation at the 7-position of the starting 6-bromoflavone (**11**) takes place, followed by a bromine exchange between the resulting carbanion intermediate and a second Date: 15-12-14 12:48:59

Pages: 13



Scheme 11.

molecule of starting material to give 6,7-dibromoflavone. The relative ease of this deprotonation may be explained by the higher stability of this carbanion. The 6,7-dibromoflavone intermediate may react with the above-mentioned carbanion to yield 7-bromoflavone (17). These hypothetical steps are consistent with the generally accepted mechanism of the "halogen dance". However, it should be emphasised that the "halogen dance" usually needs more basic conditions [LDA (dithium diisopropylamide), BuLi, NaNH₂, ArNHK, etc.]. In addition, to the best of our knowledge, this would be the first example of such a transformation to be observed in a Buchwald–Hartwig reaction.

Applying the same conditions to the reaction of 7bromoflavone (17) and various amino acid methyl ester hydrochlorides 29, the expected products (i.e., 31a-31e) were obtained (Scheme 11, Table 6), and no 6-substituted products were obtained or detected. The higher reactivity of 7-bromoflavone (17) caused by electronic effects (vide supra) leads to a fast reaction, and so excludes the "halogen dance". These results further demonstrate the synthetic utility of the Buchwald-Hartwig reaction in the field of oxygen heterocycles. On the other hand, the data summarised in Table 6 clearly show that the polarity of the amino acid has a decisive role in the efficiency of the reaction. The coupling with L-aspartic acid resulted in a low yield, and no product was obtained with L-serine or L-tryptophan. The lack of any reaction may be explained by either the low solubility of the amino acid derivative in xylene, or by ligation of the palladium(0)/palladium(II) ion to the heteroatom of the reagent.

Table 6. Synthesis of flavone-amino-acid hybrids **31** by Buchwald-Hartwig reaction.

Entry	Amino acid	Product	Yield [%]	ee [%]
1	L-Phe	31a	64	6
2	D-Phe	31a	59	10
3	L-Ala	31b	30	n.d.
4	L-Leu	31c	56	6
5	L-Met	31d	40	1
6	L-Asp	31e	11	2
7	L-Ser	31f	0	_
8	L-Trp	31g	0	-

Another drawback of coupling with amino acid esters, at least at the moment, is racemisation (Table 6). This can be attributed to a deprotonation–reprotonation sequence at the stereogenic centre of the amino acid unit.

Conclusions

In conclusion, we have demonstrated that the Buchwald– Hartwig amination of bromoflavones with halo substituents on their aromatic rings is a useful tool for the synthesis of amine derivatives that are hardly available by previously reported methods. We have pointed out that the previously published structures of some derivatives obtained from 3halo- or sulfonyloxyflavones with primary amines were erroneously assigned, and that in fact, ring-contracted products were formed. Palladium-catalysed C–N bond formation also proved to be an efficient tool in the synthesis of flavone–amino-acid hybrids, although the coupling takes place accompanied by significant racemisation of the stereogenic centre. Experiments aimed at avoiding this racemisation and extending the number of nitrogen sources are in progress, and the results will be published soon.

Experimental Section

General Remarks: Column chromatography was carried out on silica gel (Merck 60, 70–230 mesh). Thin-layer chromatography was carried out on silica gel plates 60 F_{254} (Merck, 0.2 mm). In phosphine-ligand optimisation studies, product ratios were determined by HPLC using a Pinnacle II column (C18, 150×4.6 mm, 5 µm; MeOH/H₂O, gradient: from 60 to 90% MeOH in 20 min; flow: 0.9 mL/min, 280 nm). Chiral HPLC was carried out using a Chiralpak IC column (250×4.6 mm) with a Jasco HPLC system: Jasco PU-980 HPLC pump, Jasco MD-910 multiwavelength detector. The purity of the compounds was established by GC–MS (Agilent 7890, Agilent 5975 MS detector) with positive EI at 70 eV. NMR spectra were recorded with a Bruker AM 360 (360.13 MHz for ¹H, 90.03 MHz for ¹³C) spectrometer. Chemical shifts (δ) were calibrated using the CHCl₃ signals (δ = 7.26 ppm) for ¹H and (δ =

FULL PAPER

77.00 ppm) for ¹³C spectra. Melting points were determined by using Büchi B-540 equipment. Elemental analysis (C, H) was conducted with an Elementar Vario MicroCube instrument. IR spectra were measured as KBr discs with a Jasco FTIR 4100A instrument. Microwave-assisted reactions were carried out using a CEM-Discover Focused microwave synthesis system (2450 MHz).

6-Bromoflavone (11),^[56] 7-bromoflavone (17),^[56] and amino acid methyl ester hydrochlorides^[57] were synthesised according to literature procedures.

Buchwald–Hartwig Cross-Coupling by Thermal Heating: Haloflavone (0.66 mmol), NaOtBu (88 mg, 0.92 mmol), BINAP [2,2'bis(diphenylphosphino)-1,1'-binaphthyl; 32 mg, 0.050 mmol], and amine (0.80 mmol) were mixed in dry toluene (6 mL) in a dried flask under nitrogen, and $Pd_2(dba)_3$ (30 mg, 0.032 mmol) was added. The reaction mixture was stirred at reflux for 3 h in an oil bath. The crude reaction mixture was applied directly to a silica gel column to give the pure cross-coupled product.

Buchwald–Hartwig Cross-Coupling by Microwave Activation: 6-Bromoflavone (0.33 mmol), NaOtBu (44 mg, 0.46 mmol), BINAP (15 mg, 0.025 mmol), and amine (0.80 mmol) were mixed in dry toluene (3 mL) in a sealed vial under nitrogen, and $Pd_2(dba)_3$ (15 mg, 0.016 mmol) was added. The reaction mixture was stirred for 15 min at 110 °C (200 W). The crude reaction mixture was applied directly to a silica gel column to give the pure cross-coupled product.

Only representative examples are given here. Characterisation data for the vast majority of the products are presented in the Supporting Information.

6-(Butylamino)flavone (12a): Eluent: toluene/ethyl acetate, 4:1; yield 88 mg (46%) from **11**, 15 mg (7.7%) from **21**; m.p. 124.5–126.5 °C. ¹H NMR (CDCl₃): δ = 7.89 (m, 2 H, 2'-H, 6'-H), 7.50 (m, 3 H, 3'-H, 5'-H, 4'-H), 7.39 (d, *J* = 8.6 Hz, 1 H, 8-H), 7.24 (d, *J* = 2.5 Hz, 1 H, 5-H), 6.98 (dd, *J* = 2.9, 9.0 Hz, 1 H, 7-H), 6.78 (s, 1 H, 3-H), 3.87 (s, 1 H, NH), 3.19 (t, *J* = 7.2 Hz, 2 H, 1''-H), 1.64 (m, 2 H, 2''-H), 1.45 (m, 2 H, 3''-H), 0.97 (t, *J* = 7.6 Hz, 3 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 178.6 (C-4), 162.6 (C-2), 149.3 (C-8a), 146.2 (C-6), 132.2 (C-1'), 131.1 (C-4'), 128.9 (C-3', C-5'), 126.1 (C-2', C-6'), 124.8 (C-4a), 121.2 (C-7), 118.8 (C-8), 106.7 (C-5), 103.5 (C-3), 43.9 (C-1''), 31.4 (C-2''), 20.2 (C-3''), 13.9 (C-4'') ppm. IR (KBr): \hat{v} = 3352, 2949, 2926, 2865, 1624, 1582, 1570, 1490, 1450, 1211, 912, 818, 770 cm⁻¹. MS: *m*/*z* = 293 [M]⁺, 250 (100%). C₁₉H₁₉NO₂ (293.36): calcd. C 77.79, H 6.53, N 4.77; found C 77.75, H 6.52, N 4.75.

N,*N*-**Bis(2-phenyl-4-oxo-***4H*-**chromen-6-yl)butylamine (14):** Eluent: toluene/ethyl acetate, 4:1; yield 20 mg (11%); m.p. 218–220 °C. ¹H NMR (CDCl₃): δ = 7.91 (m, 4 H, 4 H), 7.81 (d, *J* = 2.7 Hz, 2 H), 7.53 (dd, *J* = 4.8, 1.1 Hz, 5 H), 7.49 (d, *J* = 9.1 Hz, 3 H), 7.35 (dd, *J* = 9.0, 2.8 Hz, 2 H), 6.81 (s, 2 H), 3.91–3.75 (m, 2 H), 1.77–1.61 (m, 2 H), 1.42 (dd, *J* = 15.0, 7.5 Hz, 2 H), 0.95 (t, *J* = 7.3 Hz, 3 H) ppm. ¹³C NMR (CDCl₃): δ = 178.1 (C-4), 163.1 (C-2), 151.6 (C-8a), 145.1 (C-6), 131.8 (C-1'), 131.4 (C-4'), 128.9 (C-3', C-5'), 127.9 (C-7), 126.2 (C-2', C-6'), 124.9 (C-4a), 12.2 (C-7), 119.3 (C-8), 115.0 (C-5), 106.9 (C-3), 52.7 (C-1''), 29.4 (C-2''), 20.2 (C-3''), 13.9 (C-4'') ppm. IR (KBr): \tilde{v} = 3446, 3064, 2956, 2925, 2854, 1632, 1566, 1495, 1480, 1451, 1356, 1259, 910, 818, 770 cm⁻¹. MS: *m*/*z* = 326, 213 (100%), 137, 105, 77, 55. C₃₄H₂₇NO₄ (513.59): calcd. C 79.51, H 5.30, N 2.73; found C 79.70, H 5.35, N 2.75.

6-[(2-Methylphenyl)amino]flavone (12g): Eluent: toluene/ethyl acetate, 4:1; yield 114 mg (53%); m.p. 220–223 °C. ¹H NMR (CDCl₃ + [D₆]DMSO): δ = 7.92 (m, 2 H, 2'-H, 6'-H), 7.61 (d, *J* = 2.5 Hz, 1

H, 5-H), 7.53 (m, 3 H, 3'-H, 5'-H, 4'-H), 7.48 (d, J = 9.0 Hz, 1 H, 8-H), 7.31 (dd, J = 2.5, 9.7 Hz, 1 H, 7-H), 7.25 (m, 2 H, 3''-H, 6''-H), 7.18 (m, 1 H, 5''-H), 7.01 (m, 1 H, 4''-H), 6.78 (s, 1 H, 3-H), 5.88 (s, 1 H, NH), 2.27 (s, 3 H, 2-CH₃) ppm. ¹³C NMR (CDCl₃ + [D₆]DMSO): $\delta = 177.1$ (C-4), 161.9 (C-2), 149.2 (C-8a), 142.9 (C-1''), 140.1 (C-6), 131.3 (C-2''), 130.8 (C-3''), 130.5 (C-4'), 130.3 (C-1'), 128.5 (C-3', C-5'), 126.1 (C-5''), 125.6 (C-2', C-6'), 123.9 (C-4a), 122.8 (C-4''), 122.6 (C-6''), 120.7 (C-7), 118.4 (C-8), 106.9 (C-5), 105.7 (C-3), 17.6 (2''-CH₃) ppm. IR (KBr): $\tilde{v} = 3317$, 3113, 3063, 2939, 1641, 1619, 1576, 1534, 1483, 1449, 1365, 1318, 1138, 914, 832, 770, 755, 690 cm⁻¹. MS: m/z = 326 [M]⁺⁺ (100%), 168. C₂₂H₁₇NO₂ (327.38): calcd. C 80.71, H 5.23, N 4.28; found C 80.69, H 5.20, N 4.26.

6-(Dibutylamino)flavone (12j): Oil, eluent: toluene/ethyl acetate, 8:1; yield 42 mg (18%). ¹H NMR (CDCl₃): δ = 7.91 (m, 2 H, 2'-H, 6'-H), 7.51 (m, 3 H, 3'-H, 5'-H, 4'-H), 7.43 (d, J = 9.0 Hz, 1 H, 8-H), 7.29 (d, J = 2.9 Hz, 1 H, 5-H), 7.06 (dd, J = 2.9, 9.0 Hz, 1 H, 7-H), 6.78 (s, 1 H, 3-H), 3.35 (t, J = 7.6 Hz, 4 H, 1''-H), 1.60 (m, 4 H, 2''-H), 1.38 (m, 4 H, 3''-H), 0.97 (t, J = 7.2 Hz, 6 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 178.8 (C-4), 162.5 (C-2), 148.2 (C-8a), 146.0 (C-6), 132.3 (C-1'), 131.1 (C-4'), 128.9 (C-3', C-5'), 126.1 (C-2', C-6'), 124.7 (C-4a), 119.2 (C-7), 118.7 (C-8), 106.4 (C-5), 104.0 (C-3), 50.9 (C-1''), 29.3 (C-2''), 20.3 (C-3''), 14.0 (C-4'') ppm. IR (KBr): \tilde{v} = 3060, 2925, 2928, 2870, 1639, 1614, 1566, 1500, 1453, 1363, 911, 771, 689 cm⁻¹. MS: m/z = 349 [M]⁺⁻, 306 (100%). C₂₃H₂₇NO₂ (349.47): calcd. C 79.05, H 7.79, N 4.01; found C 79.07, H 7.81, N 4.02.

6-(Phenyl-methylamino)flavone (120): Eluent: hexane/ethyl acetate, 4:1; yield 67 mg (31%); m.p. 164.5–165.5 °C. ¹H NMR (CDCl₃): δ = 7.91 (m, 2 H, 2'-H, 6'-H), 7.67 (d, J = 2.5 Hz, 1 H, 5-H), 7.52 (m, 3 H, 3'-H, 5'-H, 4'-H), 7.36 (m, 3 H, 8-H, 3''-H, 5''-H), 7.28 (dd, J = 2.9, 9.0 Hz, 1 H, 7-H), 7.10 (m, 3 H, 2''-H, 6''-H, 4''-H), 6.81 (s, 1 H, 3-H), 3.39 (s, 3 H, NCH₃) ppm. ¹³C NMR (CDCl₃): δ = 178.3 (C-4), 162.8 (C-2), 150.7 (C-1''), 148.4 (C-8a), 146.5 (C-6), 131.9 (C-1'), 131.3 (C-4'), 129.5 (C-3'', C-5''), 128.9 (C-3', C-5'), 126.1 (C-2', C-6'), 125.5 (C-7), 124.6 (C-4a), 123.4 (C-4''), 123.0 (C-2'', C-6''), 118.6 (C-5), 110.6 (C-8), 106.8 (C-3), 40.7 (NCH₃) ppm. IR (KBr): \tilde{v} = 3448, 3037, 3002, 2951, 2890, 2815, 1620, 1590, 1580, 1566, 1482, 1457, 1450, 1362, 1141, 1134, 1045, 1029, 912, 832, 821, 770, 707 cm⁻¹. MS: *m/z* = 327 [M]⁺⁺, 326 (100%). C₂₂H₁₇NO₂ (327.38): calcd. C 80.71, H 5.23, N 4.28; found C 80.69, H 5.20, N 4.28.

7-(Butylamino)flavone (18a): Eluent: toluene/ethyl acetate, 8:1; yield 157 mg (81%) from **17**, 63 mg (32%) from **22**; m.p. 163.5–164.5 °C. ¹H NMR (CDCl₃): δ = 7.96 (d, *J* = 8.6 Hz, 1 H, 5-H), 7.88 (m, 2 H, 2'-H, 6'-H), 7.49 (m, 3 H, 3'-H, 5'-H, 4'-H), 6.69 (s, 1 H, 3-H), 6.62 (d, *J* = 8.6 Hz, 1 H, 6-H), 6.52 (s, 1 H, 8-H), 4.50 (s, 1 H, NH), 3.21 (q, *J* = 6.8 Hz, 2 H, 1''-H), 1.65 (m, 2 H, 2''-H), 1.46 (m, 2 H, 3''-H), 0.98 (t, *J* = 7.6 Hz, 3 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.6 (C-4), 162.0 (C-2), 158.8 (C-8a), 153.2 (C-7), 132.1 (C-1'), 104.0 (C-4'), 128.8 (C-3', C-5'), 126.5 (C-5), 125.9 (C-2', C-6'), 114.3 (C-4a), 112.9 (C-6), 107.2 (C-3), 96.2 (C-8), 43.1 (C-1''), 31.1 (C-2''), 20.2 (C-3''), 13.8 (C-4'') ppm. IR (KBr): \tilde{v} = 3316, 3139, 3069, 3034, 2948, 2927, 2865, 1632, 1586, 1566, 1503, 1448, 1378, 1248, 1095, 906, 824, 764, 670 cm⁻¹. MS: *m*/*z* = 293 [M]⁺, 250 (100%). C₁₉H₁₉NO₂ (293.36): calcd. C 77.79, H 6.53, N 4.77; found C 77.76, H 6.55, N 4.78.

7-[(4-Chlorophenyl)amino]flavone (18f): Eluent: toluene/ethyl acetate, 4:1; yield 126 mg (55%) from **17**, 83 mg (36%) from **22**; m.p. 273.5–276.5 °C. ¹H NMR (CDCl₃ + [D₆]DMSO): δ = 8.36 (s, 1 H, NH), 7.99 (d, *J* = 8.6 Hz, 1 H, 5-H), 7.90 (m, 2 H, 2'-H, 6'-H), 7.53 (m, 3 H, 3'-H, 5'-H, 4'-H), 7.33 (d, *J* = 8.6 Hz, 2 H, 3''-H,

5''-H), 7.23 (d, J = 8.6 Hz, 2 H, 2''-H, 6''-H), 7.09 (s, 1 H, 8-H), 7.02 (d, J = 8.6 Hz, 1 H, 6-H), 6.70 (s, 1 H, 3-H) ppm. ¹³C NMR (CDCl₃ + [D₆]DMSO): $\delta = 176.7$ (C-4), 161.8 (C-2), 157.6 (C-8a), 149.1 (C-7), 139.0 (C-1''), 131.3 (C-1'), 130.6 (C-4'), 128.7 (C-2', C-6'), 128.3 (C-3'', C-5''), 126.9 (C-4''), 126.0 (C-5), 125.4 (C-2', C-6'), 121.3 (C-2'', C-6''), 115.5 (C-4a), 114.2 (C-6), 106.6 (C-3), 99.2 (C-8) ppm. IR (KBr): $\tilde{v} = 3274$, 3188, 3100, 1648, 1627, 1585, 1534, 1492, 1449, 1438, 1374, 1092, 909, 813, 768, 676 cm⁻¹. MS: m/z = 347 [M]⁺⁻ (100%), 319. C₂₁H₁₄ClNO₂ (347.80): calcd. C 72.52, H 4.06, Cl 10.19, N 4.03; found C 72.55, H 4.10, Cl 10.21, N 4.01.

7-Morpholinoflavone (18k): Eluent: toluene/ethyl acetate, 4:1; yield 150 mg (74%); m.p. 208–211.5 °C. ¹H NMR (CDCl₃): δ = 8.07 (d, J = 9.0 Hz, 1 H, 5-H), 7.89 (m, 2 H, 2'-H, 6'-H), 7.51 (m, 3 H, 3'-H, 5'-H, 4'-H), 6.96 (dd, J = 2.2, 9.0 Hz, 1 H, 6-H), 6.83 (s, J = 2.2 Hz, 1 H, 8-H), 6.73 (s, 1 H, 3-H), 3.89 (m, 4 H, 3''-H, 5''-H), 3.36 (m, 4 H, 2''-H, 6''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.7 (C-4), 162.6 (C-2), 158.1 (C-8a), 155.0 (C-7), 132.0 (C-1'), 131.2 (C-4'), 128.9 (C-3', C-5'), 126.6 (C-5), 126.0 (C-2', C-6'), 115.9 (C-4a), 112.9 (C-6), 107.4 (C-3), 100.3 (C-8), 66.4 (C-3'', C-5''), 47.6 (C-2'', C-6'') ppm. IR (KBr): \tilde{v} = 3432, 3060, 2963, 2855, 1632, 1592, 1573, 1511, 1494, 1448, 1383, 1239, 1124, 819, 772 cm⁻¹. MS: m/z = 307 [M]⁺⁺ (100%), 249, 222, 147, 91. C₁₉H₁₇NO₃ (307.35): calcd. C 74.25, H 5.58, N 4.56; found C 74.24, H 5.60, N 4.54.

3-Hydroxy-1-(5-bromo-2-hydroxyphenyl)-3-phenylprop-2-en-1-one (15):



Eluent: toluene/ethyl acetate, 4:1; yield 17 mg (7.7%); m.p. 112–115 °C. This compound was obtained as a by-product from the reaction of *tert*-butylamine and 6-bromoflavone. ¹H NMR (CDCl₃): δ = 15.48 (s, 1 H, 2''-OH), 12.06 (s, 1 H, 3-OH), 7.96 (d, J = 7.2 Hz, 2 H, 2'-H, 6'-H), 7.85 (s, 1 H, 6''-H), 7.57 (d, J = 7.2 Hz, 1 H, 4''-H), 7.52 (m, 3 H, 3'-H, 5'-H, 4'-H), 6.90 (d, J = 9.0 Hz, 1 H, 3''-H), 6.75 (s, 1 H, 2-H) ppm. ¹³C NMR (CDCl₃): δ = 194.2 (C-1), 178.4 (C-3), 161.3 (C-2''), 138.3 (C-4''), 133.2 (C-1'), 132.8 (C-6''), 120.6 (C-4'), 128.8 (C-2', C-6'), 127.0 (C-3', C-5'), 120.8 (C-3''), 120.3 (C-1''), 110.7 (C-5''), 92.1 (C-2) ppm. IR (KBr): \tilde{v} = 1607, 1567, 1469, 1289, 1190, 913, 744 cm⁻¹.

7-(Butylamino)-4'-methoxyflavone (20a): Eluent: toluene/ethyl acetate, 1:1; yield 178 mg (84%); m.p. 173.5–175.5 °C. ¹H NMR $(CDCl_3)$: $\delta = 7.95$ (d, J = 9.0 Hz, 1 H, 5-H), 7.83 (d, J = 8.6 Hz, 2 H, 2'-H, 6'-H), 6.99 (d, J = 8.6 Hz, 2 H, 3'-H, 5'-H), 6.59 (m, 2 H, 3-H, 6-H), 6.50 (d, J = 1.8 Hz, 1 H, 8-H), 4.42 (s, 1 H, NH), 3.87 (s, 3 H, 4'-OCH₃), 3.20 (m, 2 H, 1''-H), 1.66 (m, 2 H, 2''-H), 1.46 (m, 2 H, 3''-H), 0.98 (t, J = 7.6 Hz, 3 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.7 (C-4), 162.1 (C-2), 161.9 (C-4'), 158.7 (C-8a), 153.0 (C-7), 127.6 (C-2', C-6'), 126.5 (C-5), 124.5 (C-1'), 114.4 (C-4a), 114.2 (C-3', C-5'), 112.7 (C-6), 105.9 (C-3), 96.3 (C-8), 55.4 (4'-OCH₃), 43.2 (C-1''), 31.2 (C-2''), 20.2 (C-3''), 13.81 (C-4'') ppm. IR (KBr): $\tilde{v} = 3302, 2950, 2929, 2860, 1624, 1608,$ 1584, 1566, 1537, 1508, 1423, 1376, 1244, 1176, 1095, 909, 822, 597 cm⁻¹. MS: $m/z = 323 \text{ [M]}^+$, 280 (100%), 148. C₂₀H₂₁NO₃ (323.39): calcd. C 74.28, H 6.55, N 4.33; found C 74.25, H 6.58, N 4.35.

3-(Butylamino)flavone (24a): Yellow oil, eluent: hexane/ethyl acetate, 3:1; yield 64 mg (33%). ¹H NMR (CDCl₃): δ = 8.24 (d, J = 7.9 Hz, 1 H, 5-H), 7.96 (d, J = 7.2 Hz, 2 H, 2'-H, 6'-H), 7.60 (t, J

 $= 7.2 \text{ Hz}, 1 \text{ H}, 7-\text{H}), 7.40-7.51 \text{ (m, 4 H, 6-H, 8-H, 3'-H, 5'-H)}, 7.36 \text{ (m, 1 H, 4'-H)}, 4.41 \text{ (s, 1 H, NH)}, 2.64 \text{ (t, } J = 6.8 \text{ Hz}, 2 \text{ H}, 1''-\text{H}), 1.36-1.42 \text{ (m, 2 H, 2''-H)}, 1.21-1.29 \text{ (m, 2 H, 3''-H)}, 0.79 \text{ (t, } J = 7.5 \text{ Hz}, 3 \text{ H}, 4''-\text{H}) \text{ ppm.} \ ^{13}\text{C} \text{ NMR} \text{ (CDCl}_3): \delta = 175.3 \text{ (C-4)}, 155.5 \text{ (C-8a)}, 147.7 \text{ (C-2)}, 133.5 \text{ (C-4a)}, 132.9 \text{ (C-7)}, 131.6 \text{ (C-1)}, 129.6 \text{ (C-5)}, 128.3 \text{ (C-2', C-6')}, 128.1 \text{ (C-3', C-5')}, 125.8 \text{ (C-4')}, 132.9 \text{ (C-4')}, 132.9 \text{ (C-4')}, 132.9 \text{ (C-4')}, 132.8 \text{ (C-4')}, 132.9 \text{ (C-4')}, 132.9 \text{ (C-4')}, 132.8 \text{ (C$

1), 129.6 (C-5), 128.3 (C-2', C-6'), 128.1 (C-3', C-5'), 125.8 (C-4'), 124.0 (C-6), 120.9 (C-3), 117.9 (C-8), 47.0 (C-1''), 32.3 (C-2''), 19.9 (C-3''), 13.8 (C-4'') ppm. IR (KBr): $\tilde{v} = 3326, 2957, 2928, 2871, 2358, 1613, 1465, 1195, 757 cm^{-1}$. MS: m/z = 293 [M]⁺, 250 (100%). C₂₄H₁₉NO₂ (353.42): calcd. C 81.56, H 5.42, N 3.96; found C 81.60, H 5.45, N 3.93.

3-N-Pyrrolidinoflavone (24d): Oil, eluent: hexane/ethyl acetate, 3:1; yield 26 mg (13%). ¹H NMR (CDCl₃): δ = 8.22 (d, *J* = 7.9 Hz, 1 H, 5-H), 8.01 (d, *J* = 7.2 Hz, 2 H, 2'-H, 6'-H), 7.64 (m, 1 H, 7-H), 7.47–7.50 (m, 4 H, 6-H, 8-H, 3'-H, 5'-H), 7.37 (m, 1 H, 4'-H), 3.16 (m, 4 H, 2''-H, 5''-H), 1.94 (m, 4 H, 3''-H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.9 (C-4), 161.9 (C-8a), 155.4 (C-2), 133.1 (C-7), 132.9 (C-1'), 130.4 (C-4a), 130.0 (C-5), 128.7 (C-2', C-6'), 128.0 (C-3', C-5'), 125.6 (C-4'), 124.5 (C-6), 124.3 (C-3), 117.9 (C-8), 50.6 (C-2'', C-5''), 26.3 (C-3'', C-4'') ppm. IR (KBr): \tilde{v} = 3437, 2929, 2851, 1615, 1602, 1466, 1376, 1307, 1198, 754, 689 cm⁻¹. MS: m/z = 291 [M]⁺⁺ (100%), 248, 222, 194, 165, 120. C₁₉H₁₇NO₂ (291.35): calcd. C 78.33, H 5.88, N 4.81; found C 78.37, H 5.90, N 4.80.

Buchwald–Hartwig Cross-Coupling of 3-Bromoflavone (23) by Microwave Activation: A mixture of 3-bromoflavone (23; 100 mg, 0.330 mmol), *n*-butylamine (40 μ L, 0.400 mmol), NaOtBu (44 mg, 0.46 mmol), and BINAP (15 mg, 0.025 mmol) in toluene (3 mL) in a capped vial (10 mL) was irradiated for 15 min (200 W, 110 °C). Then the mixture was cooled, water was added (10 mL), and the mixture was extracted with ethyl acetate (2 × 25 mL). The combined organic layers were washed with water, and dried (magnesium sulfate), and the solvent was evaporated in vacuo. The crude product was purified by column chromatography (hexane/ethyl acetate, 4:1) to give **24a** (26 mg, 27%) as a yellow oil.

Alkylation of 3-Aminoflavone (26) by Microwave Activation: A mixture of 3-aminoflavone (26; 474 mg, 2.000 mmol), butyl bromide (603 mg, 4.400 mmol), and potassium carbonate (608 mg, 4.400 mmol) in water (4 mL) was irradiated for 60 min (100 W, 120 °C) in a capped vial (10 mL). Then, the mixture was cooled and extracted with ethyl acetate (2×25 mL). The combined organic layers were washed with water, and dried (magnesium sulfate), and the solvent was evaporated in vacuo. The crude product was purified by column chromatography (hexane/ethyl acetate, 4:1) to give 24a (114 mg, 19%) as a yellow oil.

Ring Contraction: A mixture of 3-bromoflavone (100 mg, 0.33 mmol) and amine (0.825 mmol, 2.5 equiv.) in THF was stirred for 1 d at ambient temperature according to the literature procedure.^[45] Then the solvent was evaporated, and the crude reaction mixture was submitted directly to a silica gel column to give the pure product.

2-[*a*-(**Butylamino**)**benzylidene**]**benzofuran-3**(*2H*)-**one** (**25a**): Yellow oil, eluent: hexane/ethyl acetate, 5:1; yield 71 mg (74%). ¹H NMR (CDCl₃): δ = 10.02 (s, 1 H, NH), 7.84 (d, *J* = 7.6 Hz, 1 H, 5-H), 7.50 (m, 5 H, 2'-H, 3'-H, 4'-H, 5'-H, 6'-H), 7.42 (t, *J* = 7.8 Hz, 1 H, 7-H), 7.13 (m, 2 H, 6-H, 8-H), 3.23 (t, *J* = 6.8 Hz, 2 H, 1''-H), 1.56 (m, 2 H, 2''-H), 1.38 (m, 2 H, 3''-H), 0.87 (t, *J* = 7.3 Hz, 3 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.5, 160.8, 152.2, 132.0, 131.7, 130.2, 129.7, 128.9, 128.6, 124.7, 122.8, 121.6, 112.5, 44.2, 32.7, 29.6, 19.7, 13.6 ppm. IR (KBr): \tilde{v} = 3368, 3065, 2960, 2932, 2873, 1702, 1632, 1603, 1458, 1381, 1250, 755, 713 cm⁻¹. MS: *m*/*z*

FULL PAPER

= 293 [M]⁺, 250 (100%). $C_{19}H_{19}NO_2$ (293.36): calcd. C 77.79, H 6.53, N 4.77; found C 77.82, H 6.55, N 4.80.

General Procedure for the Synthesis of 6- and 7-(Diphenylmethyleneamino)flavones (27a and 27b): A mixture of 6-bromo- (11) or 7-bromoflavone (17; 1.505 g, 5.00 mmol), BINAP (235 mg, 0.375 mmol), benzophenone imine (2; 1.10 mL, 6.00 mmol), palladium acetate (55 mg, 0.250 mmol, 5 mol-%), and cesium carbonate (2.120 g, 6.50 mmol) in xylene (30 mL) was stirred at reflux overnight under a nitrogen atmosphere. The cooled mixture was poured into water (100 mL), then extracted with ethyl acetate (3×25 mL). The combined organic phases were dried with magnesium sulfate, then the solvents were removed in vacuo. The solid residue was triturated with a mixture of hexane (20 mL) and acetone (1 mL) to give pure 27a or 27b as a pale yellow solid.

7-(Diphenylmethylene)iminolflavone (27b): Yield 73%; m.p. 212–213 °C. ¹H NMR (CDCl₃): δ = 7.99 (d, *J* = 8.6 Hz, 1 H, 5-H), 7.85 (m, 2 H, Ph), 7.77 (m, 2 H, Ph), 7.51 (m, 6 H, Ph), 7.45 (d, *J* = 8.6 Hz, 1 H, 6-H), 7.29 (m, 3 H, Ph), 7.17 (m, 2 H, Ph), 6.90 (s, 1 H, 8-H), 6.74 (s, 1 H, 3-H) ppm. ¹³C NMR (CDCl₃): δ = 178.0 (C-4), 169.5 (C=N), 162.9 (C-2), 156.8 (C-8a), 138.6 (C-7), 135.2, 137.5 (C-1''), 131.8 (C-1'), 132.4, 131.4, 130.0, 129.8, 129.5, 129.1, 128.3 (Ph), 126.1 (C-7), 119.4 (C-4a), 119.0 (C-5), 108.5, 107.5 (C-3, C-8) ppm. IR (KBr): \tilde{v} = 3455, 3068, 1640, 1617, 1458, 1358, 1289, 1024, 910, 773, 697 cm⁻¹. MS: *m*/*z* = 401 [M]⁺⁻, 324, 297, 221, 165 (100%), 139, 102, 77. C₂₈H₁₉NO₂ (401.46): calcd. C 83.77, H 4.77; found C 83.80, H 4.75. C₂₈H₁₉NO₂ (401.46): calcd. C 83.77, H 4.77, N 3.49; found C 83.80, H 4.75, N 3.54.

General Procedure for the Synthesis of 6- and 7-Aminoflavones (28a and 28b): [(Diphenylmethylene)imino]flavone 27a or 27b (1.466 g, 3.651 mmol) was dissolved in THF (14 mL), and HCl (6 N aq.; 9.2 mL, 18.0 mmol, 5 equiv.) was added. After 10 min, dichloromethane (25 mL) and water (50 mL) were added, and then the mixture was stirred at room temperature overnight. Next, dichloromethane (10 mL) was added, upon which the reaction mixture became clear. The pH was adjusted to ca. 9–10 by the addition of saturated sodium hydrogen carbonate solution (vigorous evolution of gas!). The product was extracted with dichloromethane ($3 \times 50 \text{ mL}$). The combined organic extracts were dried with magnesium sulfate, and the solvent was evaporated in vacuo. The solid residue was triturated with a mixture of hexane (20 mL) and acetone (1 mL) to give pure amine 28a or 28b as a pale yellow solid.

7-Aminoflavone (28b): Yield 68%; m.p. 183–185 °C, lit. m.p.^[5] 186 °C. ¹H NMR (CDCl₃): δ = 8.01 (d, *J* = 9.2 Hz, 1 H, 5-H), 7.87 (m, 2 H, 2'-H, 6'-H), 7.50 (m, 3 H, 3'-H, 5'-H, 4'-H), 6.69 (m, 3 H, 3-H, 6-H, 8-H), 4.33 (br. s, 2 H, NH₂) ppm. ¹³C NMR (CDCl₃): δ = 177.7 (C-4), 162.4 (C-2), 158.3 (C-7), 151.9 (C-8a), 132.0 (C-1'), 131.2 (C-5), 128.9 (C-3', C-5'), 127.2 (C-4'), 126.1 (C-2', C-6'), 115.7 (C-4a), 113.7 (C-6), 107.3 (C-3), 100.2 (C-8) ppm. IR (KBr): \tilde{v} = 3462, 3332, 3194, 1633, 1586, 1569, 1450, 1388, 1370, 1248, 1186, 1026, 909, 832, 768 cm⁻¹. MS: *m/z* = 237 [M]⁺⁻, 209, 180, 152, 135, 107, 79. C₁₅H₁₁NO₂ (237.26): calcd. C 75.94, H 4.67, N 5.90; found C 75.95, H 4.69, N 5.95.

General Method for Coupling 6- and 7-Bromoflavone with Amino Acid Methyl Ester Hydrochlorides: Bromoflavone 11 or 17 (0.66 mmol), cesium carbonate (557 mg, 1.71 mmol), BINAP (32 mg, 0.050 mmol), and the amino acid methyl ester hydrochloride (0.80 mmol) were mixed in dry xylene (6 mL) in an oven-dried flask under nitrogen, and Pd(OAc)₂ (7 mg, 0.032 mmol) was added. The reaction mixture was stirred at reflux at 135 °C for 24 h in an oil bath. The crude reaction mixture was applied directly to a silica gel column to give the pure cross-coupled product.



7-{[2-Phenyl-1-(methoxycarbonyl)ethyl]amino}flavone (31a): Eluent: toluene/ethyl acetate, 2:1; yield 168 mg (64%); m.p. 171.5-173.5 °C. ¹H NMR (CDCl₃): δ = 8.00 (d, J = 8.6 Hz, 1 H, 5-H), 7.88 (m, 2 H, 2'-H, 6'-H), 7.51 (m, 3 H, 3'-H, 5-H, 4'-H), 7.31 (m, 3 H, 3"'-H, 5'''-H, 4'''-H), 7.15 (m, 2 H, 2'''-H, 6'''-H), 6.71 (s, 1 H, 3-H), 6.64 (dd, J = 1.8, 8.6 Hz, 1 H, 6-H), 6.54 (s, 1 H, 8-H), 4.83 (d, *J* = 7.9 Hz, 1 H, NH), 4.50 (m, 1 H, 1^{''}-H), 3.76 (s, 3-H, OCH₃), 3.23 (m, 2 H, 2"-H) ppm. ¹³C NMR (CDCl₃): δ = 178.2 (C-4), 172.2 (COOMe), 163.2 (C-2), 158.8 (C-8a), 151.4 (C-7), 135.4 (C-1'''), 131.8 (C-1'), 131.5 (C-4'), 129.2 (C-3''', C-5'''), 129.0 (C-2''', C-6'''), 128.8 (C-3', C-5'), 127.4 (C-5), 127.2 (C-4'''), 126.2 (C-2', C-6'), 114.9 (C-4a), 113.6 (C-6), 106.8 (C-3), 97.6 (C-8), 56.8 (C-1''), 52.6 (OCH₃), 38.1 (C-2'') ppm. IR (KBr): $\tilde{v} = 3396$, 3265, 3152, 3065, 3028, 2948, 1726, 1624, 1593, 1538, 1507, 1448, 1436, 1370, 1249, 1191, 1148, 906, 822, 766, 699 cm⁻¹. MS: m/z = 399 $[M]^{+}$, 340, 308 (100%), 248, 221, 165, 91. $C_{25}H_{21}NO_4$ (399.45): calcd. C 75.17, H 5.30, N 3.51; found C 75.21, H 5.33, N 3.54.

This product was also obtained as a by-product from the reaction of 6-bromoflavone (11) in 6% yield.

7-{[3-Methyl-1-(methoxycarbonyl)butyl]amino}flavone (31c): Eluent: toluene/ethyl acetate, 2:1; yield 136 mg (56%); m.p. 152-156 °C. ¹H NMR (CDCl₃): δ = 8.01 (d, J = 8.6 Hz, 1 H, 5-H), 7.89 (m, 2 H, 2'-H, 6'-H), 7.50 (m, 3 H, 3'-H, 5'-H, 4'-H), 6.71 (s, 1 H, 3-H), 6.68 (dd, J = 2.1, 8.6 Hz, 1 H, 6-H), 6.56 (d, J = 2.1 Hz, 1 H, 8-H), 4.80 (d, J = 8.6 Hz, 1 H, NH), 4.21 (m, 1 H, 1''-H), 3.77 (s, 3 H, OCH₃), 1.78 (m, 3 H, 2^{''}-H, 3^{''}-H), 1.02 (d, *J* = 6.5 Hz, 3 H, 4''-H), 0.97 (d, J = 6.1 Hz, 3 H, 4''-H) ppm. ¹³C NMR (CDCl₃): δ = 177.7 (C-4), 173.9 (COOMe), 162.4 (C-2), 158.5 (C-8a), 151.8 (C-7), 132.0 (C-1'), 131.1 (C-4'), 128.9 (C-3', C-5'), 127.0 (C-5), 126.1 (C-2', C-6'), 115.5 (C-4a), 113.1 (C-6), 107.4 (C-3), 97.5 (C-8), 54.6 (OCH₃), 52.4 (C-1''), 41.8 (C-2''), 24.9, 22.7 (C-4''), 22.1 (C-3'') ppm. IR (KBr): $\tilde{v} = 3289, 3153, 3087, 2953, 1724, 1620,$ 1589, 1566, 1534, 1449, 1373, 1250, 1190, 1149, 1000, 9008, 824, 768, 686, 673 cm⁻¹. MS: $m/z = 365 [M]^{+1}$, 306 (100%), 250, 91. C₂₂H₂₃NO₄ (365.43): calcd. C 72.31, H 6.34, N 3.83; found C 72.35, H 6.38, N 3.85.

Supporting Information (see footnote on the first page of this article): Experimental procedures, physical and spectroscopic data (¹H and ¹³C NMR, IR, and MS) for the products.

Acknowledgments

Financial support from the Hungarian Scientific Research Fund (OTKA) (grant number K75806) and from the New Hungary Development Plan, cofinanced by the European Social Fund and the European Regional Development Fund (project number TÁMOP-4.2.2/B-10/1-2010-0024) is greatly appreciated.

- [1] S. Khadem, R. J. Marles, Molecules 2012, 17, 191–206.
- [2] a) R. G. Naik, S. L. Kattige, S. V. Bhat, B. Alreja, N. J. de Souza, R. H. Rupp, *Tetrahedron* **1988**, *44*, 2081–2086; b) S. Khadem, R. J. Marles, *Molecules* **2012**, *17*, 191–206.
- [3] a) K. S. Kim, J. S. Sack, J. S. Tokarski, L. Qian, L. Leith, Y. F. Kelly, R. N. Misra, J. T. Hunt, S. D. Kimball, W. G. Hum-

Buchwald–Hartwig Reactions of Monohaloflavones

phreys, B. S. Wautlet, J. G. Mulheron, K. R. Webster, J. Med. Chem. 2000, 43, 4126-4134; b) H. Lu, D. J. Chang, B. Baratte, L. Meijer, U. Schulze-Gahmen, J. Med. Chem. 2005, 48, 737-743; c) A. Montagnoli, B. Valsasina, V. Croci, M. Meninchinchieri, S. Rainoldi, V. Marchesi, M. Tibolla, P. Tenca, D. Brotherton, C. Albanese, V. Patton, R. Alzani, A. Ciavolella, F. Sola, A. Molinari, D. Volpi, N. Avanzi, F. Fiorentini, M. Cattoni, S. Healy, D. Ballinari, E. Pesenti, A. Isacchi, J. Moll, A. Bensimon, E. Vanotti, C. Santocanale, Nat. Chem. Biol. 2008, 4, 357-365; d) K. S. Kim, S. D. Kimball, R. N. Misra, D. B. Rawlins, J. T. Hunt, H. Y. Xiao, S. Lu, L. Qian, W. C. Han, W. Shan, T. Mitt, Z. W. Cai, M. A. Poss, H. Zhu, J. S. Sack, J. S. Tokarski, C. Y. Chang, N. Pavletich, A. Kamath, W. G. Humphreys, P. Marathe, I. Bursuker, K. A. Kellar, U. Roongta, R. Batorsky, J. G. Mulheron, D. Bol, C. R. Fairchilds, F. Y. Lee, K. R. Webster, J. Med. Chem. 2002, 45, 3905-3927; e) X. Lü, W. E. Burgan, M. A. Cerra, E. Y. Chuang, M.-H. Tsai, P. J. Tofilon, K. Camphausen, Mol. Cancer Ther. 2004, 3, 861-872; f) D. Rathkopf, M. A. Dickson, R. D. Carvajal, M. A. Shah, N. Wu, R. Lefkowitz, M. Gonen, L. M. Cane, H. J. Dials, J. L. Winkelmann, G. J. Bosi, G. K. Schwartz, Clin. Cancer Res. 2009, 15, 7405-7411; g) K. C. Bible, P. P. Peethambaram, A. L. Oberg, W. Maples, D. L. Groteluschen, M. Boente, J. K. Burton, L. C. Gomez-Dahl, J. D. Tibodeau, C. R. Isham, J. L. Maguire, V. Shridhar, A. K. Kukla, K. J. Voll, M. J. Mauer, A. D. Colevas, J. Wright, L. A. Doyle, C. Erlichman, Gynecol. Oncol. 2012, 127, 55-62.

- [4] D. S. Bapat, K. Venkataraman, Proc. Indian Acad. Sci. 1955, 42A, 336–341.
- [5] a) M. Vijaya Lakshmi, N. V. Subba Rao, *Curr. Sci.* 1967, *36*, 401–402; b) M. Vijaya Lakshmi, N. V. Subba Rao, *Ind. J. Chem.* 1972, *10*, 34–37.
- [6] H. Gao, J. Kawabata, Bioorg. Med. Chem. 2005, 13, 1661-1671.
- [7] a) M. Cushman, D. Nagarathnam, D. L. Burg, R. L. Geahlen, J. Med. Chem. 1991, 34, 798–806; b) M. Cushman, H. Zhu, R. L. Geahlen, A. J. Kraker, J. Med. Chem. 1994, 37, 3353– 3362.
- [8] a) T. Akama, Y. Shida, T. Sugaya, H. Ishida, K. Gomi, M. Kasai, J. Med. Chem. 1996, 39, 3461–3469; b) Y. Shida, T. Sugaya, K. Gomi, M. Kasai, M. Moritomo, JPH 02256673, 1990; c) T. Akama, H. Ishida, Y. Shida, U. Kimura, K. Gomi, H. Saito, E. Fuse, S. Kobayashi, N. Yoda, M. Kasai, J. Med. Chem. 1997, 40, 1894–1900.
- [9] D. A. Vasselin, A. D. Westwell, C. S. Matthews, J. Med. Chem. 2006, 49, 3973–3981.
- [10] a) http://dctd.cancer.gov/FeaturedAgents/710464AminoflavoneProdrug.htm; b) K. M. Snader, B. R. Vishnuvajjala, T. Akama, WO 2002081462, 2002; c) A. M. Burger, B. Nguyen, WO 2010008731, 2010.
- [11] a) M. P. Goetz, J. M. Reid, Y. Qi, A. Chen, R. M. McGovern, M. J. Kuffel, P. D. Scanlon, C. Erlichman, M. M. Ames, *J. Clin.* Oncol. 2011, 29, 2546 (Suppl.); b) P. P. Nandekar, A. B. Sangamwar, *Expert Opin. Drug Discov.* 2012, 7, 771–789.
- [12] D. Dauzonne, B. Folléas, L. Martinez, G. G. Chabot, *Eur. J. Med. Chem.* 1997, 32, 71–82.
- [13] a) T. Sawada, S. Hashimoto, S. Tohma, Y. Nishioka, T. Nagai, T. Sato, K. Ito, T. Inoue, M. Iwata, K. Yamamoto, *Immuno-pharmacology* 2000, 49, 285–294; b) T. Inaba, K. Tanaka, R. Takeno, H. Nagaki, C. Yoshida, S. Takano, *Chem. Pharm. Bull.* 2000, 48, 131–139; c) F. Du, L.-J. Lü, Q. Fu, M. Dai, J.-L. Teng, W. Fan, S.-L. Chen, P. Ye, N. Shen, X.-F. Huang, J. Qian, C.-D. Bao, *Arthritis Res. Ther.* 2008, 10, R136; d) K. Tanaka, T. Yamamoto, Y. Aikawa, K. Kizawa, K. Muramoto, H. Matsuno, A. Muragami, *Rheumatology* 2003, 42, 1365–1371.
- [14] J. F. Geissler, J. L. Roesel, T. Meyer, U. P. Trinks, P. Traxler, N. B. Lydon, *Cancer Res.* **1992**, *52*, 4492–4498.
- [15] M. K. Rastogi, K. Chaudhary, R. P. Kapoor, C. P. Garg, *Indian J. Chem. B* 1978, *16*, 895–897.



- [16] Y.-M. Cheng, S.-C. Pu, Y.-C. Yu, P. T. Chou, C.-H. Huang, C.-T. Chen, T.-H. Li, W.-P. Hu, J. Phys. Chem. A 2005, 109, 11696–11706.
- [17] N. N. Agrawal, P. A. Soni, Indian J. Chem. 2005, 44B, 2601– 2603.
- [18] F. Eiden, G. Patzelt, H. Buchborn, Arch. Pharm. 1989, 322, 589–592.
- [19] M. Lorenz, M. S. Kabir, J. M. Cook, *Tetrahedron Lett.* 2010, 51, 1095–1098.
- [20] A. A. Raval, N. M. Shah, J. Org. Chem. 1956, 21, 1408-1411.
- [21] L. Reichel, G. Hampel, Justus Liebigs Ann. Chem. 1966, 693, 216–224.
- [22] G. Wurm, U. Geres, Arch. Pharm. 1978, 311, 177-183.
- [23] L. Tang, S. Zhang, J. Yang, W. Gao, J. Cui, T. Zhuang, *Molecules* 2004, *9*, 842–848.
 [24] C. Wurm, H. Cargo, Auch. Phys. 1077, 210, 600, 614.
- [24] G. Wurm, U. Geres, Arch. Pharm. 1977, 310, 609–614.
- [25] M. Cardenas, M. Marder, V. C. Blank, L. P. Roguin, *Bioorg. Med. Chem.* 2006, 14, 2966–2971.
- [26] G. Wurm, U. Geres, Arch. Pharm. 1976, 309, 273-278.
- [27] K. Görlitzer, Arch. Pharm. 1979, 312, 248-253.
- [28] R. Larget, B. Lockhart, P. Renard, M. Largeron, *Bioorg. Med. Chem. Lett.* 2000, 10, 835–838.
- [29] J. Quintin, C. Ruillier, S. Thoret, G. Lewin, *Tetrahedron* 2006, 62, 4038–4051.
- [30] X. Zheng, W.-D. Meng, Y.-Y. Xu, J.-G. Cao, F.-L. Quing, Bioorg. Med. Chem. Lett. 2003, 13, 881–884.
- [31] M. Ono, N. Yoshida, K. Ishibashi, M. Haratake, M. Arano, H. Mori, M. Nakayama, J. Med. Chem. 2005, 48, 7253–7260.
- [32] J. Lee, T. Park, S. Jeong, K.-H. Kim, C. Hong, *Bioorg. Med. Chem. Lett.* 2007, 17, 1284–1287.
- [33] C. Lecoutey, C. Fossey, L. Demuynck, F. Lefoulon, F. Fabis, S. Rault, *Tetrahedron* 2008, 64, 11243–11248.
- [34] B. Bauvois, M.-L. Puiffe, J.-B. Bongui, S. Paillat, C. Monneret, D. Dauzonne, J. Med. Chem. 2003, 46, 3900–3913.
- [35] D. Dauzonne, C. Grandjean, Synthesis 1992, 677-680.
- [36] M. Michalska, Chem. Ind. 1966, 628-629.
- [37] C. Paparao, K. V. Rao, V. Sundaramurthy, Synthesis 1981, 236– 237.
- [38] T. Patonay, K. Kónya, E. Juhász-Tóth, Chem. Soc. Rev. 2011, 40, 2797–2847.
- [39] a) T. Patonay, R. Bognár, Gy. Litkei, *Tetrahedron* 1984, 40, 2555–2562; b) T. Patonay, J. Jekő, É. Rimán, *Synth. Commun.* 2002, 32, 2403–2415; c) N. De Meyer, A. Haemers, L. Mishra, H.-K. Pandey, L. A. C. Pieters, D. A. Vanden Bergen, A. J. Vlietinck, *J. Med. Chem.* 1991, 34, 736–746.
- [40] T. Patonay, M. Rákosi, Gy. Litkei, R. Bognár, *Liebigs Ann. Chem.* 1979, 161–173.
- [41] D. Dauzonne, L. Martinez, *Tetrahedron Lett.* 1995, 36, 1845– 1848.
- [42] D. T. Patoilo, A. M. S. Silva, J. A. S. Cavaleiro, Synlett 2010, 1381–1385.
- [43] B. Zurowska, A. Erxleben, L. Glinka, M. Lecycka, E. Zyner, J. Ochoki, *Inorg. Chim. Acta* 2009, 362, 739–744.
- [44] a) R. Bognár, C. O'Brien, E. M. Philbin, S. Ushioda, T. S. Wheeler, *Chem. Ind.* **1960**, 1186; b) C. O'Brien, E. M. Philbin, S. Ushioda, T. S. Wheeler, *Tetrahedron* **1963**, *19*, 373–377.
- [45] a) A. Takechi, H. Takikawa, H. Miyake, M. Sasaki, *Chem. Lett.* **2006**, *35*, 128–129; b) H. Miyake, S. Nishino, A. Nishimura, M. Sasaki, *Chem. Lett.* **2007**, *36*, 522–523.
- [46] a) S. Guram, R. A. Rennels, S. L. Buchwald, Angew. Chem. Int. Ed. Engl. 1995, 34, 1348–1350; Angew. Chem. 1995, 107, 1456– 1459; b) J. Louie, J. F. Hartwig, Tetrahedron Lett. 1995, 36, 3609–3612; c) M. Kienle, S. R. Dubbakka, K. Brade, P. Knochel, Eur. J. Org. Chem. 2007, 4166–4176, and the references cited therein; d) A. de Meijere, F. Diederich, Metal-Catalyzed Cross-Coupling Reactions, chapter 13, pp. 699–760, Wiley-VCH, Weinheim, Germany, 2004.
- [47] B.-L. Deng, J. A. Lepoivre, G. Lemiere, Eur. J. Org. Chem. 1999, 2683–2688.

FULL PAPER

- [48] R. J. Fitzmaurice, Z. C. Etheridge, E. Jumel, D. N. Woolfson, S. Caddick, *Chem. Commun.* 2006, 4814–4816.
- [49] T. Patonay, A. Vasas, A. Kiss-Szikszai, A. M. S. Silva, J. A. S. Cavaleiro, Aust. J. Chem. 2010, 63, 1582–1593.
- [50] Y. Ju, R. S. Varma, Org. Lett. 2005, 7, 2409-2411.
- [51] J. P. Wolfe, J. Ahman, J. P. Sadighi, R. A. Singer, S. L. Buchwald, *Tetrahedron Lett.* **1997**, *38*, 6367–6370.
- [52] a) S. Lee, M. Jorgensen, J. F. Hartwig, Org. Lett. 2001, 3, 2729–2732; b) X. Huang, S. L. Buchwald, Org. Lett. 2001, 3, 3417–3419.
- [53] M. L. Lee, K. H. Kang, Y. M. Lee, D. Kim, Y. S. Park, Bull. Korean Chem. Soc. 2008, 29, 1075–1078.
- [54] Y. M. Ahn, L. Vogeti, C.-J. Liu, H. K. R. Santhapuram, J. M. White, V. Vasandani, L. A. Mitscher, G. H. Lushington, P. R.

Hanson, D. R. Powell, R. H. Himes, K. F. Roby, Q. Ye, G. I. Georg, *Bioorg. Med. Chem.* 2007, *15*, 702–713.

- [55] Among others: a) M. Schnürch, M. Spina, A. F. Kahn, M. D. Mihovilovic, P. Stanetty, *Chem. Soc. Rev.* 2007, *36*, 1046–1057;
 b) N. de Souza, M. Vinicius, *Curr. Org. Chem.* 2007, *11*, 637–646; c) X.-F. Duan, Z.-B. Zhang, *Heterocycles* 2005, *65*, 2005–2012; d) J. F. Bunnett, *Acc. Chem. Res.* 1972, *5*, 139–147.
- [56] Sz. Fekete, T. Patonay, A. M. S. Silva, J. A. S. Cavaleiro, ARKI-VOC 2012, 5, 210–225.
- [57] M. Brenner, W. Huber, Helv. Chim. Acta 1953, 36, 1109–1115. Received: August 19, 2014 Published Online: ■

Buchwald-Hartwig Reactions of Monohaloflavones



Amination of Flavones



K. Kónya, D. Pajtás, A. Kiss-Szikszai, T. Patonay^{*} 1–13

Buchwald-Hartwig Reactions of Monohaloflavones

Keywords: Homogeneous catalysis / Crosscoupling / Amination / Palladium / Amino acids / Oxygen heterocycles

A new and efficient approach to (substituted) aminoflavones and hybrids of flavones and amino acids using Buchwald– X = H, Y = alkyl/aryl amino Hartwig coupling was developed and optimised.